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(54) Title: LACTOFERRIN RECEPTOR GENES OF MORAXELLA (57) Abstract <p>Purified and isolated nucleic acid molecules are provided which encode lactoferrin receptor proteins of <i>Moraxella</i>, such as <i>M. catarrhalis</i>, or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid sequence may be used to produce recombinant lactoferrin receptor proteins Lbp1, Lbp2 or ORF3 of the strain of <i>Moraxella</i> free of other proteins of the <i>Moraxella</i> strain for purposes of diagnostics and medical treatment. Furthermore, the nucleic acid molecule may be used in the diagnosis of infection.</p>		

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TITLE OF INVENTIONLACTOFERRIN RECEPTOR GENES OF MORAXELLAFIELD OF INVENTION

5 The present invention relates to the molecular cloning of genes encoding lactoferrin receptor (LfR) proteins and, in particular, to the cloning of lactoferrin binding protein genes (*lbp* genes) from *Moraxella* (*Branhamella*) *catarrhalis*.

10 REFERENCE TO RELATED APPLICATION

This application is a continuation-in-part of copending United States patent application No. 08/867,941 filed June 3, 1997.

BACKGROUND OF THE INVENTION

15 *Moraxella* (*Branhamella*) *catarrhalis* bacteria are Gram-negative diplococcal pathogens which are carried asymptotically in the healthy human respiratory tract. However, in recent years, *M. catarrhalis* has been recognized as an important causative agent of otitis
20 media. In addition, *M. catarrhalis* has been associated with sinusitis, conjunctivitis, and urogenital infections, as well as with a number of inflammatory diseases of the lower respiratory tract in children and adults, including pneumonia, chronic bronchitis,
25 tracheitis, and emphysema (refs. 1 to 8). (Throughout this application, various references are cited in parentheses to describe more fully the state of the art to which this invention pertains. Full bibliographic information for each citation is found at the end of the
30 specification, immediately preceding the claims. The disclosures of these references are hereby incorporated by reference into the present disclosure). Occasionally, *M. catarrhalis* invades to cause

septicaemia, arthritis, endocarditis, and meningitis (refs. 9 to 13).

5 *M. catarrhalis* colonizes the human upper respiratory tract and is an important cause of otitis media in infants and children as well as lower respiratory tract infections in adults with chronic obstructive pulmonary disease.

10 Otitis media is one of the most common illnesses of early childhood and approximately 80% of all children suffer at least one middle ear infection before the age of three (ref. 14). Chronic otitis media has been associated with auditory and speech impairment in children, and in some cases, has been associated with learning disabilities. Conventional treatments for
15 otitis media include antibiotic administration and surgical procedures, including tonsillectomies, adenoidectomies, and tympanocentesis. In the United States, treatment costs for otitis media are estimated to be between one and two billion dollars per year.

20 In otitis media cases, *M. catarrhalis* is commonly co-isolated from middle ear fluid along with *Streptococcus pneumoniae* and non-typable *Haemophilus influenzae*, which are believed to be responsible for 50% and 30% of otitis media infections, respectively. *M.*
25 *catarrhalis* is believed to be responsible for approximately 20% of otitis media infections (ref. 15). Epidemiological reports indicate that the number of cases of otitis media attributable to *M. catarrhalis* is increasing, along with the number of antibiotic-resistant isolates of *M. catarrhalis*. Thus, prior to
30 1970, no β -lactamase-producing *M. catarrhalis* isolates had been reported, but since the mid-seventies, an increasing number of β -lactamase-expressing isolates have been detected. Recent surveys suggest that up to

80 to 85% of clinical isolates produce β -lactamase (ref. 16, 22, 23).

Iron-restriction is a general host defence mechanism against microbial pathogens. A number of bacterial species including *Neisseria meningitidis* (ref. 17, 24), *N. gonorrhoeae* (ref. 25) and *M. catarrhalis* (ref. 17), express outer membrane proteins which specifically bind human lactoferrin.

M. catarrhalis infection may lead to serious disease. It would be advantageous to provide a recombinant source of lactoferrin binding proteins as antigens in immunogenic preparations including vaccines, carriers for other antigens and immunogens and the generation of diagnostic reagents. The genes encoding lactoferrin binding proteins and fragments thereof are particularly desirable and useful in the specific identification and diagnosis of *Moraxella* and for immunization against disease caused by *M. catarrhalis* and for the generation of diagnostic reagents.

SUMMARY OF THE INVENTION

The present invention is directed towards the provision of purified and isolated nucleic acid molecules encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid molecules and isolated and purified lactoferrin binding proteins provided herein are useful for the specific detection of strains of *Moraxella* and for diagnosis of infection by *Moraxella*. The purified and isolated nucleic acid molecules provided herein, such as DNA, are also useful for expressing the *lbp* genes by recombinant DNA means for providing, in an economical manner, purified and isolated lactoferrin receptor

proteins free of other *Moraxella* proteins, as well as subunits, fragments or analogs thereof.

The lactoferrin receptor, subunits or fragments thereof or analogs thereof, as well as nucleic acid molecules encoding the same and vectors containing such
5 nucleic acid molecules, are useful in immunogenic compositions for vaccinating against diseases caused by *Moraxella*, the diagnosis of infection by *Moraxella*, and as tools for the generation of immunological reagents.

10 Monoclonal antibodies or mono-specific antisera (antibodies) raised against the lactoferrin receptor protein produced in accordance with aspects of the present invention are useful for the diagnosis of infection by *Moraxella*, the specific detection of
15 *Moraxella* (in, for example, *in vitro* and *in vivo* assays) and for the treatment of diseases caused by *Moraxella*.

In accordance with one aspect of the present invention, there is provided a purified and isolated
20 nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella*, more particularly a strain of *M. catarrhalis*, specifically *M. catarrhalis* strain 4223, Q8 or VH19 or a fragment or an analog of the lactoferrin receptor protein. A fragment of the
25 lactoferrin receptor protein is a portion of the protein which retains the immunological properties of the protein.

In one preferred embodiment of the invention, the nucleic acid molecule may encode only the Lbp1 protein
30 of the *Moraxella* strain or only the Lbp2 protein of the *Moraxella* strain or only the ORF3 protein of the *Moraxella* strain. In another preferred embodiment of the invention, the nucleic acid may encode a fragment

of the lactoferrin receptor protein of a strain of *Moraxella* having a conserved amino acid sequence.

In a further aspect of the present invention, there is provided an isolated and purified nucleic acid molecule encoding at least one lactoferrin binding protein of *Moraxella* having a restriction map as shown in Figure 3 for *M. catarrhalis* 4223, Figure 5 for *M. catarrhalis* Q8 or Figure 17 for *M. catarrhalis* VH19 or the equivalent map from other strains of *Moraxella*.

In another aspect of the present invention, there is provided a purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of (a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto; (b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and (c) a DNA sequence encoding a functional lactoferrin receptor preprotein of *Moraxella*, which may be a DNA sequence which hybridizes under stringent conditions to any one of the DNA sequences defined in (a) or (b). The DNA sequence defined in (c) may have at least about 90% sequence identity with any one of the DNA sequences defined in (a) or (b). Stringent conditions of hybridization are described below. Sequence identity is determined in the manner described below.

In an additional aspect, the present invention includes a vector adapted for transformation of a host, comprising a nucleic acid molecule as provided herein and may have the characteristics of a nucleotide sequence contained within vectors pLD3, pLDW3, pLD1-8 and pLDW1.

The vector may be adapted for expression of the encoded lactoferrin receptor protein, fragments or

analogs thereof, in a heterologous or homologous host, in either a lipidated or non-lipidated form. Accordingly, a further aspect of the present invention provides an expression vector adapted for transformation of a host comprising a nucleic acid molecule as provided herein and expression means operatively coupled to the nucleic acid molecule for expression by the host of the lactoferrin receptor protein or the fragment or analog of the lactoferrin receptor protein.

In specific embodiments of this aspect of the invention, the nucleic acid molecule may encode substantially all the lactoferrin receptor protein, only the Lbp1 protein of the *Moraxella* strain, only the Lbp2 protein of the *Moraxella* strain, only the ORF3 protein of the *Moraxella* strain, or fragments of the Lbp1, Lbp2 or ORF3 proteins.

The expression means may include a nucleic acid portion encoding a leader sequence for secretion from the host of the lactoferrin receptor protein or the fragment or the analog of the lactoferrin receptor protein. The expression means also may include a nucleic acid portion encoding a lipidation signal for expression from the host of a lipidated form of the lactoferrin receptor protein or the fragment or the analog of the lactoferrin receptor protein. The host may be selected from, for example, *Escherichia coli*, *Bacillus*, *Bordetella*, *Haemophilus*, *Moraxella*, fungi, yeast or baculovirus and Semliki Forest virus expression system may be used. In a particular embodiment, the plasmid adapted for expression of Lbp2 is pRD2A, pRD2B, pQW2A or pQW2B; the plasmid adapted for expression of Lbp1 is pRD1A, pRD1B, PQ1A or pQ1B; and the plasmid adapted for expression of ORF3 is pLRD3 or pLQW3.

In an additional aspect of the invention, there is provided a transformed host containing an expression vector as provided herein. The invention further includes a recombinant lactoferrin receptor protein or
5 fragment or analog thereof of a strain of *Moraxella* producible by the transformed host.

Such recombinant lactoferrin receptor protein may be provided in substantially pure form according to a further aspect of the invention, which provides a method
10 of forming a substantially pure recombinant lactoferrin receptor protein, which comprises growing the transformed host provided herein and isolating and purifying the lactoferrin receptor protein, analog or fragment thereof. The lactoferrin receptor protein may
15 be expressed in inclusion bodies, which may be purified free from cellular material and soluble proteins and lactoferrin receptor protein solubilized from the purified inclusion bodies, and the lactoferrin receptor protein purified free from other solubilized materials.
20 The substantially pure recombinant lactoferrin receptor protein may comprise Lbp1 alone, Lbp2 alone, ORF3 or a mixture of two or more of such proteins. The recombinant protein is generally at least about 70% pure, preferably at least about 90% pure.

25 Further aspects of the present invention, therefore, provide recombinantly-produced Lbp1 protein (or a fragment or analog thereof) of a strain of *Moraxella* devoid of the Lbp2 and ORF3 proteins of the *Moraxella* strain and any other protein of the *Moraxella*
30 strain, recombinantly-produced Lbp2 protein (or a fragment or analog thereof) of a strain of *Moraxella* devoid of the Lbp1 and ORF3 proteins of the *Moraxella* strain and any other protein of the *Moraxella* strain, and recombinantly-produced ORF3 protein (or a fragment
35 or analog thereof) of a strain of *Moraxella* devoid of

the Lbp1 and Lbp2 proteins of the *Moraxella* strain and any other protein of the *Moraxella* strain. The *Moraxella* strain may be *M. catarrhalis* 4223, Q8 or VH19 strain.

5 The invention further includes, in an additional aspect, an open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the lactoferrin binding protein which is encoded by region downstream from the genes encoding Lbp2 and Lbp1
10 proteins of the *Moraxella* strain. The ORF3 may be from a strain of *M. catarrhalis*, which may be strain 4223 or Q8. The Lbp3 may have a molecular mass of about 60 kDa.

 In accordance with another aspect of the
15 invention, an immunogenic composition is provided which comprises at least one active component selected from at least one nucleic acid molecule as provided herein, at least one recombinant protein as provided herein or at least one novel protein as provided herein, and a
20 pharmaceutically acceptable carrier therefor or vector therefor. The at least one active component produces an immune response when administered to a host.

 The immunogenic compositions provided herein may be formulated as a vaccine for *in vivo* administration
25 to a host to provide protection against disease caused by *M. catarrhalis*. For such purpose, the compositions may be formulated as a microparticle, capsule, ISCOM or liposome preparation. The immunogenic composition may be provided in combination with a targeting molecule
30 for delivery to specific cells of the immune system or to mucosal surfaces. The immunogenic compositions of the invention (including vaccines) may further comprise at least one other immunogenic or immunostimulating material and the immunostimulating material may be at
35 least one adjuvant or at least one cytokine.

Suitable adjuvants for use in the present invention include (but are not limited to) aluminum phosphate, aluminum hydroxide, QS21, Quil A, derivatives and components thereof, ISCOM matrix, calcium phosphate, calcium hydroxide, zinc hydroxide, a glycolipid analog, an octadecyl ester of an amino acid, a muramyl dipeptide, polyphosphazene, ISCOPREP, DC-chol, DDBA and a lipoprotein and other adjuvants to induce a TH1 response. Advantageous combination of adjuvants are described in copending United States Patent Applications No. 08/261,194 filed June 16, 1994 and 08/483,856 filed June 7, 1995, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference (WO 95/34308, published November 21, 1995).

In accordance with another aspect of the invention, there is provided a method for generating an immune response in a host, comprising the step of administering to a susceptible host, such as a human, an effective amount of the immunogenic composition as recited above. The immune response may be humoral or a cell-mediated immune response and may provide protection against disease caused by *Moraxella*. Hosts in which protection against disease may be conferred include primates, including humans.

In a further aspect, there is provided a live vector for delivery of lactoferrin receptor to a host, comprising a vector containing the nucleic acid molecule as described above. The vector may be selected from *Salmonella*, *Mycobacterium bovis*, BCG, adenovirus, poxvirus, vaccinia and poliovirus.

The nucleic acid molecules provided herein are useful in diagnostic applications. Accordingly, in a further aspect of the invention, there is provided a method of determining the presence, in a sample, of

nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising the steps of:

5 a) contacting the sample with a nucleic acid molecule as provided herein to produce duplexes comprising the nucleic acid molecule and any nucleic acid molecule encoding the lactoferrin receptor protein of a strain of *Moraxella* present in the sample and specifically hybridizable therewith; and

b) determining the production of the duplexes.

10 In addition, the present invention provides a diagnostic kit for determining the presence, in a sample, of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising:

a) a nucleic acid molecule as provided herein;

15 b) means for contacting the nucleic acid molecule with the sample to produce duplexes comprising the nucleic acid molecule and any such nucleic acid present in the sample and hybridizable with the nucleic acid present in the sample and hybridizable with the nucleic acid molecule; and

20 c) means for determining production of the duplexes.

The invention further includes the use of the nucleic acid molecules and proteins provided herein as medicines. The invention additionally includes the use of the nucleic acid molecules and proteins provided herein in the manufacture of medicaments for protection against disease caused by strains of *Moraxella*.

Advantages of the present invention include:

- 30 - an isolated and purified nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein;
- recombinantly-produced lactoferrin receptor proteins, including Lbp1, Lbp2 and ORF3 and fragments
- 35

and analogs thereof free from each other and other *Moraxella* proteins;

- open reading frame protein 3; and
 - diagnostic kits and immunological reagents for
- specific identification of *Moraxella*.

BRIEF DESCRIPTION OF DRAWINGS

The present invention will be further understood from the following description with reference to the drawings, in which:

Figure 1 shows partial sequence of the 2.2 kb PCR amplified fragments of the *lbpA* genes from *M. catarrhalis* 4223 or Q8, which were used to probe the phage libraries. In the figure, Tbp1 is the deduced 4223 Tbp1 sequence (as described in United States Patent Application No. 08/613,009 filed March 8, 1996, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference) (SEQ ID No: 19), Lbp1 is the deduced full-length 4223 Lbp1 sequence (SEQ ID No: 3) used here solely for aligning the PCR fragments, PCR4 is the 4223 PCR fragment (SEQ ID No: 20), and PCR5 is a partial sequence of the Q8 PCR fragment (SEQ ID No: 21). Only single strand sequence was obtained for the PCR fragments and "X" has been inserted where there was a doubtful sequence. Dashes have been used for maximum alignment. The underlined sequence in Lbp1 (MVQYTYRKGKKNKAH - SEQ ID No: 22) represents the position of a CNBr peptide used to generate the 5'-PCR primer.

Figure 2 shows the nucleotide (SEQ ID No: 1, full sequence; SEQ ID No: 2, Lbp2 coding sequence; SEQ ID No: 3, Lbp1 coding sequence, first methionine; SEQ ID No: 4, Lbp1 coding sequence, second methionine; SEQ ID No: 5, ORF3 coding sequence) and deduced amino acid sequences (SEQ ID No: 11, Lbp2; SEQ ID No: 12, Lbp1, first methionine; SEQ ID No: 13, Lbp1, second

methionine; SEQ ID No: 14, ORF3) of the putative *lfr* locus from *M. catarrhalis* 4223. There are three tandem genes in the putative *lfr* locus identified as *lbpB*, *lbpA* and *orf3*. Potential promoter elements found upstream of the *lbpB* and *lbpA* genes are indicated by underlining.

Figure 3 shows a restriction map of clone pLD1-8 containing the *lbpA*, *lbpB*, and *orf3* genes from *M. catarrhalis* isolate 4223.

Figure 4 shows the nucleotide (SEQ ID No: 6, full sequence; SEQ ID No: 7, *Lbp2* coding sequence; SEQ ID No: 8, *Lbp1* coding sequence, first methionine; SEQ ID No: 9, *Lbp2*, second methionine; SEQ ID No: 10, ORF3 coding sequence) and deduced amino acid sequences (SEQ ID No: 15, *Lbp2*; SEQ ID No: 16, *Lbp1*, first methionine; SEQ ID No: 17, *Lbp1*, second methionine; SEQ ID No: 18, *Lbp3*) of the putative *lfr* locus from *M. catarrhalis* Q8. There are three tandem genes in the putative *lfr* locus identified as *lbpB*, *lbpA* and *orf3*. Potential promoter elements found upstream of the *lbpB* and *lbpA* genes are indicated by underlining.

Figure 5 shows a restriction map of clone pLDW1 containing the *lbpA*, *lbpB* and *orf3* genes from *M. catarrhalis* isolate Q8.

Figure 6 shows a comparison of the amino acid sequences of *Lbp1* from *M. catarrhalis* strains 4223 (SEQ ID No: 12) and Q8 (SEQ ID No: 16), *N. meningitidis* strains BNCV (SEQ ID No: 23) and H44/76 (SEQ ID No: 75), and *N. gonorrhoeae* strain FA19 (SEQ ID No: 24). Also shown is the partial carboxy terminal sequence of *Lbp2* from *N. meningitidis* strains BNCV (SEQ ID No: 76) and H44/76 (SEQ ID No: 77) and *N. gonorrhoeae* strain FA19 (SEQ ID No: 88). Dots indicate identical residues

and dashes have been introduced to achieve maximum sequence alignment.

Figure 7 shows a comparison of the amino acid sequences of Lbp2 from *M. catarrhalis* strains 4223 (SEQ ID No: 11), Q8 (SEQ ID No: 15) and VH19 (SEQ ID No: 70). Dots indicate identical residues. The arrow indicates the lipidated cysteine of a potential mature Lbp2 lipoprotein. The residues conserved with Tbp2 proteins are underlined and the RGD sequence is italicized.

Figure 8 shows a comparison of the amino acid sequences of Tbp2 (USPA No: 08/613,009) (SEQ ID No: 25) and Lbp2 from *M. catarrhalis* strain 4223 (SEQ ID No: 11). Dots indicate identical residues and dashes have been inserted to achieve maximum sequence alignment. The asterisks indicate conserved residues and the putative site of lipidation for both proteins is indicated by the arrow.

Figure 9 shows a comparison of the amino acid sequences of ORF3 from *M. catarrhalis* strains 4223 (SEQ ID No: 14) and Q8 (SEQ ID No: 18). Dots indicate identical residues and dashes have been introduced for maximum alignment.

Figure 10 shows the construction of plasmids for expression of recombinant Lbp1 protein from *E. coli*. Plasmids pRD1A and pRD1B express 4223 Lbp1 from the first or second methionine residues, respectively. Plasmids pQW1A and pQW1B express Q8 Lbp1 from the first or second methionine residues, respectively.

Figure 11, comprising panels A and B, shows the expression of recombinant Lbp1 (rLbp1) proteins from *E. coli*. Panel A shows the expression of the Q8 Lbp1 proteins and panel B shows the expression of the 4223 Lbp1 proteins. Lane 1, molecular weight marker. Lanes 2 and 3 demonstrate the induced expression of the

longer Lbp1 starting from the first methionine residues and lanes 4 and 5 illustrate the expression of the shorter Lbp1 proteins starting from the second methionine residues. Lanes 6, 7, 8 and 9 are uninduced samples.

Figure 12 shows the construction of plasmids for expression of recombinant Lbp2 (rLbp2) protein from *E. coli*. Plasmids pRD2A and pRD2B express 4223 Lbp2 with or without the native leader sequence, respectively. Plasmids pQW2A and pQW2B express Q8 Lbp2 with or without the native leader sequence, respectively.

Figure 13 shows the construction of a plasmid for expression of recombinant ORF3 (rORF3) proteins from *E. coli*.

Figure 14 shows a purification scheme for rLbp1 expressed from *E. coli*.

Figure 15 shows an SDS PAGE gel of the purification of Q8 Lbp1 from *E. coli*. Lane 1, BL21(DE3) lysate; lane 2, soluble proteins after 50 mM Tris/5 mM AEBSF/0.5 M NaCl, pH 8.0 extraction; lane 3, soluble proteins after 50 mM Tris/0.5% Triton X-100/10 mM EDTA, pH 8.0 extraction; lane 4, soluble proteins after 50 mM Tris-HCl/1% octylglucoside, pH 8.0 extraction; lane 5, solubilized inclusion bodies; lane 6, purified Lbp1.

Figure 16 shows the nucleotide sequence (SEQ ID No: 69) of the *M. catarrhalis* strain VH19 *lbpB* gene and the deduced amino acid sequence (SEQ ID No: 70) of the corresponding Lbp2 protein.

Figure 17 shows a partial restriction map of the *M. catarrhalis* strain VH19 *lbpB* gene.

Figure 18, comprising panels A, B and C, shows SDS-PAGE gels of the purification of recombinant Lbp proteins. Panel A shows an SDS-PAGE gel of the purification of Q8 rLbp1. Panels B and C show the

purification of Q8 rLbp2 and 4223 rLbp2, respectively. Lane 1, molecular weight markers; lane 2, whole cell lysates; lane 3, inclusion bodies; lane 4, purified protein.

5 Figure 19, comprising panels A and B, shows binding of recombinant Lbp proteins to lactoferrin. Panel A shows an SDS PAGE gel of purified recombinant proteins. Panel B shows the binding of recombinant proteins to human lactoferrin. Lane 1, molecular
10 weight markers; lane 2, Q8 rLbp1; lane 3, Q8 rLbp2; lane 4, 4223 rLbp2.

 Figure 20, comprising panels A, B and C, shows an immunoblot of *M. catarrhalis* strains reacted with anti-rLbp1 and anti-rLbp2 antibodies. Panel A: whole cell
15 lysates probed with anti-Q8 rLbp1 + anti-Q8 rLbp2 antisera. All cells were grown in the presence of EDDA. Panel B: whole cell lysates probed with anti-Q8 rLbp1 antibody. Panel C: whole cell lysates probed with anti-Q8 rLbp2 antibody. Lane 1, strain Q8; lane
20 2, strain 4223; lane 3, strain VH19; lane 4, strain LES-1; lane 5, strain H-04; lane 6, strain 3. + indicates growth in the presence of EDDA and - indicates growth in the absence of EDDA.

GENERAL DESCRIPTION OF THE INVENTION

25 Any *Moraxella* strain may be conveniently used to provide the purified and isolated nucleic acid, which may be in the form of DNA molecules, comprising at least a portion of the nucleic acid coding for a lactoferrin receptor as typified by embodiments of the
30 present invention. Such strains are generally available from clinical sources and from bacterial culture collections, such as the American Type Culture Collection.

In this application, the terms "lactoferrin receptor" (LfR) and "lactoferrin binding proteins" (Lbp) are used to define a family of Lbp1, Lbp2 and/or ORF3 proteins which includes those having variations in their amino acid sequences including those naturally occurring in various strains of, for example, *Moraxella*. The purified and isolated DNA molecules comprising at least a portion coding for lactoferrin receptor of the present invention also includes those encoding functional analogs of lactoferrin receptor proteins Lbp1, Lbp2 and/or Lbp3 of *Moraxella*. In this application, a first protein is an "analog" of a second protein if the first protein is immunologically related to and/or has the same function as the second protein. The analog may be, for example, a substitution, addition or deletion mutant thereof.

Lactoferrin receptor proteins were purified from *M. catarrhalis* membrane preparations by affinity chromatography on biotinylated human lactoferrin. Cyanogen bromide fragments were generated and amino acid sequence analysis of a 13 kDa fragment provided an internal Lbp1 sequence of MVQYTYRKGKENKAH (SEQ ID No: 22) underlined in Figure 6. The C-terminus of *M. catarrhalis* Tbp1 (United States Patent Application No. 08/613,009), *N. meningitidis* Tbp1 (ref. 27) and *H. influenzae* Tbp1 (ref. 31) has a conserved LEMKF (SEQ ID No: 26) sequence. Oligonucleotide primers were generated based upon these two sequences and used to PCR amplify an approximately 2.2kb fragment of the *lbpA* gene from *M. catarrhalis* strains 4223, Q8 and VH19. Partial sequence analysis demonstrated that the amplified genes were *lbpA* and not *tbpA* (see Fig. 1). The 2.2 kb PCR fragments were used to screen genomic libraries.

Chromosomal DNA from 4223, Q8 and VH19 was partially digested with Sau3A I and 15 to 23 kb fragments were purified before cloning into BamH I arms of the lambda vector EMBL3. The libraries were screened with the PCR fragment and positive clones were subjected to three rounds of plaque purification. Phage clone 4223LfR.17 containing an approximately 16 kb insert from 4223 and phage clone Q8LfR.13 containing an approximately 16 kb insert from Q8 were selected for further analysis.

Restriction enzyme and Southern blot analyses revealed that an internal Hind III fragment of approximately 9 kb contained at least a portion of the *lbpA* gene for both phage clones. The approximately 9 kb Hind III fragments were subcloned into pUC or pBluescript-based plasmids and sequenced. In each case, they contained the complete *lbpA* gene as well as an upstream gene identified as *lbpB*, and a downstream gene designated as *orf3*. The *lbpB-lbpA* gene arrangement is the same as present for *Neisseria* strains, but there has been no identification of a third gene for these organisms.

The gene arrangement is different than that observed for the *M. catarrhalis* *tfr* operon which was *tbpA-orf-tbpB* (United States Patent Application No. 08/613,009). There are promoter elements found upstream of both the *lbpB* and *lbpA* genes from strains 4223 and Q8. The third ORF is located immediately downstream of *lbpA*, separated by a single nucleotide.

By analogy with the *N. meningitidis* and *N. gonorrhoeae* transferrin receptor operons (ref. 26, 27, 28), the lactoferrin receptor operon was presumed to consist of two genes encoding lactoferrin binding proteins 1 and 2 (*Lbp1* and *Lbp2*) (ref. 29). However,

we report here that, for *M. catarrhalis*, there also appears to be a third gene located immediately downstream of *lbpA* encoding a potential lactoferrin binding protein 3 (ORF3).

5 The *M. catarrhalis* 4223 and Q8 *lbpA* genes encode proteins of molecular mass about 110 kDa and that are highly conserved with only seven residues difference between them. The N-terminal sequence of the native Lbp protein is unknown and there are two possible ATG
10 start codons at positions 1 or 16. The first of these is adjacent to consensus sequences for promoter elements and the second is followed by a putative signal sequence. The exact peptide sequence used to design the PCR amplification primers was not found.
15 When compared with other known Lbp1 sequences from *N. meningitidis* (refs. 31, 24) or *N. gonorrhoeae* (ref. 25) there is about 32% sequence identity and about 50% sequence homology between the *M. catarrhalis* and the *Neisseria* proteins. There is some homology between the
20 *M. catarrhalis* Lbp1 and Tbp1 proteins as shown in Figure 1, but it is very scattered.

 The *M. catarrhalis* 4223, Q8 and VH19 *lbpB* genes encode 898, 894 and 906 amino acid proteins, respectively. The *M. catarrhalis* Lbp2 proteins from
25 strains 4223 and Q8 are 92% identical and 95% homologous while that from VH19 is 77% identical and 84% similar to the 4223 and Q8 Lbp2 proteins (Figure 7). There is a consensus sequence for lipidation at the Cys³² residue, suggesting that Lbp2 is a lipoprotein
30 like Tbp2. There is little homology between the *M. catarrhalis* Lbp2 and Tbp2 proteins (Fig. 8) with the exception of a previously identified peptide sequence (LEGGFY (SEQ ID No: 27)) that is also found in *N. meningitidis* and *H. influenzae* Tbp2 (ref. 30).

The sequence of the proposed *M. catarrhalis* *lfr*-related downstream *orf3* is conserved between strains 4223 and Q8. The encoded 4223 and Q8 ORF3 proteins when compared to the PIR and Swiss Prot protein databases were found to be previously unknown. The ORF3 protein may bind lactoferrin itself or may be an associated or regulatory protein for Lbp1 and/or Lbp2.

Expression vectors have been assembled from the *lbpA* and *lbpB* genes and recombinant Lbp1 and Lbp2 proteins isolated and purified, as described in detail in the Examples below.

Results shown in Table 1 below illustrate the ability of anti-Lbp1 guinea pig antiserum, produced by immunization with affinity purified Lbp1, to lyse *M. catarrhalis*. The results show that the antisera produced by immunization with Lbp1 protein isolated from *M. catarrhalis* isolate 4223 was bactericidal against a homologous non-clumping *M. catarrhalis* strain RH408 (a strain previously deposited in connection with United States Patent Application No. 08/328,589, assigned to the assignee hereof (WO 96/12733 published May 2, 1996)) derived from isolate 4223. In addition, antisera produced by immunization with Lbp1 protein isolated from *M. catarrhalis* 4223 were bactericidal against the heterologous non-clumping strain Q8. The results in Table 3 show that similarly-produced anti-Lbp2 guinea pig antiserum was bactericidal for the homologous strain and for three of five heterologous strains. The ability of isolated and purified lactoferrin binding protein to generate bactericidal antibodies is *in vivo* evidence of utility of these proteins as vaccines to protect against disease caused by *Moraxella*.

Thus, in accordance with another aspect of the present invention, there is provided a vaccine against *Moraxella* comprising an immunogenically-effective amount of lactoferrin binding protein or fragment or analog thereof, or a nucleic acid molecule (DNA or RNA) encoding the lactoferrin binding protein or fragment or analog thereof, and a physiologically-acceptable carrier therefor. The lactoferrin binding protein or fragment or analog thereof provided herein may also be used as a carrier protein for haptens, polysaccharide or peptides to make conjugate vaccines against antigenic determinants unrelated to lactoferrin binding proteins.

In additional embodiments of the present invention, therefore, the lactoferrin binding protein as provided herein may be used as a carrier molecule to prepare chimeric molecules and conjugate vaccines (including glycoconjugates) against pathogenic bacteria, including encapsulated bacteria. Thus, for example, glycoconjugates of the present invention may be used to confer protection against disease and infection caused by any bacteria having polysaccharide antigens including lipooligosaccharides (LOS) and PRP. Such bacterial pathogens may include, for example, *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Escherichia coli*, *Neisseria meningitidis*, *Salmonella typhi*, *Streptococcus mutans*, *Cryptococcus neoformans*, *Klebsiella*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Particular antigens which can be conjugated to lactoferrin binding protein and methods to achieve such conjugations are described in U.S. Patent Application No. 08/433,522 filed November 23, 1993 (WO 94/12641), assigned to the assignee hereof and the disclosure of which is hereby incorporated by reference thereto.

In another embodiment, the carrier function of lactoferrin binding protein may be used, for example, to induce an immune response against abnormal polysaccharides of tumour cells, or to produce anti-tumour antibodies that can be conjugated to chemotherapeutic or bioactive agents.

The lactoferrin binding protein provided herein is useful as a diagnostic reagent, as an antigen or for the generation of anti-lactoferrin protein binding antibodies, antigen for vaccination against disease caused by species of *Moraxella* and for detecting infection by *Moraxella* and other such bacteria.

The invention extends to lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same from *Moraxella catarrhalis* for use as an active ingredient in a vaccine against disease caused by infection with *Moraxella*. The invention also extends to a pharmaceutical vaccinal composition containing lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same from *Moraxella catarrhalis* and optionally, a pharmaceutically acceptable carrier and/or diluent.

In a further aspect the invention provides the use of lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same for the preparation of a pharmaceutical vaccinal composition for immunization against disease caused by infection with *Moraxella*.

It is clearly apparent to one skilled in the art, that the various embodiments of the present invention have many applications in the fields of vaccination, diagnosis, treatment of, for example, *Moraxella* infections and the generation of immunological and other

diagnostic reagents. A further non-limiting discussion of such uses is further presented below.

1. Vaccine Preparation and Use

Immunogenic compositions, suitable to be used as
5 vaccines, may be prepared from immunogenic lactoferrin
receptor proteins, analogs and fragments thereof encoded
by the nucleic acid molecules as well as the nucleic
acid molecules disclosed herein. The vaccine elicits an
immune response which produces antibodies, including
10 anti-lactoferrin receptor antibodies and antibodies that
are opsonizing or bactericidal. Should the vaccinated
subject be challenged by *Moraxella*, the antibodies bind
to the lactoferrin receptor and thereby prevent access
of the bacteria to an iron source which is required for
15 viability. Furthermore, opsonizing or bactericidal
anti-lactoferrin receptor antibodies may also provide
protection by alternative mechanisms.

Immunogenic compositions, including vaccines, may
be prepared as injectables, as liquid solutions or
20 emulsions. The lactoferrin receptor proteins, analogs
and fragments thereof and encoding nucleic acid
molecules as well as the nucleic acid molecules
described herein may be mixed with pharmaceutically
acceptable excipients which are compatible with the
25 lactoferrin receptor proteins, fragments, analogs or
nucleic acid molecules. Such excipients may include
water, saline, dextrose, glycerol, ethanol, and
combinations thereof. The immunogenic compositions and
vaccines may further contain auxiliary substances, such
30 as wetting or emulsifying agents, pH buffering agents,
or adjuvants, to enhance the effectiveness of the
vaccines. Immunogenic compositions and vaccines may be
administered parenterally, by injection subcutaneously,
intradermally or intramuscularly. Alternatively, the
35 immunogenic compositions provided according to the

present invention, may be formulated and delivered in a manner to evoke an immune response at mucosal surfaces. Thus, the immunogenic composition may be administered to mucosal surfaces by, for example, the nasal or oral (intragastric) routes. The immunogenic composition may be provided in combination with a targeting molecule for delivery to specific cells of the immune system or to mucosal surfaces. Some such targeting molecules include vitamin B12 and fragments of bacterial toxins, as described in WO 92/17167 (Biotech Australia Pty. Ltd.), and monoclonal antibodies, as described in U.S. Patent No. 5,194,254 (Barber et al). Alternatively, other modes of administration, including suppositories and oral formulations, may be desirable. For suppositories, binders and carriers may include, for example, polyalkalene glycols or triglycerides. Oral formulations may include normally employed incipients such as, for example, pharmaceutical grades of saccharine, cellulose and magnesium carbonate. These compositions may take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 1 to 95% of the lactoferrin receptor proteins, fragments, analogs and/or nucleic acid molecules.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective, protective and immunogenic. The quantity to be administered depends on the subject to be treated, including, for example, the capacity of the individual's immune system to synthesize antibodies, and, if needed, to produce a cell-mediated immune response. Precise amounts of active ingredient required to be administered depend on the judgement of the practitioner. However, suitable dosage ranges are readily determinable by one skilled in

the art and may be of the order of micrograms of the lactoferrin receptor proteins, analogs and fragments thereof and/or nucleic acid molecules. Suitable regimes for initial administration and booster doses are also
5 variable, but may include an initial administration followed by subsequent administrations. The dosage of the vaccine may also depend on the route of administration and will vary according to the size of the host.

10 The nucleic acid molecules encoding the lactoferrin receptor of *Moraxella* may be used directly for immunization by administration of the DNA directly, for example, by injection for genetic immunization or by constructing a live vector, such as *Salmonella*, BCG,
15 adenovirus, poxvirus, vaccinia or poliovirus containing the nucleic acid molecules. A discussion of some live vectors that have been used to carry heterologous antigens to the immune system is contained in, for example, O'Hagan (ref. 18). Processes for the direct
20 injection of DNA into test subjects for genetic immunization are described in, for example, Ulmer et al. (ref. 19).

Immunogenicity can be significantly improved if the antigens are co-administered with adjuvants, commonly
25 used as an 0.05 to 1.0 percent solution in phosphate - buffered saline. Adjuvants enhance the immunogenicity of an antigen but are not necessarily immunogenic themselves. Adjuvants may act by retaining the antigen locally near the site of administration to produce a
30 depot effect facilitating a slow, sustained release of antigen to cells of the immune system. Adjuvants can also attract cells of the immune system to an antigen depot and stimulate such cells to elicit immune responses.

Immunostimulatory agents or adjuvants have been used for many years to improve the host immune responses to, for example, vaccines. Intrinsic adjuvants, such as lipopolysaccharides, normally are the components of killed or attenuated bacteria used as vaccines. Extrinsic adjuvants are immunomodulators which are typically non-covalently linked to antigens and are formulated to enhance the host immune responses. Thus, adjuvants have been identified that enhance the immune response to antigens delivered parenterally. Some of these adjuvants are toxic, however, and can cause undesirable side-effects, making them unsuitable for use in humans and many animals. Indeed, only aluminum hydroxide and aluminum phosphate (collectively commonly referred to as alum) are routinely used as adjuvants in human and veterinary vaccines. The efficacy of alum in increasing antibody responses to diphtheria and tetanus toxoids is well established and an HBsAg vaccine has been adjuvanted with alum.

A wide range of extrinsic adjuvants can provoke potent immune responses to antigens. These include saponins complexed to membrane protein antigens (immune stimulating complexes), pluronic polymers with mineral oil, killed mycobacteria and mineral oil, Freund's complete adjuvant, bacterial products, such as muramyl dipeptide (MDP) and lipopolysaccharide (LPS), as well as lipid A, and liposomes.

To efficiently induce humoral immune responses (HIR) and cell-mediated immunity (CMI), immunogens are often emulsified in adjuvants. Many adjuvants are toxic, inducing granulomas, acute and chronic inflammations (Freund's complete adjuvant, FCA), cytotoxicity (saponins and pluronic polymers) and pyrogenicity, arthritis and anterior uveitis (LPS and MDP). Although FCA is an excellent adjuvant and widely

used in research, it is not licensed for use in human or veterinary vaccines because of its toxicity.

Desirable characteristics of ideal adjuvants include:

- 5 (1) lack of toxicity;
- (2) ability to stimulate a long-lasting immune response;
- (3) simplicity of manufacture and stability in long-term storage;
- 10 (4) ability to elicit both CMI and HIR to antigens administered by various routes, if required;
- (5) synergy with other adjuvants;
- (6) capability of selectively interacting with populations of antigen presenting cells (APC);
- 15 (7) ability to specifically elicit appropriate T_H1 or T_H2 cell-specific immune responses; and
- (8) ability to selectively increase appropriate antibody isotype levels (for example, IgA) against antigens.

20 U.S. Patent No. 4,855,283 granted to Lockhoff et al on August 8, 1989, which is incorporated herein by reference thereto, teaches glycolipid analogues including N-glycosylamides, N-glycosylureas and N-glycosylcarbamates, each of which is substituted in the
25 sugar residue by an amino acid, as immuno-modulators or adjuvants. Thus, Lockhoff et al. 1991 (ref. 20) reported that N-glycolipid analogs displaying structural similarities to the naturally-occurring glycolipids, such as glycopospholipids and glycoglycerolipids, are
30 capable of eliciting strong immune responses in both herpes simplex virus vaccine and pseudorabies virus vaccine. Some glycolipids have been synthesized from long chain-alkylamines and fatty acids that are linked directly with the sugars through the anomeric carbon

atom, to mimic the functions of the naturally occurring lipid residues.

U.S. Patent No. 4,258,029 granted to Moloney, assigned to the assignee hereof and incorporated herein by reference thereto, teaches that octadecyl tyrosine hydrochloride (OTH) functions as an adjuvant when complexed with tetanus toxoid and formalin inactivated type I, II and III poliomyelitis virus vaccine. Also, Nixon-George et al. 1990, (ref. 21) reported that octadecyl esters of aromatic amino acids complexed with a recombinant hepatitis B surface antigen, enhanced the host immune responses against hepatitis B virus.

2. Immunoassays

The lactoferrin receptor proteins, analogs and/or fragments thereof of the present invention are useful as immunogens, as antigens in immunoassays including enzyme-linked immunosorbent assays (ELISA), RIAs and other non-enzyme linked antibody binding assays or procedures known in the art for the detection of anti-*Moraxella*, lactoferrin receptor protein antibodies. In ELISA assays, the lactoferrin receptor protein, analogs and/or fragments corresponding to portions of Lfr protein, are immobilized onto a selected surface, for example, a surface capable of binding proteins or peptides such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed lactoferrin receptor, analogs and/or fragments, a non-specific protein such as a solution of bovine serum albumin (BSA) or casein that is known to be antigenically neutral with regard to the test sample may be bound to the selected surface. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by non-specific bindings of antisera onto the surface.

The immobilizing surface is then contacted with a sample, such as clinical or biological materials, to be tested in a manner conducive to immune complex (antigen/antibody) formation. This procedure may include diluting the sample with diluents, such as BSA, bovine gamma globulin (BGG) and/or phosphate buffered saline (PBS)/Tween. The sample is then allowed to incubate for from about 2 to 4 hours, at temperatures such as of the order of about 25° to 37°C. Following incubation, the sample-contacted surface is washed to remove non-immunocomplexed material. The washing procedure may include washing with a solution such as PBS/Tween or a borate buffer.

Following formation of specific immunocomplexes between the test sample and the bound lactoferrin receptor protein, analogs and/or fragments and subsequent washing, the occurrence, and even amount, of immunocomplex formation may be determined by subjecting the immunocomplex to a second antibody having specificity for the first antibody. If the test sample is of human origin, the second antibody is an antibody having specificity for human immunoglobulins and in general IgG. To provide detecting means, the second antibody may have an associated activity such as an enzymatic activity that will generate, for example, a color development upon incubating with an appropriate chromogenic substrate. Quantification may then be achieved by measuring the degree of color generation using, for example, a spectrophotometer.

30 3. Use of Sequences as Hybridization Probes

The nucleotide sequences of the present invention, comprising the sequence of the lactoferrin receptor gene, now allow for the identification and cloning of the lactoferrin receptor genes from any species of *Moraxella*.

The nucleotide sequences comprising the sequence of the lactoferrin receptor genes of the present invention are useful for their ability to selectively form duplex molecules with complementary stretches of other *lfr* genes. Depending on the application, a variety of hybridization conditions may be employed to achieve varying degrees of selectivity of the probe toward the other *lfr* genes. For a high degree of selectivity, relatively stringent conditions are used to form the duplexes, such as low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of between about 50°C to 70°C. For some applications, less stringent hybridization conditions are required such as 0.15 M to 0.9 M salt, at temperatures ranging from between about 20°C to 55°C. Hybridization conditions can also be rendered more stringent by the addition of increasing amounts of formamide, to destabilize the hybrid duplex. Thus, particular hybridization conditions can be readily manipulated, and will generally be a method of choice depending on the desired results. In general, convenient hybridization temperatures in the presence of 50% formamide are: 42°C for a probe which is 95 to 100% homologous to the target fragment, 37°C for 90 to 95% homology and 32°C for 85 to 90% homology.

Such hybridization conditions may be employed to determine DNA sequences which encode a functional lactoferrin receptor of *Moraxella* and which hybridize under stringent conditions to any one of the DNA sequences (a) or (b), described above.

In a clinical diagnostic embodiment, the nucleic acid sequences of the *lfr* genes of the present invention may be used in combination with an appropriate means, such as a label, for determining hybridization. A wide

variety of appropriate indicator means are known in the art, including radioactive, enzymatic or other ligands, such as avidin/biotin and digoxigenin-labelling, which are capable of providing a detectable signal. In some
5 diagnostic embodiments, an enzyme tag such as urease, alkaline phosphatase or peroxidase, instead of a radioactive tag may be used. In the case of enzyme tags, colorimetric indicator substrates are known which can be employed to provide a means visible to the human
10 eye or spectrophotometrically, to identify specific hybridization with samples containing *lfr* gene sequences.

The nucleic acid sequences of *lfr* genes of the present invention are useful as hybridization probes in
15 solution hybridizations and in embodiments employing solid-phase procedures. In embodiments involving solid-phase procedures, the test DNA (or RNA) from samples, such as clinical samples, including exudates, body fluids (e. g., serum, amniotic fluid, middle ear
20 effusion, sputum, bronchoalveolar lavage fluid) or even tissues, is adsorbed or otherwise affixed to a selected matrix or surface. The fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes comprising the nucleic acid sequences of
25 the *lfr* genes or fragments thereof of the present invention under desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required depending on, for example, the G+C contents, type of target nucleic
30 acid, source of nucleic acid, size of hybridization probe etc. Following washing of the hybridization surface so as to remove non-specifically bound probe molecules, specific hybridization is detected, or even quantified, by means of the label. It is preferred to
35 select nucleic acid sequence portions which are

conserved among species of *Moraxella*. The selected probe may be at least 18bp and may be in the range of about 30 to 90 bp.

4. Expression of the Lactoferrin Receptor Genes

5 Plasmid vectors containing replicon and control sequences which are derived from species compatible with the host cell may be used for the expression of the lactoferrin receptor genes in expression systems. The vector ordinarily carries a replication site, as well as
10 marking sequences which are capable of providing phenotypic selection in transformed cells. For example, *E. coli* may be transformed using pBR322 which contains genes for ampicillin and tetracycline resistance and thus provides easy means for identifying transformed
15 cells. The pBR322 plasmid, or other microbial plasmid or phage, must also contain, or be modified to contain, promoters which can be used by the host cell for expression of its own proteins.

In addition, phage vectors containing replicon and
20 control sequences that are compatible with the host can be used as a transforming vector in connection with these hosts. For example, the phage in lambda GEMTM-11 may be utilized in making recombinant phage vectors which can be used to transform host cells, such as *E.*
25 *coli* LE392.

Promoters commonly used in recombinant DNA construction include the β -lactamase (penicillinase) and lactose promoter systems and other microbial promoters, such as the T7 promoter system as described in U.S.
30 Patent No. 4,952,496. Details concerning the nucleotide sequences of promoters are known, enabling a skilled worker to ligate them functionally with genes. The particular promoter used will generally be a matter of choice depending upon the desired results. Hosts that

are appropriate for expression of the lactoferrin receptor genes, fragments or analogs thereof, may include *E. coli*, *Bacillus* species, *Haemophilus*, fungi, yeast, *Moraxella*, *Bordetella*, or the baculovirus expression system may be used.

In accordance with this invention, it is preferred to produce the lactoferrin receptor protein, fragment or analog thereof, by recombinant methods, particularly since the naturally occurring LfR protein as purified from a culture of a species of *Moraxella* may include trace amounts of toxic materials or other contaminants. This problem can be avoided by using recombinantly produced LfR protein in heterologous systems which can be isolated from the host in a manner to minimize contaminants, including other proteins of the *Moraxella* strain, in the purified material. Particularly desirable hosts for expression in this regard include Gram positive bacteria which do not have LPS and are, therefore, endotoxin free. Such hosts include species of *Bacillus* and may be particularly useful for the production of non-pyrogenic lactoferrin receptor proteins, fragments or analogs thereof. Furthermore, recombinant methods of production permit the manufacture of Lbp1 or Lbp2 or ORF3 or respective analogs or fragments thereof, separate from one another which is distinct from the normal combined proteins present in *Moraxella*.

Sequence Alignment and Analysis

Sequence alignments were performed using the ALIGN (Trademark) or GENALIGN (Trademark) computer programs (Inteligenetics Suite 5.4, Oxford Molecular). ALIGN® uses the Needleman-Wunsch algorithm (ref. 35) and its later modifications to locate regions of similarity between two sequences. Finding regions of maximum

similarity between two sequences can be solved in a rigorous manner using the iterative matrix calculation of the Needleman and Wunsch 1997 algorithm. The analysis is restricted to regions with no internal deletions or insertions, joined by a minimum number of loop-outs or deletions. Sellers (ref. 36) developed a true metric measure of the "distance" between sequences and Waterman (ref. 37) extended this algorithm to include insertions and deletions of arbitrary length. Smith (ref. 38) improved the early algorithms to find the subsequences of maximum similarity. The algorithm has been used to analyze sequences as long as 5000 bases by dividing these sequences into segments of 200 to 400 bases, and then reassembling them into a final best match. This method of dividing the sequence and then reassembling it has proven quite robust. The algorithm permits the size of the segment to be specified which the program searches for similarities. The program then assembles the segments after checking overlaps of adjacent subsequences. The weighting of deletions and the relative size of overlaps may be controlled. The program displays the results to show the differences in closely related sequences.

GENALIGN® is a multiple alignment program. Up to 99 sequences using the Martinez/Regions (ref. 39) or Needleman-Wunsch (ref. 35) method may be analyzed for alignment. GENALIGN places the sequences in an order that puts the most closely aligned sequence pairs adjacent to each other. A consensus sequence is displayed under the multiple sequence alignments. The sequences used in developing the consensus sequence file for use in other programs. GENALIGN allows the parameters of the search to be changed so that alternate alignments of the sequences can be formed.

These programs are used employing their default settings. The default settings are as follows:

FastDB

5 AMINO-Res-length = 2
 DEletion-weight = 5.00
 LEngth-factor = 0
 Matching-weight = 1.00
 NUCLEIC-Res-length = 4
 SPread-factor = 50

10

Findseq

Search Parameters:

Similarity matrix Unitary
 K-tuple 4
 Mismatch penalty 1
 15 Joining Penalty 30
 Randomization group length 0
 Cutoff score 5

Alignment Parameters:

Window size 32
 20 Gap penalty 1.00
 Gap size penalty 0.33

Such procedures may be used to determine DNA sequences which encode a functional lactoferrin receptor of *Moraxella* and which may have at least about 90% sequence identity with any one of the DNA sequences (a) or (b), described above.

25

Biological Deposits

Certain vectors that contain at least a portion coding for a lactoferrin receptor protein from strains of *Moraxella catarrhalis* strain 4223 and Q8 and a strain of *M. catarrhalis* RH408 that are described and referred to herein have been deposited with the American Type Culture Collection (ATCC) located at 12301 Parklawn Drive, Rockville, Maryland 20852, USA, pursuant to the

30

Budapest Treaty and prior to the filing of this application. Samples of the deposited vectors and bacterial strain will become available to the public and the restrictions imposed on access to the deposits will be removed upon grant of a patent based upon this United States patent application. In addition, the deposit will be replaced if viable samples cannot be dispensed by the Depository. The invention described and claimed herein is not to be limited in scope by the biological materials deposited, since the deposited embodiment is intended only as an illustration of the invention. Any equivalent or similar vectors or strains that encode similar or equivalent antigens as described in this application are within the scope of the invention.

Deposit Summary

Deposit	ATCC Designation	Date deposited
Plasmid pLD1-8	97,997	April 23, 1997
Plasmid pLDW1	97,998	April 23, 1997
Strain RH408	55,637	Dec. 9, 1994

EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific Examples. These Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations.

Methods of molecular genetics, protein biochemistry and immunology used but not explicitly described in this disclosure and these Examples are amply reported in the scientific literature and are well within the ability of those skilled in the art.

Example 1

This Example illustrates the generation of oligonucleotide primers for PCR amplification of *M. catarrhalis* *lbpA*.

Native Lbp1 was purified by affinity chromatography using high stringency conditions as described in United States Patent Application No. 08/552,232, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference, and in ref. 40.

The purified Lbp1 protein was digested overnight with cyanogen bromide, then fragments separated by SDS PAGE and submitted to sequence analysis on an ABI model 477A. A 13 kDa protein fragment was found to have the N-terminal sequence MVQYTYRKGKKNKAH (SEQ ID No: 22). A degenerate oligonucleotide primer (4393.RD) was prepared based upon this sequence:

	Q	Y	T	R	K	G	E	N	K	A	(SEQ ID No: 28)
25	5'										3'
	CAA	TAT	ACI	CGT	AAA	GGT	GAA	AAT	AAA	GC	(SEQ ID No: 29)
	CAA	TAT	ACI	CGT	AAA	GGC	GAA	AAC	AAA	GC	(SEQ ID No: 30)
	CAA	TAT	ACI	CGT	AAA	GGT	GAA	AAC	AAA	GC	(SEQ ID No: 31)
	CAA	TAT	ACI	CGT	AAA	GGC	GAA	AAT	AAA	GC	(SEQ ID No: 32)
30	CAA	TAT	ACI	CGC	AAA	GGC	GAA	AAC	AAA	GC	(SEQ ID No: 33)
	CAA	TAT	ACI	CGC	AAA	GGC	GAA	AAT	AAA	GC	(SEQ ID No: 34)
	CAA	TAT	ACI	CGC	AAA	GGT	GAA	AAT	AAA	GC	(SEQ ID No: 35)
	CAA	TAT	ACI	CGC	AAA	GGT	GAA	AAC	AAA	GC	(SEQ ID No: 36)

The Y⁶ and K¹⁰ residues were omitted from the sequence analysis report for the N-terminal sequence and the oligonucleotides used to PCR amplify the 2.2 kb fragment were incorrect, but nevertheless were successful.

There is a conserved C-terminal pentapeptide found in all known Lbp1 and Tbp1 protein sequences: LEMKF (SEQ ID No. 26). An oligonucleotide primer (4572.RD) was prepared based upon the complementary DNA sequence encoding this pentapeptide:

L E M K F *

5' CTT GAA ATG AAG TTT TAA 3' (SEQ ID No: 37)

3' GAA CTT TAC TTC AAA ATT 5' 4572.RD (SEQ ID No: 38)

Example 2

This Example illustrates the preparation of chromosomal DNA from *M. catarrhalis* strains 4223 and Q8.

M. catarrhalis isolate 4223 was inoculated into 100 ml of BHI broth, and incubated for 18 hr at 37°C with shaking. The cells were harvested by centrifugation at 10,000 x g for 20 min. The pellet was used for extraction of *M. catarrhalis* 4223 chromosomal DNA.

The cell pellet was resuspended in 20 ml of 10 mM Tris-HCl (pH 7.5)-1.0 mM EDTA (TE). Pronase and SDS were added to final concentrations of 500 µg/ml and 1.0%, respectively, and the suspension was incubated at 37°C for 2 hr. After several sequential extractions with phenol, phenol:chloroform (1:1), and chloroform:isoamyl alcohol (24:1), the aqueous extract was dialysed, at 4°C, against 1.0 M NaCl for 4 hr, and against TE (pH 7.5) for a further 48 hr with three buffer changes. Two volumes of ethanol were added to the dialysate, and the DNA was spooled onto a glass

rod. The DNA was allowed to air-dry, and was dissolved in 3.0 ml of water. Concentration was estimated, by UV spectrophotometry, to be about 290 µg/ml.

M. catarrhalis strain Q8 was grown in BHI broth.

5 Cells were pelleted from 50 ml of culture by centrifugation at 5000 rpm for 20 minutes, at 4°C. The cell pellet was resuspended in 10 ml of TE (10 mM Tris-HCl, 1 mM EDTA, pH 7.5) and proteinase K and SDS were added to final concentrations of 500 µg/ml and 1%,
10 respectively. The sample was incubated at 37°C for 4 hours until a clear lysate was obtained. The lysate was extracted twice with Tris-saturated phenol/chloroform (1:1), and twice with chloroform. The final aqueous phase was dialysed for 24 hours
15 against 2 X 1000 ml of 1 M NaCl at 4°C, changing the buffer once, and for 24 hours against 2 x 1000 ml of TE at 4°, changing the buffer once. The final dialysate was precipitated with two volume of 100% ethanol. The DNA was spooled, dried and resuspended in 5 to 10 ml of
20 TE buffer.

Example 3

This Example illustrates the PCR amplification of a fragment of *M. catarrhalis lbpA* and the generation of probes for screening libraries.

25 PCR amplification was performed on chromosomal DNA isolated in Example 2 using primers 4393.RD and 4572.RD under the following cycling conditions: 25 cycles of 94°C for 1 min, 47°C for 30 sec and 72°C for 1 min. PCR4 is the amplification of the 4223 *lbpA* fragment and
30 PCR5 is the amplification of the Q8 *lbpA* fragment. A specific band of about 2.2 kb was amplified and partial sequence analysis was performed to ensure that the gene product was related to *lbpA* and was not *tbpA*. The derived amino acid sequences are shown in Figure 1 and
35 have been aligned with the complete 4223 *Lbp1* sequence

to show their placement and the 4223 Tbp1 sequence (USAN 08/613,009) to indicate their uniqueness.

The full-length 2.2 kb gene fragment was randomly labeled with ^{32}P and used to probe genomic libraries.

5 **Example 4**

This Example illustrates the generation and screening of the EMBL 3 libraries.

Chromosomal DNA was prepared as described in Example 2. A series of Sau3AI restriction digests of
10 chromosomal DNA, in final volumes of 10 μL each, were carried out in order to optimize the conditions necessary to generate maximal amounts of restriction fragments within a 15 to 23 kb size range. Using the optimized digestion conditions, a large-scale digestion
15 was set up in a 100 μL volume, containing the following: 50 μL of chromosomal DNA (290 $\mu\text{g}/\text{ml}$), 33 μL water, 10 μL 10X Sau3A buffer (New England Biolabs), 1.0 μL BSA (10 mg/ml , New England Biolabs), and 6.3 μL Sau3A (0.04 U/ μL). Following a 15 min. incubation at
20 37°C, the digestion was terminated by the addition of 10 μL of 100 mM Tris-HCl (pH 8.0)-10 mM EDTA-0.1% bromophenol blue-50% glycerol (loading buffer). Digested DNA was electrophoresed through a 0.5% agarose gel in 40 mM Tris acetate-2 mM $\text{Na}_2\text{EDTA} \cdot 2\text{H}_2\text{O}$ (pH 8.5) (TAE
25 buffer) at 50 V for 6 hr. The region containing restriction fragments within a 15 to 23 kb molecular size range was excised from the gel, and placed into dialysis tubing containing 3.0 ml of TAE buffer. DNA was electroeluted from the gel fragment by applying a
30 field strength of 1.0 V/cm for 18 hr. Electroeluted DNA was extracted once each with phenol and phenol:chloroform (1:1), and precipitated with ethanol. The dried DNA was dissolved in 5.0 μL water.

Size-fractionated chromosomal DNA was ligated with

*Bam*HI-digested EMBL3 arms (Promega), using T4 DNA ligase in a final volume of 9 μ L. The entire ligation mixture was packaged into lambda phage using a commercial packaging kit (Amersham), following manufacturer's instructions.

The packaged DNA library was amplified on solid media. 0.1 ml aliquots of *Escherichia coli* strain NM539 in 10 mM MgSO_4 ($\text{OD}_{260} = 0.5$) were incubated at 37°C for 15 min. with 15 to 25 μ L of the packaged DNA library. Samples were mixed with 3 ml of 0.6% agarose containing 1.0% BBL trypticase peptone-0.5% NaCl (BBL top agarose), and mixtures were plated onto 1.5% agar plates containing 1.0% BBL trypticase peptone-0.5% NaCl, and incubated at 37°C for 18 hr. 3 ml quantities of 50 mM Tris-HCl (pH 7.5)-8 mM magnesium sulfate heptahydrate-100 mM NaCl-0.01% (w/v) gelatin (SM buffer) were added to each plate, and plates were left at 4°C for 7 hr. SM buffer containing phage was collected from the plates, pooled together, and stored in a screwcap tube at 4°C, with chloroform.

Ten μ L aliquots of phage stock were combined each with 100 μ L of *E. coli* strain LE392 in 10 mM MgSO_4 ($\text{OD}_{260} = 0.5$) (plating cells), and incubated at 37°C for 15 min. The samples were mixed with 3 ml each of BBL top agarose, and the mixtures were poured onto 1.5% agarose plates containing 1% bacto tryptone-0.5% bacto yeast extract-0.05% NaCl (LB agarose; Difco) and supplemented with 200 μ M EDDA. The plates were incubated at 37°C for 18 hr. Plaques were lifted onto nitrocellulose filters (Amersham Hybond-C Extra) which were hybridized with the ^{32}P -labelled 2.2 kb PCR fragment. Several putative phage clones were obtained from each library and clones 4223LfR.17 and Q8LfR.13 were chosen for further analysis.

Example 5

This Example illustrates the subcloning of the phage clones containing *M. catarrhalis lfr* genes.

5 Restriction enzyme analysis and Southern blotting using the screening probes, indicated that at least a portion of *lbpA* was localized to an about 9 kb Hind III fragment from each phage clone. The about 9 kb Hind III fragment from 4223Lfr.17 was subcloned into pUC 18, generating clone pLD1-8. The about 9 kb Hind III
10 fragment from Q8Lfr.13 was subcloned into pBluescript, generating plasmid pLDW1. Internal about 5.5 kb EcoR V fragments were subcloned generating plasmids pLD3 and pLDW3 for the 4223 and Q8 genes, respectively.

15 **Example 6**

This Example illustrates the sequence analysis of clones containing the *M. catarrhalis lfr* genes from strains 4223 and Q8.

Sequence analysis of the 5.5 kb EcoR V fragments
20 from pLD3 and pLDW3, revealed that they each contained the 3'-end of *lbpB*, the complete *lbpA* gene, and a third complete gene designated *orf3*. The remainder of the *lbpB* genes was found on the about 9 kb Hind III fragments from pLD1-8 and pLDW1. Partial restriction
25 enzyme analysis of the 4223 *lbpA*, *lbpB*, and *orf3* genes, based upon the nucleotide sequences is shown in Figure 3. Partial restriction enzyme analysis of the Q8 *lbpA*, *lbpB*, and *orf3* genes, based upon the nucleotide
30 sequences is shown in Figure 5. The complete sequences of the *lbpB*, *lbpA*, and *orf3* genes comprising the putative *lfr* locus from *M. catarrhalis* 4223 and Q8 is shown in Figures 2 and 4, respectively. The intergenic distance between the *lbpB* and *lbpA* genes is 184 nucleotides, while a single nucleotide separates the

lbpA and *orf3* genes. A putative promoter and ribosome binding site is indicated by underlining upstream of both *lbpB* and *lbpA*. A fourth potential gene was cloned on the approximately 9 kb Hind III fragments.

5 The N-terminal sequence of the native Lbp1 protein is unknown. Examination of the deduced amino acid sequence of the *lbpA* gene indicates that there are two possible ATG start codons at positions 1 and 16. The first position is downstream of strong promoter
10 elements found in the *lbpB-lbpA* intergenic region and the second position is followed by a putative signal sequence. The *M. catarrhalis* 4223 and Q8 Lbp1 proteins (from the first ATG) have molecular mass values of about 110 kDa and are 99% identical. The deduced Lbp1
15 protein sequences from *M. catarrhalis* strains 4223 and Q8 are compared in Figure 6. They are also compared with the *iroA/lbpA* gene from *N. meningitidis* strain BNCV (ref. 24) and the *lbpA* gene from *N. gonorrhoeae* strain FA19 (ref. 25). The *M. catarrhalis* proteins are
20 found to be about 32% identical and about 50% similar to the *Neisseria* proteins. As shown in Figure 1, there is very limited sequence homology between the *M. catarrhalis* Tbp1 and Lbp1 sequences.

 The deduced Lbp2 protein sequences from *M.*
25 *catarrhalis* strains 4223 and Q8 are compared in Figure 7. The 4223 and Q8 Lbp2 proteins both have molecular masses of about 99 kDa and are 92% identical and 95% similar to each other. A comparison to the *M. catarrhalis* Tbp2 proteins shows very little homology
30 except the LEGGFY (SEQ ID No: 27) epitope previously identified in *H. influenzae* and *N. meningitidis* Tbp2 proteins (Fig. 8). A cysteine residue at position 32 is preceded by a consensus sequence for lipoproteins suggesting that Lbp2, like Tbp2, is a lipoprotein. An

unusual feature of the Lbp2 proteins is the high combined aspartic acid and asparagine content which is nearly 20%. In addition, the 4223 Lbp2 amino acid composition from residues 698 to 751 is about 52% aspartic acid.

The 4223 and Q8 *lfr orf3* genes would encode proteins of molecular mass about 60 kDa, respectively. A notable feature of the ORF3 protein is a potential signal sequence, a terminal phenylalanine which is often associated with membrane anchored proteins, an internal repeat sequence of DGLG (SEQ ID No: 39), and a high leucine content of 15%. The deduced Lbp3 protein sequences are compared in Figure 9. These proteins are 98% identical and 99% similar.

Example 7

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp1 from the first methionine in *E. coli*.

There are two possible start codons at the beginning of the *lbpA* gene and hence two expression constructs were made. The construction scheme for 4223 or Q8 *lbpA* expressed from the first methionine is shown in Figure 10. An approximately 200 bp fragment of the 5'-end of *lbpA* from the ATG to a BstE II site was PCR amplified using primers 5405.RD and 5407.RD. An Nde I site was engineered at the 5'-end to facilitate cloning into the pT7-7 vector.

NdeI

M S K S I T (SEQ ID No: 40)

5' GGAATTCCAT ATG TCA AAA TCT ATC ACA AA 3' 5405.RD

(SEQ ID No: 41)

BstE II

L D A I T V T A A (SEQ ID No: 42)

5' T TTA GAT GCC ATC ACG GTA ACC GCC GCC CC 3' (SEQ ID No: 43)

3' A AAT CTA CGG TAG TGC CAT TGG CGG CGG GG 5' 5407.RD

(SEQ ID No: 44)

5 In order to subclone the *lbpA* gene into pT7-7, a approximately 515 bp fragment of the 3'-end of the gene from an Sph I site to the stop codon was PCR amplified using primers 5281.RD and 5282.RD and a BamH1 site was engineered at 3'-end.

Sph I

10 G K L D L H A M T S (SEQ ID No: 45)

5' GGC AAA CTG GAT TTG CAT GCC ATG ACA TCA 3' 5281.RD

(SEQ ID No: 46)

S L E M K F * (SEQ ID No: 47)

15 5' AGT CTT GAA ATG AAG TTT TAA 3'

(SEQ ID No: 48)

3' TCA GAA CTT TAC TTC AAA ATT GCC CTA GGG C 5' 5282.RD

BamH I (SEQ ID No: 49)

20 For the Q8 subclone, plasmid pLDW3, prepared as described in Example 5, was digested with BstE II and Sph I generating a 2.3kb fragment of *lbpA* which was ligated with the Nde I-BstE II and SphI-BamH I PCR fragments and cloned into pT7-7 digested with NdeI and
25 BamH I. The resulting plasmid pQW1A thus contains the full-length Q8 *lbpA* gene from the first methionine, under the control of the T7 promoter. DNA from pQW1A was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate strain
30 QW1A which was grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 protein was visualized by Coomassie blue staining (Fig. 11).

35 For the 4223 subclone, plasmid pLD3, prepared as described in Example 5 was digested with BstEII and SphI, generating a 2.3 kb fragment of *lbpA*, which was

ligated with the Nde I-BstE II and SphI-BamH I PCR fragments and cloned into pT7-7 digested with NdeI and BamH I. The resulting plasmid pRD1A thus contains the full-length 4223 *lbpA* gene from the first possible methionine under the control of the T7 promoter. DNA from pRD1A was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate strain RD1A which was grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 protein was visualized by Coomassie blue staining (Fig. 11).

The Q8 Lbp1 protein was expressed at very high levels but the 4223 Lbp1 protein was expressed at substantially lower levels.

15 **Example 8**

This Example illustrates the extraction and purification of rLbp1 from *E. coli*. The procedure is illustrated generally in Figure 14.

E. coli cells from a 500 ml culture, prepared as described in Example 7, were resuspended in 40 ml of 50 mM Tris-HCl, pH 8.0 containing 5 mM AEBSF (protease inhibitor) and 0.1 M NaCl, and disrupted by sonication (3 x 10 min, 70% duty cycle). The extract was centrifuged at 20,000 x *g* for 30 min and the resultant supernatant, which contained greater than 95% of the soluble proteins from *E. coli*, was discarded. The remaining pellet (Figure 14, PPT1) was further extracted in 40 ml of 50 mM Tris, pH 8.0 containing 0.5% Triton X-100 and 10 mM EDTA. The mixture was stirred at 4°C for at least 1 hour and then centrifuged at 20,000 x *g* for 30 min and the supernatant containing residual soluble proteins and the majority of the membrane proteins was discarded. The resultant pellet (Figure 14, PPT2) was further extracted in 40 ml of 50

mM Tris, pH 8.0 containing 1% octylglucoside. The mixture was stirred at 4°C for at least 1 hour and then centrifuged at 20,000 x g for 30 min. The supernatant containing residual contaminating proteins was discarded. The resultant pellet (Figure 14, PPT3) obtained after the above extractions contained the Lbp1 protein as inclusion bodies.

The rLbp1 protein was solubilized from the inclusion bodies in 50 mM Tris, pH 8.0, containing 6 M guanidine and 5 mM DTT. After centrifugation, the resultant supernatant was further purified on a Superdex 200 gel filtration column equilibrated in 50 mM Tris-HCl, pH 8.0, containing 2 M guanidine and 5 mM DTT. The fractions were analysed by SDS-PAGE and those containing purified rLbp1 were pooled. Triton X-100 was added to the pooled rLbp1 fraction to a final concentration of 0.1%. The fraction was dialysed overnight at 4°C against PBS, and then centrifuged at 20,000 x g for 30 min. The purified rLbp1 was stored at -20°C. Samples from the purification were analyzed by SDS-PAGE (Fig. 15).

Example 9

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp1 from the second methionine in *E. coli*.

The construction scheme for 4223 or Q8 *lbpA* expressed from the second methionine is shown in Figure 10. An approximately 200 bp fragment of the 5'-end of *lbpA* from the ATG to a BstE II site was PCR amplified using primers 5406.RD and 5407.RD. An Nde I site was engineered at the 5'-end to facilitate cloning into the pT7-7 vector.

NdeI

M T T H R L

(SEQ ID No: 50)

47

5' GGAATTCCCAT ATG ACC ACG CAC CGC TTA AA 3' 5406.RD

(SEQ ID No: 51)

BstE II

5 L D A I T V T A A

5' T TTA GAT GCC ATC ACG GTA ACC GCC GCC CC 3'

3' A AAT CTA CGG TAG TGC CAT TGG CGG CGG GG 5' 5407.RD

The 3'-end of the *lbpA* gene was PCR amplified from the SphI restriction site to the stop codon using primers 5281.RD and 5282.RD as described in Example 8. The 2.3 kb BstE II-Sph I fragments described in Example 8 were ligated to the Nde I-BstE II and Sph I-BamH I PCR fragments and cloned into pT7-7 that had been digested with NdeI and BamH I. Plasmid pQW1B thus contains a full-length Q8 *lbpA* gene from the second methionine and plasmid pRD1B contains a full-length 4223 *lbpA* gene from the second methionine under the direction of the T7 promoter. DNA was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate recombinant strains which were grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 proteins were visible by Coomassie blue staining (Fig. 11).

As seen for the longer protein in Example 8, the shorter Lbp1 from Q8 was expressed to much higher levels than the corresponding 4223 protein.

Example 10

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp2 with a leader sequence from *E. coli*.

The construction scheme is illustrated in Figure 12. There are two BspH I sites within the *lbpB* genes of strains 4223 and Q8. The 5'-end of the *lbpB* gene was PCR amplified from the ATG start codon through the

first BspH I site generating an approximately 201 bp fragment. An NdeI site was engineered at the ATG to facilitate cloning into the pT7-7 expression vector. The oligonucleotides used for amplification are illustrated below:

NdeI

M S T V K T P H (SEQ ID No: 52)

5' GGAATTCCAT ATG AGT ACT GTC AAA ACC CCC CAC A 3' 5533.RD

(SEQ ID No: 53)

10

BspH I

I P N T G H D N T N (SEQ ID No: 54)

5' A ATA CCG AAC ACA GGT CAT GAC AAC ACC AAT 3'

(SEQ ID No: 55)

T TAT GGC TTG TGT CCA GTA CTG TTG TGG TTA 5' 5534.RD

15

(SEQ ID No: 56)

The 3'-end of the *lbpB* gene was PCR amplified from the second BspH I site to the TAA stop codon generating a 381 bp fragment. A BamH I site was introduced after the stop codon for cloning purposes. The oligonucleotides used for amplification are illustrated below:

N E P T H E K T F A (SEQ ID No: 57)

5' AAT GAG CCT ACT CAT GAA AAA ACC TTT GCC 3' 5535.RD

25

(SEQ ID No: 58)

G A V F G A V K D K * (SEQ ID No: 59)

5' GG GCT GTC TTT GGG GCT GTT AAA GAT AAA TAA 3'

30

(SEQ ID No: 60)

CC CGA CAG AAA CCC CGA CAA TTT CTA TTT ATT CCTAGGGC 5' 5536.RD

BamH I (SEQ ID No: 61)

Plasmids pLD1-8 or pLDW1, prepared as described in Example 4, were digested with BspH I to release a 2.1 kb internal fragment of the *lbpB* gene which was ligated with the 5'- and 3'-PCR fragments and cloned into pT7-7

35

that had been digested with NdeI and BamH I. The resulting plasmids, pLD2A and pLDW2A, contain the full-length 4223 and Q8 *lbpB* genes under the control of the T7 promoter, respectively.

5 **Example 11**

This Example illustrates the construction of vectors to express the mature *M. catarrhalis* Lbp2 proteins from *E. coli*.

10 The construction scheme is illustrated in Figure 12. The putative mature Lbp2 lipoproteins start at the Cys³² residue. A scheme similar to that described in Example 10 can be used to generate expression clones. To amplify the 5'-end of the *lbpB* gene, a sense PCR primer is designed that includes an NdeI site for
15 subsequent cloning and an ATG start codon for initiation of translation followed immediately by the Cys³² residue. The antisense primer is the same as that described in Example 9 (5534.RD) and includes the BspH I cloning site. The amplified fragment is ~112 bp
20 long. The oligonucleotides are illustrated below:

NdeI

M C R S D D I S V N

(SEQ ID No: 62)

5' GGAATTCCAT ATG TGC CGC TCT GAT GAC ATC AGC GTC AAT 3' .RD

25 (SEQ ID No: 63)

BspH I

I P N T G H D N T N (SEQ ID No: 54)

5' A ATA CCG AAC ACA GGT CAT GAC AAC ACC AAT 3'

30 (SEQ ID No: 55)

3' T TAT GGC TTG TGT CCA GTA CTG TTG TGG TTA 5' 5534.RD

(SEQ ID No: 56)

35 The BspH I-BamH I 3'-end of the *lbpB* gene is PCR amplified as in Example 9 and the plasmid expressing

mature Lbp2 is constructed by ligating the 5'- and 3'-PCR fragments with the 2.1 kb BspH I fragment and vector pT7-7 digested with NdeI and BamH I. The resulting plasmids, pLD2B and pLDW2B, contain the *lbpB* gene encoding the mature Lbp2 proteins from strains 4223 and Q8 under the direction of the T7 promoter, respectively.

Example 12

This Example illustrates the construction of a vector to express the *M. catarrhalis* *lfr* Lbp3 from *E. coli*.

The construction scheme is illustrated in Figure 13. Oligonucleotides were used to generate the 5'-end of the *orf3* gene from the ATG start codon to an AlwN I site. An NdeI site was engineered at the 5'-end for subsequent cloning into pT7-7. The oligonucleotides are shown below:

```

NdeI
20      M   T   C   L   P   K   T   N   P   A   L   K   V   K   H   R
5'  T ATG ACC TGT TTA CCA AAG ACC AAC CCT GCT TTA AAA GTC AAG CAC AGA
3'      AC TGG ACA AAT GGT TTC TGG TTG GGA CGA AAT TTT CAG TTC GTG TCT

      AlwN I
          F   L   K   Q   V
25      TTT TTA AAG CAG GTG          3'      5532.RD      (SEQ ID No: 65)
          AAA AAT TTC GTC              5'      5457.RD      (SEQ ID No: 66)

```

The pLD1-8 or pLDW1 plasmid, prepared as described in Example 5, was digested with BstE II generating a 4.6 kb fragment which was filled in with Klenow polymerase before being digested with AlwNI. The resultant 1.8 kb fragment was ligated with the annealed NdeI-AlwN I oligonucleotides and cloned into pT7-7 that had been digested with NdeI and SmaI. The resulting plasmids, pLRD3 and pLQW3, contain the full-length *orf3*

genes from strains 4223 and Q8 under the direction of the T7 promoter, respectively.

Example 13

5 This Example describes the cloning and sequencing of the *lbpB* gene from *M. catarrhalis* strain VH19.

Chromosomal DNA was prepared from *M. catarrhalis* strain VH19, as described previously in Example 2. Oligonucleotide primers were designed based upon the flanking sequence of the 4223 *lbpB* gene. The sense
10 primer was 5' AAGCTTAGCATGATGGCATCGGCT 3' (SEQ ID No: 67) and the antisense primer was 5' TTAGCCCAAGGCAAATCTGGTGCA 3' (SEQ ID No: 68). PCR was performed in buffer containing 10mM Tris-HCl (pH 8.3), 50 mM potassium chloride and 1.5 mM magnesium chloride.
15 Each 100 µl reaction mixture contained 1 µg chromosomal DNA, 0.1 µg each primer, 2.5 units amplitaq DNA polymerase (Perkin Elmer Cetus, Foster City, California) and 10 mM of each dNTP (Perkin Elmer Cetus). The cycling conditions were 24 cycles of 94°C
20 for 1 min, 47°C for 30 sec and 72°C for 1 min. Specific 2.9 kb fragments were amplified from two independent reactions and subcloned into pCR II (Invitrogen, Carlsbad, California), generating plasmids pVH19pcr1 and pVH19pcr2 for sequence analysis. A third
25 PCR amplification was performed without subcloning the resultant DNA. Plasmid DNA from pVH19pcr1 and pVH19pcr2 was prepared from 50 ml overnight cultures using the Qiagen Plasmid Midi kit (Qiagen Inc, Chatsworth, California). PCR amplified DNA was
30 purified for direct sequencing using a Qiagen PCR purification kit. DNA samples were sequenced on an ABI model 373A DNA sequencer using dye terminator chemistry. Oligonucleotide primers 17 to 25 bases in length were used to sequence both strands of the DNA.

The nucleotide sequence (SEQ ID No: 69) of the VH19 *lbpB* gene and the deduced amino acid sequence of the corresponding Lbp2 protein (SEQ ID No: 70) are shown in Figure 16. The encoded VH19 Lbp2 protein is 906 amino acids and is 77% identical and 84% similar to the 4223 and Q8 Lbp2 proteins. There is a putative lipoprotein signal sequence which is very similar to the 4223 and Q8 signal sequences. The high Asp and Asn content found in the 4223 and Q8 Lbp2 proteins is also present in the VH19 LbpB protein, as is the RGD sequence. A partial restriction map of the VH19 *lbpB* gene is shown in Figure 17.

An alignment of the Lbp2 proteins from *M. catarrhalis* strains 4223, Q8 and VH19 is shown in Figure 7. The *M. catarrhalis* Lbp2 proteins are also compared with partial Lbp2 sequences from *N. meningitis* strains BNCV (ref. 31) and H44/76 (ref. 24) and *N. gonorrhoeae* strain FA19 (ref. 25). There are small scattered regions of sequence homology to the known bacterial Tbp2 proteins (ref. 32). Residues that are conserved among the Tbp2 proteins and the *M. catarrhalis* Lbp2 proteins are underlined in Figure 7 and include the LEGGFY (SEQ ID No: 75) motif.

Example 14

This Example describes the construction of vectors for expression of the *M. catarrhalis* Lbp2 protein.

By analogy with Tbp2 proteins, Lbp2 was assumed to be a lipoprotein and constructs were designed for expression of Lbp2 with or without a lipopeptide signal sequence. There is a unique Bgl I site in *lbpB*. To express the full-length Lbp2 protein with leader sequence (construct A), an approximately 429 bp 5'-fragment from the Met¹ start codon to the Bgl I site was PCR amplified and to express the mature protein

(construct B), an approximately 329 bp 5'-fragment from the putative Cys³² start to the Bgl I site was PCR amplified. The following sense primers were used:

5' Nde I
 5' M S T V K T P H (SEQ ID No: 52)
 5' GGAATTCCAT ATG AGT ACT GTC AAA ACC CCC CAC A 3' (SEQ ID No: 53)
 for construct A or

10' Nde I
 10' M C R S D D I S V N (SEQ ID No: 62)
 5' GGAATTCCAT ATG TGC CGC TCT GAT GAC ATC AGC GTC AAT 3' (SEQ ID No: 63)
 for construct B and the anti-sense primer was:

15' G K N L R G P I (SEQ ID No: 72)
 5' GGT AAA AAC TTG CGT CAG CCC ATC 3' (SEQ ID No: 73)
 3' CCA TTT TTG AAC GCA GTC GGG TAG 5' (SEQ ID No: 74)
 Bgl I

20' The Q8 *lfr*-containing plasmid, pLDW1 (Example 5), was digested with Bgl I and EcoR I to release a 2.3 kb *lbpB* fragment which was ligated with the Nde I - Bgl I PCR fragment and cloned into pT7-7 that had been digested with Nde I and EcoR I. The resulting plasmids, pQW2A and pQW2B, thus contain the Q8 *lbpB* gene encoding the full-length or mature Lbp2 proteins under the direction of the T7 promoter. The plasmids expressing the 4223 full-length or mature Lbp2 proteins were constructed in a similar manner and designated pRD2A and pRD2B. There was no measurable expression of rLbp2 from constructs containing the signal sequence, however the mature rLbp2 proteins were expressed at 5 to 10% of total proteins as inclusion bodies and were purified by the same process as that described for rLbp1 in Example 8. Samples from the purification were analyzed by SDS-PAGE (Figure 18).

Example 15

This Example describes the functional characterization of the recombinant lactoferrin binding proteins.

5 Human lactoferrin (Sigma) was conjugated to horseradish peroxidase using an EZ-Link maleimide activated horseradish peroxidase (HRP) kit (Pierce, Rockford, Illinois) according to the manufacturer's instructions. The lactoferrin binding activity of
10 rLbp1 or rLbp2 was assessed by modifying the procedure described for transferrin binding proteins (ref. 17). Briefly, purified rLbp1 or rLbp2 was subjected to discontinuous electrophoresis through a 12.5% SDS PAGE gel. The proteins were electrophoretically transferred
15 to a polyvinylidene difluoride (PVDF) membrane (Millipore, Bedford, Massachusetts) and incubated with horseradish peroxidase-conjugated human lactoferrin (1:20 dilution) at 4°C overnight. LumiGLO substrate (Kirkegaard and Perry Laboratories, Inc., Gaithersburg,
20 Maryland) was used for chemiluminescent detection of HRP activity according to the manufacturer's instructions. The Q8 rLbp1 protein did not bind human lactoferrin under these conditions, but the 4223 rLbp2 and Q8 rLbp2 proteins did (Fig. 19).

25 Example 16

This Example describes the immunization of animals and immunoassays.

Groups of two guinea pigs (Hartley outbred, Charles River, Quebec) were immunized intramuscularly (i.m.)
30 with 5 µg doses of purified rLbp1 or rLbp2 protein emulsified in CFA or IFA. Anti-Lbp antibody titers in guinea pig immune sera were determined by antigen-specific ELISA. Microtiter wells (Nunc-MAXISORB, Nunc, Denmark) were coated with 50 µl of protein (0.5 µg ml⁻¹

¹). The reactive titer of an antiserum was defined as the reciprocal of the highest dilution consistently showing a two-fold increase in absorbance at 450 nm over that obtained with the pre-immune serum samples. The recombinant proteins elicited high titre antibodies as shown in Tables 1 and 2.

Example 17

This Example describes the antigenic conservation of Lbp1 and Lbp2 in *M. catarrhalis* strains.

To demonstrate the iron-dependent expression of the *lbpA* and *lbpB* genes, representative *M. catarrhalis* strains were grown in BHI ± 25 mM EDDA. Whole cell lysates were separated by SDS PAGE and electrophoretically transferred to nitrocellulose membrane. Guinea pig anti-Q8 rLbp1, anti-Q8 rLbp2 and anti-4223 rLbp2 antisera were used as first antibodies and horseradish peroxidase-conjugated protein G (ZYMED) was used as secondary antibody. To assess antigenic conservation, approximately 90 *M. catarrhalis* strains, obtained from North America or Finland were grown in BHI + 25 mM EDDA, and immunoblots were probed with guinea pig anti-4223 rLbp2 antibody, as above. All strains showed a protein band reactive with anti-rLbp2 antibody. There was very little size heterogeneity for the Lbp2 proteins from the 90 *M. catarrhalis* strains, ranging from approximately 100 kDa to 105 kDa. Representative immunoblots are illustrated in Fig. 19.

Example 18

This Example describes the assay used to determine the bactericidal antibody activity of anti-Lbp antibodies.

The assay was performed as described by ref. 33. Briefly, the *M. catarrhalis* strains were grown to an OD₅₇₈ of 0.5 in BHI medium containing 25 mM EDDA. The

bacteria were diluted so that the pre-bleed control plates contained 100 to 300 cfu. Guinea pig anti-rLbp1 or anti-rLbp2 antisera and pre-bleed controls, were heated to 56°C for 30 min to inactivate endogenous complement and were diluted 1:64 with veronal buffer containing 0.1% BSA (VBS). Guinea pig complement (Biowhittaker, Walkersville, Maryland) was diluted 1:10 in VBS. Twenty-five µl each of diluted antiserum, bacteria and complement were added to duplicate wells of a 96 well microtiter plate (Nunc). The plates were incubated at 37°C for 60 min, gently shaking at 70 rpm on a rotary platform. Fifty µl of each reaction mixture were plated onto Mueller Hinton agar plates (Becton-Dickinson, Cockeysville, Maryland) which were incubated at 37°C for 24 h, then room temperature for 24 h, before the bacteria were counted. Antisera were determined to be bactericidal if $\geq 50\%$ of bacteria were killed compared with negative controls.

Six strains of different geographical and anatomical origins were tested. The data in Table 3 illustrates that anti-4223 rLbp2 antibody was bactericidal for the homologous strain and three of five heterologous strains.

SUMMARY OF THE DISCLOSURE

In summary of this disclosure, the present invention provides purified and isolated DNA molecules containing lactoferrin receptor genes from *Moraxella catarrhalis*, the sequences of these lactoferrin receptor genes, and the derived amino acid sequences thereof. The genes and DNA sequences are useful for diagnosis, immunization, and the generation of diagnostic and immunological reagents. Immunogenic compositions, including vaccines, based upon expressed recombinant Lbp1 and/or Lbp2 and/or ORF3, portions

thereof, or analogs thereof, can be prepared for prevention of diseases caused by *Moraxella*. Modifications are possible within the scope of this invention.

Table 1

Bactericidal antibody titres for anti-native Lbp1

Antibody	Bactericidal titre - RH408		Bactericidal titre - Q8	
	Pre-immune	Immune	Pre-immune	Immune
Anti-4223 Lbp1	<8	114-330	<8	128-512

Bactericidal titres are expressed as the reciprocal dilution of antiserum capable of killing 50% of *M. catarrhalis* cells

Table 2 ELISA titers for guinea pig anti-Lbp antibodies raised against recombinant lactoferrin binding proteins

Coated antigen	Anti-Q8 rLbp1	Anti-Q8 rLbp2	Anti-4223 rLbp2
Q8 rLbp1	3,200 25,600	-	-
Q8 rLbp2	-	1,638,400 1,638,400	409,600 409,600
4223 rLbp2	-	409,600 409,600	819,200 819,200

Table 3. Bactericidal antibody activity of guinea pig anti-rLbp2 antibodies

Strain	locale ¹	source ²	Lbp2 size	Bactericidal antibody activity ³	
				Anti-4223 rLbp2	Anti-Q8 rLbp2
4223	New York	MEF	105 kDa	++	-
Q8	Quebec	sputum	105 kDa	±	-
VH19	Texas	MEF	105 kDa	+	NT ⁴
LES-1	Finland	MEF	102 kDa	-	NT
H-04	Nova Scotia	MEF	100 kDa	+	NT
3	New York	sputum	100 kDa	++	NT

¹ geographic locale where strain was isolated

² anatomical source of clinical isolate. MEF is middle ear fluid from otitis media patients

³ killing by antiserum diluted 1:64, compared to negative controls: - indicates 0-25% killing; ± indicates 26-49% killing; + indicates 50-75% killing; ++ indicates 76-100% killing.

⁴ NT = not tested

REFERENCES

1. Brorson, J-E., A. Axelsson, and S.E. Holm. 1976. Studies on *Branhamella catarrhalis* (*Neisseria catarrhalis*) with special reference to maxillary sinusitis. *Scan. J. Infect. Dis.* 8:151-155.
2. Catlin, B.W., 1990. *Branhamella catarrhalis*: an organism gaining respect as a pathogen. *Clin. Microbiol. Rev.* 3: 293-320.
3. Hager, H., A. Verghese, S. Alvarez, and S.L. Berk. 1987. *Branhamella catarrhalis* respiratory infections. *Rev. Infect. Dis.* 9:1140-1149.
4. McLeod, D.T., F. Ahmad, M.J. Croughan, and M.A. Calder. 1986. Bronchopulmonary infection due to *M. catarrhalis*. Clinical features and therapeutic response. *Drugs* 31(Suppl.3):109-112.
5. Nicotra, B., M. Rivera, J.I. Luman, and R.J. Wallace. 1986. *Branhamella catarrhalis* as a lower respiratory tract pathogen in patients with chronic lung disease. *Arch.Intern.Med.* 146:890-893.
6. Ninane, G., J. Joly, and M. Kraytman. 1978. Bronchopulmonary infection due to *Branhamella catarrhalis* 11 cases assessed by transtracheal puncture. *Br.Med.Jr.* 1:276-278.
7. Srinivasan, G., M.J. Raff, W.C. Templeton, S.J. Givens, R.C. Graves, and J.C. Mel. 1981. *Branhamella catarrhalis* pneumonia. Report of two cases and review of the literature. *Am. Rev. Respir. Dis.* 123:553-555.
8. West, M., S.L. Berk, and J.K. Smith. 1982. *Branhamella catarrhalis* pneumonia., *South. Med. J.* 75:1021-1023.
9. Christensen, J.J., and B. Bruun. 1985. Bacteremia caused by a beta-lactamase producing strain of *Branhamella catarrhalis*. *Acta. Pathol. Microbiol. Immunol. Scand. Sect. B* 93:273-275.
10. Craig, D.B., and P.A. Wehrle. 1983. *Branhamella catarrhalis* septic arthritis. *J. Rheumatol.* 10:985-986.
11. Guthrie, R., K. Bakenhaster, R. Nelson, and R. Woskobnick. 1988. *Branhamella catarrhalis* sepsis: a case report and review of the literature. *J. Infect. Dis.* 158:907-908.

12. Hiroshi, S., E.J. Anaissie, N. Khardori, and G.P. Bodey. 1988. *Branhamella catarrhalis* septicemia in patients with leukemia. *Cancer* 61:2315-2317.
13. O'Neill, J.H., and P.W. Mathieson. 1987. Meningitis due to *Branhamella catarrhalis*. *Aust. N.Z. J. Med.* 17:241-242.
14. Murphy, T.F. 1989. The surface of *Branhamella catarrhalis*: a systematic approach to the surface antigens of an emerging pathogen. *Pediatr. Infect. Dis. J.* 8:S75-S77.
15. Van Hare, G.F., P.A. Shurin, C.D. Marchant, N.A. Cartelli, C.E. Johnson, D. Fulton, S. Carlin, and C.H. Kim. Acute otitis media caused by *Branhamella catarrhalis*: biology and therapy. *Rev. Infect. Dis.* 9:16-27.
16. Jorgensen, J.H., Doern, G.V., Maher, L.A., Howell, A.W., and Redding, J.S., 1990. Antimicrobial resistance among respiratory isolates of *Haemophilus influenza*, *Moraxella catarrhalis*, and *Streptococcus pneumoniae* in the United States. *Antibicrob. Agents Chemother.* 34: 2075-2080.
17. Schryvers, A.B. and Lee, B.C. (1988) Comparative analysis of the transferrin and lactoferrin binding proteins in the family *Neisseriaceae*. *Can. J. Microbiol.* 35, 409-415.
18. O'Hagan, DT. 1992. Oral delivery of vaccines. Formulation and clinical pharmacokinetic considerations. *Clin. Pharmacokinet* 22(t): 1-10.
19. Ulmer et al. 1993. *Curr. Opin. Invest. Drugs* 2:983-989.
20. Lockhoff, O., 1991. Glycolipids as immunomodulators: Synthesis and properties.
21. Nixon-George A., et al., 1990. The adjuvant effect of stearyl tyrosine on a recombinant subunit hepatitis B surface antigen. *J Immunol* 144 (12): 4798-4802.
22. Wallace, R.J. et al., 1990. Antibiotic susceptibilities and drug resistance in *Moraxella* (*Branhamella*) *catarrhalis*. *Am. J. Med.* 88(5A): 465-505.
23. Nissinen A, et al., 1995. Development of beta-lactamase-mediated resistance to penicillin in middle-ear isolates of *Moraxella catarrhalis* in

- Finnish children, 1978-1993. Clin Infect Dis 21 (5): 1193-1196.
24. Pettersson, A., et al., 1994. Identification of iroa Gene Product of *Neisseria meningitidis* as a Lactoferrin Receptor. J. Bacteriol. 176(6): 1764-1766.
 25. Biswas GD, Sparring PF. 1995. Characterization of lbpa, the structural gene for a lactoferrin receptor in *Neisseria gonorrhoeae*. Infect Immun 63 (8): 2958-2967.
 26. Legrain M, et al. 1993. Cloning and characterization of *Neisseria meningitidis* genes encoding the transferrin-binding proteins Tbp1 and Tbp2. Gene 130 (1): 73-80.
 27. Cornelissen CN, Biswas GD, Sparling PF. 1993. Expression of gonococcal transferrin-binding protein 1 causes *Escherichia coli* to bind human transferrin. J Bacteriol 175 (8): 2448-2450.
 28. Anderson JE, Sparling PF, Cornelissen CN. 1994. Gonococcal transferrin-binding protein 2 facilitates but is not essential for transferrin utilization. J Bacteriol 176 (11): 3162-3170.
 29. Ogunnariwo JA, Schryvers AB. 1996. Rapid identification and cloning of bacterial transferrin and lactoferrin receptor protein genes. J Bacteriol 178 (24): 7326-7328.
 30. Loosmore SM, et al. 1996. Cloning and expression of the *Haemophilus influenzae* transferrin receptor genes. Mol Microbiol 19 (3): 575-586.
 31. Pettersson, A. et al. 1993. Molecular Characterization of the 98-Kilodalton Iron-Regulated Outer membrane Protein of *Neisseria meningitidis*. Infect. Immun. 61 (ti): 4724-4733.
 32. Ogunnariwo, J.A., Woo, T.K.W., Lo, R.Y.C., Gonzalez, G.C., and Schryvers, A.B. (1997) Characterization of the *Pasteurella haemolytica* transferrin receptor genes and the recombinant receptor proteins. Microbial Pathog **23**:273-284.
 33. Yang, Y.P., Myers, L.E., McGuinness, U., Chong, P., Kwok, Y., Klein, M.H., and Harkness, R.E. (1997) The outer membrane protein, CD, extracted from *Moraxella (Branhamella) catarrhalis* is a potential vaccine antigen that induces

bactericidal antibodies. *FEMS Immun Med Microbiol* **17**:187-199.

34. Pettersson, A., Klarenbeek, V., van Deurzen, J., Poolman, J.T., and Tommassen, J. (1994a) Molecular characterization of the structural gene for the lactoferrin receptor of the meningococcal strain H44/76. *Microb Pathog* **17**:395-408.
35. Needleman, S.B., and Wunsch, C.D. 1970, *J. Mol Biol.* **48**:443-453.
36. Sellers, P.H. 1974 On the theory and computation of evolutionary distances. *J. Appl. Math(Siam)* **26**:787-793.
37. Waterman, M.S., Smith, T.F., and Beyer, W.A. 1976. *Advan. Math.* **20**:367-387.
38. Smith, T.F., and Waterman, M.S. 1981 Identification of common molecular subsequences. *J. Mol. Biol.* **147**:195-197.
39. Sobel, E. and Martinez, H.M. 1985 A Multiple Sequence Alignment Program. *Nucleic Acid Res.* **14**:363-374.
40. Bonnat, R.A., Yu, R.H. and Schryvers, A.B. 1995, Biochemical Analysis of Lactoferrin Receptors in the *Neisseriaceae*: Identification of a Second Lactoferrin Receptor Protein. *Microb. Pathog.* **19**:285-297.

CLAIMS

What we claim is:

1. A purified and isolated nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein.
2. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the lactoferrin receptor binding protein 1 (Lbp1) of the *Moraxella* strain.
3. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the lactoferrin receptor binding protein 2 (Lbp2) of the *Moraxella* strain.
4. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the open reading frame protein 3 (ORF3) of the *Moraxella* strain.
5. The nucleic acid molecule of claim 1 wherein the strain of *Moraxella* is a strain of *Moraxella catarrhalis*.
6. The nucleic acid molecule of claim 5 wherein the strain of *Moraxella catarrhalis* is *Moraxella catarrhalis* 4223, Q8 or VH16.
7. A purified and isolated nucleic acid molecule encoding at least one lactoferrin binding protein of *Moraxella* and having a restriction map as shown in Figure 3 for *M. catarrhalis* 4223, Figure 5 for *M. catarrhalis* Q8 or Figure 17 for *M. Catarrhalis* VH19 or the equivalent map for another strain of *M. catarrhalis*.
8. A purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of:

- (a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto;
 - (b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and
 - (c) a DNA sequence encoding a functional lactoferrin receptor protein of *Moraxella*.
9. The nucleic acid molecule of claim 8, wherein the DNA sequence defined in (c) hybridizes under stringent conditions to any one of the sequences defined in (a) or (b).
10. The nucleic acid molecule of claim 8, wherein the DNA sequence defined in (c) is that encoding the equivalent lactoferrin receptor protein from another strain of *Moraxella*.
11. A vector adapted for transformation of a host comprising the nucleic acid molecule of claim 1 or 8.
12. The vector of claim 11 encoding at least a fragment of a lactoferrin receptor protein and having the characteristics of a plasmid selected from the group consisting of pLD3, pLDW3, pLD1-8 (ATCC 97,997), pLDW1 (ATCC 97,998), pVH19pcr1 and pVH19pcr2.
13. The vector of claim 11 further comprising expression means operatively coupled to the nucleic acid molecule for expression of said lactoferrin receptor protein of a strain of *Moraxella* or the fragment or the analog of the lactoferrin receptor protein by the host containing the vector.
14. The vector of claim 13 having the characteristics of plasmid pRD1A, pRD1B, pQW1A, pQW1B, pRD2B, pQW2B, pLRD3 and pLQW3.

15. A transformed host containing an expression vector as claimed in claim 13.

16. A method of forming a substantially pure recombinant lactoferrin receptor protein, which comprises:

growing the transformed host of claim 15 to express a lactoferrin receptor protein as inclusion bodies,

purifying the inclusion bodies free from cellular material and soluble proteins,

solubilizing lactoferrin receptor protein from the purified inclusion bodies, and

purifying the lactoferrin receptor protein free from other solubilized materials.

17. The method of claim 16 wherein said lactoferrin receptor protein comprises Lbp1 alone, Lbp2 alone, ORF3 alone or a mixture of two or more of Lbp1, Lbp2 and ORF3.

18. The method of claim 17 wherein said lactoferrin receptor protein is at least about 70% pure.

19. The method of claim 18 wherein said lactoferrin receptor protein is at least about 90% pure.

20. A recombinant lactoferrin receptor protein or fragment or analog thereof producible by the transformed host of claim 14.

21. The protein of claim 20 which is lactoferrin receptor binding protein 1 (Lbp1) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

22. The protein of claim 20 which is lactoferrin receptor binding protein 2 (Lbp2) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

23. The protein of claim 20 which is open reading frame protein 3 (ORF3) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

24. The protein of claim 20 wherein the strain of *Moraxella* is a strain of *Moraxella catarrhalis*.

25. An open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the lactoferrin binding protein.

26. The protein of claim 25 wherein said *Moraxella* strain is a strain of *M. catarrhalis*.

27. The protein of claim 26 wherein said *M. catarrhalis* strain is *M. catarrhalis* 4223 or Q8.

28. The protein of claim 24 having a deduced amino acid as set forth in Figure 2 or 4 (SEQ ID No: 14, 18).

29. An immunogenic composition, comprising at least one active component selected from the group consisting of:

(A) a purified and isolated nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of lactoferrin receptor protein;

(B) a purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of:

(a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto;

(b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and

(c) a DNA sequence encoding a functional lactoferrin receptor protein of *Moraxella*;

(C) a recombinant lactoferrin receptor protein or fragment or analog thereof producible by a transformed host containing an expression vector comprising a nucleic acid molecule as defined in (A) or (B) and expression means operatively coupled to the nucleic acid molecule for expression by the host of the recombinant lactoferrin receptor protein or fragment or analog thereof; or

(D) an open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the open reading frame protein;

and a pharmaceutically acceptable carrier therefor, said at least one active component producing an immune response when administered to a host.

30. A method for generating an immune response in a host, comprising administering to the host an immunoeffective amount of the immunogenic composition of claim 29.

31. A method of determining the presence, in a sample, of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising the steps of:

(a) contacting the sample with the nucleic acid of claim 1 or 7 to produce duplexes comprising the nucleic acid molecule and any nucleic acid molecule encoding the lactoferrin receptor protein of a strain of *Moraxella* present in the sample and specifically hybridizable therewith; and

(b) determining production of the duplexes.

32. A diagnostic kit for determining the presence, in a sample of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising:

(a) the nucleic acid molecule of claim 1 or 7;

(b) means for contacting the nucleic acid molecule with the sample to produce duplexes comprising the

nucleic acid molecule and any said nucleic acid present in the sample and hybridizable with the nucleic acid molecule; and

(c) means for determining production of the duplexes.

FIG.1A

Alignment of translated 2.2kb *lbpA* PCR fragments

MNQSKQNNKSKKQVLKLSALSLGLLNITQVALANTTADK	Tbp1
MSKSITKTQTSPSVHTMTTHRLNLAIKAAALFGVAVLPLSVWAQENTQTDAN	Lbp1
AEATDKTNLVVVLDETVVTAKKNAPVSRKANEVGTGLGKVVKTAETINKEQ	Tbp1
SDAKDTKTPVVYLDAITVTAAPSA--RFDTDVTGLGKTVKKTADTLAKEQ	Lbp1
VLNIRDLTRYDPGIAVVEQGRGASSGYSIRGMDKNRVAVLVDGINQAQHY	Tbp1
VQGI RDLVRYETGVSVVEQGRGSSGFAIHGV DKNRVGITVDGIAQIQSY	Lbp1
--QGPVAGKNYAAGGAIN EIEYENVRVSVEISKGANSSEYSGALSGSVAFVT	Tbp1
ALKDESTKRAGAGSGAMNEIEIENIAAVAINKGCNALEAGSGALGGSVAFHT	Lbp1
KTADDIIKDGKDWGVQTKTAYASKNNAWVNSVAAAGKAGSFGLIIYTDR	Tbp1
KDVSDVLKSGKNLGAQSKTTYN SKNDHFSQTLAAAGKTERVEAMVOYTYR	Lbp1
QYT-R	PCR4
QYT-R	PCR5
RGQ EYKAHDDAYQGSQSFDRAVA TDPNNRTFLIANECANGNYEACAAGG	Tbp1
<u>KGKENKAHSD</u> LNGINQSLYRLGAWQQKYDLRKPNELFAGTSYITESCLAS	Lbp1
KG-ENKAHSDLNGINQSLYRLGAWQQKYDLRKPNELFAGTSYITESCLAS	PCR4
KG-ENKAHSDLNGINQSLYRLGAWQQKYDLRKPNELFAGTSYITESCLAS	PCR5
QTKLQAKPTNVRDKVNVKDYTGPNRLIPNPLTQDSKSLLLRPGYQLNDKH	Tbp1
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQYLASTHPHEVVS AK	Lbp1
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQYLASTHPHEVVS AK	PCR4
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQYLASTHPHEVVS AK	PCR5

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FIG. 1B

YVGGVYEITKQNYAMQDKTVPAYLAVHDIEKSRLSNHAQANGYYQNNLGERIRD	Tbp1
DYTGIRLLPDPMDYRSDSYLARLNKIKITPNLVSKLLEDTKQTYNIRDM	Lbp1
DYTGIRLLPDPMDYRSDSYLARLNKIKITPNLVXKLLLEDTKQTYNIRDM	PCR4
DYTGIRLLPDPMDYRSDSYLARLNKIKITPNLVSKLLEDTKQTYNIRDM	PCR5
IGPDSGYGINYAHGVFYDEKHQKDRGLGLEYVYDSKGENKWFDDVRVSYDKQDIT	Tbp1
RHCSYHGARLGNDGKPPANGGSI VLCDDYQEYLNANDASQALFRPGANDAP	Lbp1
RHCSYHGARLGNDGKPPANGGSI VLCDDYQEYLNANDASQALFRPGANDAP	PCR4
RHCSYHGARLGNDGKPPANGGSI VLCDDYQEYLNANDASQASFRPGANDAP	PCR5
LRSQLTNTHCSTYPHIDKNCTPDVNKPFVSKEVDNNAYKEQHNLKAVFN	Tbp1
IPKLAYARSSVFNQEHGKTRYGLSFEFKPDTPWFKQAKLNLHQQNIQIIN	Lbp1
IPKLAYARSSVFNQEHGKTRYGLSFEFKPDTPWFKQAKLNLHQQNIQIIN	PCR4
IPKLAYARSSVFNQEHGKTRYGLSFEFKPDTPWFKQAKLNLHQQNIQIIN	PCR5
KKMALGSTHHHINLQVGYDKFNSSLSRVEYRLATHQSYQKLDYTPPSNPL	Tbp1
HDIKKSCSQYPKVDLNCGIS EIGHYEQNNYRYKEGRASLTGKLDNFNFDL	Lbp1
HDIKKSCSQYPKVDNSNCGIS EIGHYEQXNYRYKEGRASLTGKLDNFNFDL	PCR4
PDKFKPIILGSNNKPICLDAYGYGHDHPQACNAKNSTYQNF A I KKGIEQYN	Tbp1
LGQHDLT VLAGADKVKSQFRANNPRRTI IDTTQGD A I I DESTLTAQEQA K	Lbp1
LGQHDLT VLAGTDKVKSQFRANNPRRTI IDTTQGD A I I DESTLTAQEQA K	PCR4

FIG.1C

QKNTNIDYQAIIDQYDKQNPNSTLKPFEEKIKQSLGQEKYNKIDELGFK	Tbp1
FKQSGAAWIVKNRLGRLEEKDACGNANECERAPIHGSNQYVGINNLYTPN	Lbp1
FKQSGAAWIVKNRLGRLEEKDACGNANECERAPIHGSNQYVGINNLYTPN	PCR4
AYKDLRNEWAGWTNDNSQQNANKGTDNIYQPNQATVVKDDKCKYSETNSY	Tbp1
DYVDLSFGGRLDKQRIHSTDNSIIISKYTNKSYNFGAAVHLTPDFSLLYK	Lbp1
DYVDXSFGRLDKQRIHSTDNSIIISKYTNKSYNFGAAVHLTPDFSLLYK	PCR4
TDSNIIISKYTNKSYNFGAAVHXTPDFSLLYK	PCR5
ADCSTTRHISGDNFYIALKDNMTINKYVDLGLGARYDRIKHKSDVPLVDNSASNQLSWNFGVV	Tbp1
TAKGFRTPSFYELYNYNSTAAQHKNDPDVSFPKRAVDVKPETSNTNEYGF	Lbp1
TAKGFRTPSFYELYNYNSTAAQHKNDPDVSFPKRAVDVKPETSNTNEYGF	PCR4
TAKGFRTPSFYELYNYNSTAAQHKNDPDVSFPKRAVDVKPETSNTNEYGF	PCR5
VKPTNWLDIAYRSSQGFRMPSESEMYGERFGVTIGKGTQHGCKGLYYICQQTV	Tbp1
RYQHPWGDVEMSMFKSRYKMDMLDKAIPNLTKAQQEQYCKAHLDSNECVGNP	Lbp1
RYQHPWGDVEMSMFKSRYKMDMLDKAIPNLTKAQQEQYCKAHLDSNECVGNP	PCR4
RYQHPWGDVEMSMFKSRYKMDMLDKAIPNLTKAQQEQYCKAHLDSNECVGNP	PCR5
HQTKLKPEKSFNQEIIGATLHNHLGSLEVSFYKNRYTDLIVGKSEEIRTLT	Tbp1
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK	Lbp1
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK	PCR4
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK	PCR5
QGDNAGKQRGKGLGFHNGQDADLTGINILGRLDLNAANSRLPYGLYSTL	Tbp1

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FIG.1D

AYNKVDVKGKTLNPTLAGTNILFDAIQPSRYVVGGLGYDAPSQKWGANAI	Tbp1
LGKFDYIAPKDADGWYQARPAFWDAITPARYVVGGLNYDHPDPSQVWGIGTTL	Lbp1
LGKFXYIAPKDADGWYQARPAFWDAITPARYVVGGLNYDHPDPSQVWGIGATL	PCR4
LGKFXYIAPKDADGWYQARPAFWDAITPARYVVGGLNYDHPDPSQVWGIGTTL	PCR5
HSDAKNPSELLADKNLGNNGNIQTKQATKAKSTPWQTL-DLSGYVNIKDNFT	Tbp1
THSKQKDENELSALRIR-NGKRETQTLTHTI PKAYTLLDMTGYYSPTESIT	Lbp1
THSKQKDENELSALRIR-NGKRETQTLTHTI PKAYTLLDMTGYYSPTESIT	PCR4
THSKQKDENELSALRIR-NGKREIQTLTHTI PKAYTLLDMTGYYSPTESIT	PCR5
LRAGVYNVFNYYTTWEALRQTAKGAVNQHTGLSQDKHYGRYAAPGRNYQLALEMKF*	Tbp1
ARLGINNVLNTRYTTWEAARQ-----LPSEAASTQSTRYIAPGRSYFASLEMKF*	Lbp1
ARLGINNVLNTRYTTWEAARQ-----LPSEAASTQSTRYIAPGRSYFASLEMKF*	PCR4
ARLGINNVLNTRYTTWEAARQ-----LPSEAASTQSTRYIAPGRSYFASLEMKF*	PCR5

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FIG.2A

M. catarrhalis 4223 *lfr* sequence

```

AAGCTTAGCATGATGGCATCGGCTGATTGT      10
                                20      30
CTTTTGGCCCTTGTGTGTGTGTGTGGGAGT      40      50      60

-35
TGATTGTACTTACCTTAGTGGTGGATGCTT      70      80      90
                                -10
GGGCTGATTTTAATAAAGCGGCTCTCAACAAC      100      110      120
                                5/130

RBS                               Lbp2
ACACCAACGAGATATCACCATGAGTACTG      130      140      150
VAL  LYS  THR  PRO  HIS  ILE  PHE  TYR  GLN  LYS  A
TCAAAACCCCCACATTTCTTACCACAAAC      160      170      180

RG  THR  LEU  SER  LEU  ALA  ILE  ALA  SER  ILE
GCACCTTAGCCCTTGCCATCGCCAGTATTT      190      200      210
PHE  ALA  ALA  LEU  VAL  MET  THR  GLY  CYS  ARG  S
TTGCTGCCCTTGGTGATGACAGGCTGCCCT      220      230      240

```

FIG.2B

ER ASP ILE SER VAL ASN ALA PRO ASN
 CTGATGACATCAGCGTCAATGCAACCAATG
 250 260 270
 VAL THR GLN LEU PRO GLN GLY THR VAL SER P
 TTAACCAACTGCCCCCAAGGCACGGTTTCAC
 280 290 300

RO ILE PRO ASN THR GLY HIS ASP ASN THR
 CAATACCGAACACAGGTCAATGACAAACCA
 310 320 330
 ASN ASN THR ASN ASN GLN GLY ASN ASN THR A
 ATAACCAACAATACTAGGGCAACAACACGG
 340 350 360

SP ASN SER THR SER THR THR ASP PRO ASN
 ATAACAGCACCAAGCACAACTGACCCAAATG
 370 380 390
 GLY ASP ASN ASN GLN LEU THR GLN ALA GLN L
 GCGATAACAACCAACTGTGACCAAGCACAA
 400 410 420

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FIG.2C

YS THR ALA ALA ALA GLY PHE PHE VAL
 AGACCGCCGCTGCCGCAAGGGTTT TTTGTA
 430 440 450
 MET GLY LYS ILE ARG ASP THR SER PRO LYS A
 TGGGTAAATAATTCGTGATACCAAGCCCAAAA
 460 470 480

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SN ASP PRO ASP TYR SER ASN ASP LEU VAL
 ATGACCCAGATTATAGCAATGATTAGTAC
 490 500 510
 GLN GLN TRP GLN GLY LYS LEU TYR VAL GLY I
 AGCAGTGCGCAAGGCCAAATTAATGTTGGTA
 520 530 540

LE ASP ALA HIS ARG PRO ASP GLY ILE GLY
 TTGATGCCCATCGCCCAAGATGGCATCGGCA
 550 560 570
 THR GLY LYS ASN LEU ARG GLN PRO ILE THR A
 CAGGTAAACCTTGCGTCAGCCCATCACCG
 580 590 600

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FIG.2D

LA ASN ASP ILE LYS PRO LEU TYR PHE ASN
 CCAATGACATCAAAACCCCTTGTAATTTAACA
 610 620 630
 LYS PHE PRO ALA LEU SER ASP LEU HIS LEU A
 ATCCCTGCATTGTCTGATTGCAATTAG
 640 650 660

SP SER GLU ARG HIS ARG PHE ASP PRO LYS
 ACAGTGAAACGCCACCGTTTGTGACCCCAAA
 670 680 690
 LYS LEU ASN THR ILE LYS VAL TYR GLY TYR G
 AGCTAAACACCACTTAAGTGTTATGGTTATG
 700 710 720

LY ASN LEU THR THR PRO SER LYS ASN ASN
 GCAACTTAACAACCCCTCTAAACAACA
 730 740 750
 THR TYR ILE ASN HIS GLN GLN ALA ASP ASN L
 CTACATCAATCATCAGCAAGCTGATAATA
 760 770 780

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FIG.2E

YS LYS ASN ASN LYS PRO VAL ASP PRO TYR
 A G A A A T A C A G C C T G T T G A C C C T T A T
 790 800 810
 GLU ASN ILE ARG PHE GLY TYR LEU GLU LEU G
 G A A A T A T C C G T T T T G G G T A T C T T G A A C T A C
 820 830 840
 LN GLY SER SER LEU THR GLN LYS ASN ALA
 A G G A A G C A G T C T G A C C C A A A A A A T G C C G
 850 860 870
 ASP THR PRO ASN ASP LYS ASP ARG ILE PRO L
 A T A C T C C A A A T G A C A A A G A C C G C A T T C C C A
 880 890 900
 YS PRO MET PRO ILE LEU PHE TYR HIS GLY
 A C C C A T G C C C A T T T T G T T T A T C A C G G A G
 910 920 930
 GLU ASN ALA SER SER GLN LEU PRO SER ALA G
 A A A C G C C A G C A G C C A G C T G C C C A G T G C T G
 940 950 960

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FIG.2F

LY LYS PHE ASN TYR THR GLY ASN TRP LEU
 GTAAATTTAAC TACACAGGCAACTGGCTGT
 970 980 990
 TYR LEU SER ASP VAL LYS LYS ARG PRO ALA L
 ACC TAG TGA TGTCAA AAAACGCCCTGCAC
 1000 1010 1020

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 EU SER ALA SER ASP ARG VAL GLY VAL
 TTTCAGCATCAGATGATCGAGTGGGGTCT
 1030 1040 1050
 TYR LEU ASN ALA SER GLY LYS SER ASN GLU G
 ATCTCAATGCCAGTGGC AAAATCCAA TGAGG
 1060 1070 1080

LY ASP VAL VAL SER ALA ALA HIS ILE TYR
 GCGATGTCGTCAGTGCCCGCCACATTTATC
 1090 1100 1110
 LEU ASN GLY PHE GLN TYR LYS HIS THR PRO A
 TAAACGGCTTTCAATATTAAGCACACGCTG
 1120 1130 1140

FIG.2G

LA THR TYR GLN VAL ASP PHE ASP THR ASN
 CCACTATCAGGTGGATTTTGACACAAACT
 1150 1160 1170
 SER LEU THR GLY LYS LEU SER TYR TYR ASP A
 CATTACAGGCAAGCTGTCCTTATTATGACA
 1180 1190 1200
 SN PRO ASN GLN GLN THR ALA GLN GLY LYS
 ATCCCAACCAAGCAACTGCCCAAGGCAAAAT
 1210 1220 1230
 TYR ILE LYS SER GLN PHE ASP THR THR LYS L
 ACATCAAAAGCCAAATTTGACACTACCAAAA
 1240 1250 1260
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 YS VAL ASN GLU THR ASP VAL TYR GLN ILE
 AAGTCAATGAACCGGATGTGTATCAAAATG
 1270 1280 1290
 ASP ALA LYS ILE ASN GLY ASN ARG PHE VAL G
 ATGCCAAATAACAACGGCAACCGCTTCGTCG
 1300 1310 1320

FIG.2H

LY THR ALA LYS SER LEU VAL ASN GLU ASN
 GTACGGCCAAATCTTTGGTTAATGAGAACAA
 1330 1340 1350
 THR GLU THR ALA PRO PHE ILE LYS GLU LEU P
 CAGAAACCGCACCTTTTATCAAGAGCTGT
 1360 1370 1380
 HE SER LYS LYS ALA ASN PRO ASN ASN PRO
 TCTCCAAATAAGCCCAATCCCAATAACCCAA
 1390 1400 1410
 ASN PRO ASN SER ASP THR LEU GLU GLY P
 ACCCTAATTTCAGACACGCTAGAGGCGGT
 1420 1430 1440
 HE TYR GLY GLU SER GLY ASP GLU LEU ALA
 TTTATGGTGAGTCGGGCGATGAGCTGGCGG
 1450 1460 1470
 GLY LYS PHE LEU SER ASN ASP ASN ALA SER T
 GTAAATTTTATCCCAATGACACGCACTCTT
 1480 1490 1500

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FIG.2I.

YR VAL VAL PHE GLY GLY LYS ARG ASP LYS
 A T G T G G T C T T T G G T G G T A A A C G A G A C A A A A
 1510 1520 1530
 THR ASP LYS PRO VAL ALA THR LYS THR VAL T
 C A G A C A A A C C T G T C G C C A C A A A A C G G T G T
 1540 1550 1560
 YR PHE SER ALA GLY PHE GLU LYS PRO SER
 A T T T A G T G C A G G C T T T G A A A A C C T A G C A
 1570 1580 1590
 THR SER PHE VAL ASP ASN GLU THR ILE GLY A
 C C A G T T T T G T G G A T A A T G A A A C G A T T G G C A
 1600 1610 1620
 RG ILE ILE ASN SER LYS LYS LYS LEU ASN ASP
 G A A T T A T T A C A G C A A A A A G T T A A A T G A T G
 1630 1640 1650
 ALA VAL ASN GLU LYS ILE ASP ASN GLY ASP I
 C G G T G A A T G A G A A A A T T G A T A A T G G T G A T A
 1660 1670 1680

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FIG.2J

LE PRO THR SER ASP GLU ARG TYR ASP GLU
 TTCC TAC CAG TGA TGA A C G C T A T G A T G A A T
 1690 1700 1710
 PHE PRO TRP GLY GLU LYS LYS ALA GLU PHE T
 TTCC T T G G G C G A A A A A A A G C A G A A T T C A
 1720 1730 1740

HR LYS LYS VAL SER SER SER THR GLN ALA
 C C A A A A A G T C A G C A G C A C C C A A G C C G
 1750 1760 1770
 VAL PRO ALA TYR PHE GLY GLN HIS ASP LYS P
 T G C C A G C T T A T T T T G G G C A A C A T G A T A A A T
 1780 1790 1800

HE TYR PHE ASN GLY ASN TYR TYR ASP LEU
 T T T A T T T A A T G G C A A C T A T T A T G A C C T A T
 1810 1820 1830
 SER ALA SER SER VAL ASP LYS LYS LEU ALA PRO A
 C A G C C A G C A G T G T T G A T A A A T T G G C C C C T G
 1840 1850 1860

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FIG.2K

LA ASP ALA VAL LYS ALA ASN GIN SER ILE
 CCGATGCTGTC A AAGCCCAACCAATCCATT A
 1870 1880 1890
 LYS GLU LYS TYR PRO ASN ALA THR LEU ASN L
 AAGAAATAATACCCCTAATGCCACCTAATAA
 1900 1910 1920

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 YS ASP ASN GIN VAL THR ALA ILE VAL LEU
 AGGACAAACCAAGTTACCGCCCATCGTGCTAC
 1930 1940 1950
 GLN GLU ALA LYS ASP ASN LYS PRO TYR THR A
 AAGAGCCCAAGATAATAAGCCTTATACCG
 1960 1970 1980

LA ILE ARG ALA LYS SER TYR GIN HIS ILE
 CCATTCTGTC CAAAGCTATCAGCACATCA
 1990 2000 2010
 SER PHE GLY GLU THR LEU TYR ASN ASP ALA A
 GTTTTGCGGAGACGCTGTATACGATGCCAA
 2020 2030 2040

FIG.2L

SN GLN THR PRO THR ARG SER TYR PHE VAL
 ACCA AACCCCAACACGCGAGTTATTTTGTGC
 2050 2060 2070
 GLN GLY GLY ARG ALA ASP THR SER THR THR L
 AAGCGGCTAGGGCAGATACCGCACCCAGC
 2080 2090 2100
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 EU PRO LYS ALA GLY LYS PHE THR TYR ASN
 TGCCCAAGGCAAGGTAAATTCACTTACAACG
 2110 2120 2130
 GLY LEU TRP ALA GLY TYR LEU ILE GLN LYS L
 GTCCTTGGCGCAGGCTATCTTATCCAAATAA
 2140 2150 2160
 YS ASP LYS GLY TYR SER ASN ASN GLU GLU
 AGGACAAAGGTTATAGCAATAATGAGAAA
 2170 2180 2190
 THR ILE LYS LYS LYS GLY HIS GLN ASP TYR L
 CCATCAAGAAATAAGGCCATCAAGATTATC
 2200 2210 2220

FIG.2M

EU LEU THR GLU ASP PHE THR PRO GLU ASP
 TGT T A C C G A G A C T T C A C C C A G A G A T G
 2230 2240 2250
 ASP ASP ASP LEU THR ALA SER ASP ASP S
 A T G A C G A T G A T T T G A C C G C A T C T G A T G A T T
 2260 2270 2280
 ER GIN ASP ASP ALA HIS GLY ASP ASP
 C A C A G A T G A T G C A C A T G G C G A T G A T G
 2290 2300 2310
 ASP LEU ILE ALA SER ASP ASP SER GIN ASP A
 A T T T G A T T G C A T C T G A T G A T T C A C A G A T G
 2320 2330 2340
 SP ASP ALA ASP GLY ASP ASP SER ASP
 A T G A C G C A G A T G G C G A T G A C G A T T C A G A T G
 2350 2360 2370
 ASP LEU GLY ASP GLY ALA ASP ASP ALA ALA A
 A T T T G G G T G A T G G T G C A G A T G A C G C C C C G
 2380 2390 2400

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FIG.2N

LA GLY LYS VAL TYR HIS ALA GLY ASN ILE
 CAGGCAAGTGTTATCATGCAAGGTAATATTC
 2410 2420 2430
 ARG PRO GLU PHE GLU ASN LYS TYR LEU PRO I
 GCCCTGAATTGTGAACAACAATACTTGCCCA
 2440 2450 2460

LE ASN GLU PRO THR HIS GLU LYS THR PHE
 TTAATGAGCCTACTCATGAAACAACCTTTG
 2470 2480 2490
 ALA LEU ASP GLY LYS ASN LYS ALA LYS PHE A
 CCTAGATGGTTAAATAATAAGCTAAGTTTG
 2500 2510 2520

SP VAL ASP PHE ASP THR ASN SER LEU THR
 ATGTGGATTTTGACACCAACAGCCTAAC TG
 2530 2540 2550
 GLY LYS LEU ASN ASP GLU ARG GLY ASP ILE V
 GTAAATTAAACGATGAGAGAGGTGATATCG
 2560 2570 2580

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FIG.20

AL PHE ASP ILE LYS ASN GLY LYS ILE ASP
 T C T T G A T A T C A A A A T G G C A A A A T T G A T G
 2590 2600 2610
 GLY THR GLY PHE THR ALA LYS ALA ASP VAL P
 G C A C A G G C T T T A C C G C C A A A G C C G A T G T G C
 2620 2630 2640
 RO ASN TYR ARG GLU GLU VAL GLY ASN ASN
 C A A C T A T C G T G A G A G T G G G T A A C A A C C
 2650 2660 2670
 GLN GLY GLY PHE LEU TYR ASN ILE LYS A
 A A G G T G G C G G T T C T T A T A C A C A T C A A A G
 2680 2690 2700
 SP ILE ASP VAL LYS GLY GLN PHE PHE GLY
 A T A T G A T G T C A A G G G G C A A T T T T T G G C A
 2710 2720 2730
 THR ASN GLY GLU GLU LEU ALA GLY GLN LEU G
 C A A T G G C C G A G A G T T G G C A G G C A G T T A C
 2740 2750 2760

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FIG.2P

LN TYR ASP LYS GLY ASP GLY ILE ASN ASP
 AGTACGACAAAGGCCGATGGCATCAATGACA
 2770 2780 2790
 THR ALA GLU LYS ALA GLY ALA VAL PHE GLY A
 CCGCCGAAAGCAGGGCTGTCTTTGGGG
 2800 2810 2820

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 LA VAL LYS ASP LYS ***
 CTGTTAAAGATAAATAAGCCCCCTTCATC
 2830 2840 2850
 ATCGTTTAGTCGCCTTGACCGACAGTTGATG
 2860 2870 2880

ACGCCCTTGGCAATGTCTTAAACAGCACT
 2890 2900 2910
 TTGAACAGTGCCCTTGGGCGAATTCTTGA
 2920 2930 2940

TAAATGCCACGATTTGCCCTTGGGCTAATA
 2950 2960 2970

-35
 TCTTGATATAAACATCGCCATAAAATAGAAA
 2980 2990 3000

-10

FIG.2Q

A T A A A G T T T A G G A T T T T T T A T G T C A A A A T
 RBS MET SER LYS
 3010 3020 3030
 SER ILE THR LYS THR GLN THR PRO SER VAL H
 C T A T C A C A A A A C A C A A C A C C A T C A G T C C
 3040 3050 3060
 2nd possible start
 IS THR MET THR THR HIS ARG LEU ASN LEU
 A T A C C A T G A C C A C G C A C C G C T T A A C C T T G
 3070 3080 3090
 ALA ILE LYS ALA ALA LEU PHE GLY VAL ALA V
 C C A T C A A A G C G C G T T A T T G G T G T G C A G
 3100 3110 3120
 21/130
 AL LEU PRO LEU SER VAL TRP ALA GLN GLU
 T T T A C C C C T A T C C G T C T G G G C G C A G A G A
 3130 3140 3150
 ASN THR GLN THR ASP ALA ASN SER ASP ALA L
 A C A C T C A G A C A G A T G C C A A C T C T G A T G C C A
 3160 3170 3180

FIG.2R

YS ASP THR LYS THR PRO VAL VAL TYR LEU
 A G A C A C A A A A C C C C T G T C G T C T A T T T A G 3200
 3190
 ASP ALA ILE THR VAL THR ALA ALA PRO SER A
 A T G C C A T C A C G G T A A C C G C C C C C A T C T G 3220
 3230 3240
 LA PRO VAL SER ARG PHE ASP THR ASP VAL
 C C C C T G T T T C T C G G T T T G A C A C C G A T G T A A 3260
 3250
 THR GLY LEU GLY LYS THR VAL LYS THR ALA A
 C A G G G C T T G G C A A A C G G T C A A A C C G C T G 3280
 3290 3300
 SP THR LEU ALA LYS GLU GLN VAL GLN GLY
 A C A C G C T G G C A A A G A A C A A G T G C A G G G C 3320
 3310
 ILE ARG ASP LEU VAL ARG THR GLU THR GLY V
 A T T C G T G A T T T G G T G C G T T A T G A A C T G G G G 3340
 3350 3360

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FIG.2S

AL SER VAL VAL GLU GLN GLY ARG GLY GLY
 T G A G T G T G G T T G A G C A G G G C G T G G T G G C A
 3370 3380 3390
 SER SER GLY PHE ALA ILE HIS GLY VAL ASP L
 G C A G C G G A T T T G C C C A T T C A T G G C G T G G A T A
 3400 3410 3420

YS ASN ARG VAL GLY ILE THR VAL ASP GLY
 A A A C C G A G T G G G C A T T A C C G T A G A T G G C A
 3430 3440 3450
 ILE ALA GLN ILE GLN SER TYR LYS ASP GLU S
 T T G C C C A A A T T C A A T C C C T A C A A A G A T G A A T
 3460 3470 3480

ER THR LYS ARG ALA GLY ALA GLY SER GLY
 C C A C C A A A C G A G C T G G T G C A G G C T C T G G G G
 3490 3500 3510
 ALA MET ASN GLU ILE GLU ILE GLU ASN ILE A
 C G A T G A A T G A G A T A G A G A T T G A A A C A T T G
 3520 3530 3540

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FIG.2T

LA ALA VAL ALA ILE ASN LYS GLY GLY ASN
 CCGCCGTTGCCATCAATAAAGGTGGTAATG
 3550 3560 3570

ALA LEU GLU ALA GLY SER GLY ALA LEU GLY G
 CCTAGAGCAGGCTCTGGTGGCTTGGGCG
 3580 3590 3600

LY SER VAL ALA PHE HIS THR LYS ASP VAL
 GTTCGGTGGCGTTTCATACCAAGATGTGA
 3610 3620 3630

SER ASP VAL LEU LYS SER GLY LYS ASN LEU G
 GCGATGTCCTTAAATCTGGTAAATACTTG
 3640 3650 3660

LY ALA GLN SER LYS THR THR TYR ASN SER
 GCGCTCAAGCAAAACCACTTATAACAGCA
 3670 3680 3690

LYS ASN ASP HIS PHE SER GLN THR LEU ALA A
 AAAATGACCAATTTAGTCAGACCGCTGGCAG
 3700 3710 3720

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FIG.2U

LA ALA GLY LYS THR GLU ARG VAL GLU ALA
 C G G C A G G T A A A C C G A G C G T G T G G A A G C G A
 3730 3740 3750
 MET VAL GLN TYR THR TYR ARG LYS GLY LYS G
 T G G T G C A A T A T A C C T A C C G T A A A G G C A A A G
 3760 3770 3780
 LU ASN LYS ALA HIS SER ASP LEU ASN GLY
 A A A C A A A G C A C A C A G C G A C C T A A A T G G C A
 3790 3800 3810
 ILE ASN GLN SER LEU TYR ARG LEU GLY ALA T
 T C A A C C A A A G C C T A T A T C G C T T G G G T G C A T
 3820 3830 3840
 RP GLN GLN LYS TYR ASP LEU ARG LYS PRO
 G G C A C A A A A T A T G A T T T A A G A A A G C C C A
 3850 3860 3870
 ASN GLU LEU PHE ALA GLY THR SER TYR ILE T
 A T G A A C T G T T T G C A G G C A C A A G C T A C A T C A
 3880 3890 3900

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FIG.2V

HR GLU SER CYS LEU ALA SER ASP ASP PRO
 CCGAAGCTGTTTGGCAAGTGATGACCCAA
 3910 3920 3930
 LYS SER CYS VAL GLN TYR PRO TYR VAL TYR T
 AAGCTGCGTACAATACCCCTTATGTC TACA
 3940 3950 3960

HR LYS ALA ARG PRO ASP GLY ILE GLY ASN
 CCAAGCCCGACCAAGATGGCAATCGGCAATC
 3970 3980 3990
 ARG ASN PHE SER GLU LEU SER ASP ALA GLU L
 GCAATTCTCTGAGTTAAGCGATGCTGAAA
 4000 4010 4020

YS ALA GLN TYR LEU ALA SER THR HIS PRO
 AAGCAATAATTGGCAATCCACGCAACCCCC
 4030 4040 4050
 HIS GLU VAL VAL SER ALA LYS ASP TYR THR G
 ATGAGGTTGTCCTGCCCCAAGATTATACAG
 4060 4070 4080

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FIG.2W

LY ILE TYR ARG LEU LEU PRO ASP PRO MET
 G C A T T A T C G G T T G T T A C C C T G A C C C C A T G G
 4090 4100 4110
 ASP TYR ARG SER ASP SER TYR LEU ALA ARG L
 A C T A T C G T T C A G A C T C G T A T T G G C A C G C C
 4120 4130 4140

EU ASN ILE LYS ILE THR PRO ASN LEU VAL
 T T A C A T C A A A T C A C C C C A A A T C T G G T C A
 4150 4160 4170
 SER LYS LEU LEU LEU GLU ASP THR LYS GLN T
 G T A A C T G T T A T T A G A G A C A C C A G C A A A
 4180 4190 4200

HR TYR ASN ILE ARG ASP MET ARG HIS CYS
 C A T A C A C A T T C G T G A T A T G C G T C A T T G T A
 4210 4220 4230
 SER TYR HIS GLY ALA ARG LEU GLY ASN ASP G
 G T T A C C A T G G G G C A A G A T T G G C A A T G A T G
 4240 4250 4260

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FIG.2X

LY LYS PRO ALA ASN GLY GLY SER ILE VAL
 GTAGCC TGGCC AATGG TGGCTCC ATTGTT C 4280
 4270
 LEU CYS ASP ASP TYR GIN GLU TYR LEU ASN A
 TTTGGG ATGATTATCAAGAGTATCTAAACG 4310
 4300 4320
 LA ASN ASP ALA SER GIN ALA LEU PHE ARG
 CCAATGACGCATCAACAGCA TTATTAGAC 4340
 4330
 PRO GLY ALA ASN ASP ALA PRO ILE PRO LYS L
 CAGGTGCTAATGATGCCCCCATTC CCAAC 4360
 4370 4380
 EU ALA TYR ALA ARG SER SER VAL PHE ASN
 TGGCTTATGCCAGAGCAGTGTTGTTTAAAC 4400
 4390
 GIN GLU HIS GLY LYS THR ARG TYR GLY LEU S
 AAGAGCATGGCAAAACTCGCTATGGGTTAA 4420
 4430 4440

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FIG.2Y

ER PHE GLU PHE LYS PRO ASP THR PRO TRP
 GTT TGA GTT TAA G C C T G A C A C G C C A T G G T
 4450 4460 4470
 PHE LYS GLN ALA LYS LEU ASN LEU HIS GLN G
 T T A G C A A G C A A A A T T A A A C C T A C A C C A A C
 4480 4490 4500
 LN ASN ILE GLN ILE ILE ASN HIS ASP ILE
 A A A T A T C C A A A T C A T T A A C C A T G A C A T T A
 4510 4520 4530
 LYS LYS SER CYS SER GLN TYR PRO LYS VAL A
 A A A A T C G T G C A G C C A A T A T C C T A A G G T G G
 4540 4550 4560
 SP LEU ASN CYS GLY ILE SER GLU ILE GLY
 A T T A A A T G T G G C A T C A G T G A A A T T G G G C
 4570 4580 4590
 HIS TYR GLU TYR GLN ASN ASN TYR ARG TYR L
 A T T A T G A A T A T C A A A A T A A T A C C G T T A T A
 4600 4610 4620

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FIG.2Z

YS GLU GLY ARG ALA SER LEU THR GLY LYS
 A A G A A G G G C G T G C C A G C T T G A C A G G C A A A C
 4630 4640 4650
 LEU ASP PHE ASN PHE ASP LEU LEU GLY GLN H
 T T G A T T T A A T T T G A C C T G C T G G G T C A G C
 4660 4670 4680

IS ASP LEU THR VAL LEU ALA GLY ALA ASP
 A C G A T T G A C G G T G T T G G C T G G T G C A G A T A
 4690 4700 4710
 LYS VAL LYS SER GLN PHE ARG ALA ASN ASN P
 A A G T T A A A G C C C A A T T C G T G C C A A C A C C
 4720 4730 4740 4750

RO ARG ARG THR ILE ILE ASP THR THR GLN
 C C A G A C G C A C A A T C A T T G A C A C C A C C C A A G
 4750 4760 4770
 GLY ASP ALA ILE ILE ASP GLU SER THR LEU T
 G C G A T G C C A T C A T T G A T G A A A G C A C G C T G A
 4780 4790 4800

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FIG.2A'

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HR  ALA  GLN  GLU  GLN  ALA  LYS  PHE  LYS  GLN
CAGCACAGGAGCAAGCCAAATTTAAGCAAT
4810                                4820 4830
SER  GLY  ALA  ALA  TRP  ILE  VAL  LYS  ASN  ARG  L
CGGGGGCGGCATGGATTGTCAAAATCGCC
4840                                4850 4860

EU  GLY  ARG  LEU  GLU  GLU  LYS  ASP  ALA  CYS
TTGGACGCTTAGAAGAAAGACGCTGTG
4870                                4880 4890
GLY  ASN  ALA  ASN  GLU  CYS  GLU  ARG  ALA  PRO  I
GCAATGCCCAATGAATGTGAACGCGCCCCA
4900                                4910 4920

LE  HIS  GLY  SER  ASN  GLN  TYR  VAL  GLY  ILE
TTCATGGCAGTAACCAATAATGTGGCAATA
4930                                4940 4950
ASN  ASN  LEU  TYR  THR  PRO  ASN  ASP  TYR  VAL  A
ACAACTTTATATACCAATAATGATTGTG
4960                                4970 4980

```

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FIG.2B'

SP LEU SER PHE GLY GLY ARG LEU ASP LYS
 A T T A A G T T T G G T G G A C G C C T T G G A T A A A C
 4990 5000 5010
 G L N A R G I L E H I S S E R T H R A S P S E R A S N I L E I
 A A C G C A T T C A C A G C A C C G A T T C A A C A T C A
 5020 5030 5040

LE SER LYS THR TYR THR ASN LYS SER TYR
 T C A G C A A A A C T T A C A C C A A A A G C T A T A
 5050 5060 5070
 A S N P H E G L Y A L A A L A V A L H I S L E U T H R P R O A
 A T T T G G A G C G G C G G T T C A T C T G A C A C C T G
 5080 5090 5100

SP PHE SER LEU LEU TYR LYS THR ALA LYS
 A T T T A G C C C T G T T G T A T A A A A C T G C C A A A G
 5110 5120 5130
 G L Y P H E A R G T H R P R O S E R P H E T Y R G L U L E U T
 G C T T T C G T A C G C C A A G T T T T A T G A A C T G T
 5140 5150 5160

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FIG.2C'

YR ASN TYR ASN SER THR ALA ALA GLN HIS
 A C A A C T A T A C A G C A C C G C C C A G C A T A
 5170 5180 5190
 LYS ASN ASP PRO ASP VAL SER PHE PRO LYS A
 A A A T G A C C C T G A T G T G T C T T T C C C A A A C
 5200 5210 5220

RG ALA VAL ASP VAL LYS PRO GLU THR SER
 G A G C G G T T G A T G T C A A A C C T G A A C T T C C A
 5230 5240 5250
 ASN THR ASN GLU TYR GLY PHE ARG TYR GLN H
 A T A C C A A T G A A T A C G G C T T C G C T A T C A G C
 5260 5270 5280

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IS PRO TRP GLY ASP VAL GLU MET SER MET
 A C C C T T G G G G G A T G T T G A G A T G A G C A T G T
 5290 5300 5310
 PHE LYS SER ARG TYR LYS ASP MET LEU ASP L
 T C A A A A G C C G T T A C A A G G A C A T G T T A G A T A
 5320 5330 5340

FIG.2D'

YS ALA ILE PRO ASN LEU THR LYS ALA GLN
 AAGCCATAACCGAACCTAACCAAAGCCCCAAC
 5350 5360 5370
 GLN GLU TYR CYS LYS ALA HIS LEU ASP SER A
 AAGAGTATTGTAAGGCTCATTTGGATTCCA
 5380 5390 5400

SN GLU CYS VAL GLY ASN PRO PRO THR PRO
 ATGAATGTGTTGGCAATCCGCCACGCCCA
 5410 5420 5430
 LYS THR SER ASP GLU VAL PHE ALA ASN LEU T
 AACCAAGTGATGAGGTATTTGCCCACTTAT
 5440 5450 5460

YR ASN ALA THR ILE LYS GLY VAL SER VAL
 AATGCCCAACCATCAAGGGGTGAGTGTC A
 5470 5480 5490
 LYS GLY LYS LEU ASP LEU HIS ALA MET THR S
 AAGGCAAACTGGATTGCA TGCCATGACAT
 5500 5510 5520

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FIG.2E'

ER LYS LEU PRO ASP GLY LEU GLU MET THR
 C A A A C T G C C A G A T G G T C T T G A A A T G A C C T
 5530 5540 5550
 LEU GLY TYR GLY HIS THR LYS LEU GLY LYS P
 T G G G T T A T G G T C A T A C C A A A T T G G G G A A A T
 5560 5570 5580
 35/130
 HE ASP TYR ILE ALA PRO LYS ASP ALA ASP
 T T G A T T A C A T T G C A C C C A A A G A T G C C G A T G
 5590 5600 5610
 GLY TRP TYR GLN ALA ARG PRO ALA PHE TRP A
 G T T G G T A T C A G G C T C G C C C T G C T T T T G G G
 5620 5630 5640
 SP ALA ILE THR PRO ALA ARG TYR VAL VAL
 A T G C C A T C A C C C A G C G C G C T A T G T G G T C G
 5650 5660 5670
 GLY LEU ASN TYR ASP HIS PRO SER GLN VAL T
 G T C T A A A C T A T G A C C A C C C C A G T C A G T A T
 5680 5700

FIG.2F

RP GLY ILE GLY THR THR LEU THR HIS SER
 GGGGCA TTGGGCACAACTTTAACGCACAGCA
 5710 5720 5730
 LYS GLN LYS ASP GLU ASN GLU LEU SER ALA L
 AACAAAGATGAATAAGCTAAGTGCCC
 5740 5750 5760
 EU ARG ILE ARG ASN GLY LYS ARG GLU THR
 TTAGAA TCCGAAATGGCAAAAGAGAAACAC
 5770 5780 5790
 GLN THR LEU THR HIS THR ILE PRO LYS ALA T
 AACCTTAACGCACACAAATACCCAAAGCCT
 5800 5810 5820
 YR THR LEU LEU ASP MET THR GLY TYR TYR
 ATACCTTACTGGACATGACAGGCTATTATA
 5830 5840 5850
 SER PRO THR GLU SER ILE THR ALA ARG LEU G
 GCCCAACTGAGAGCATCACCGCTCGTCTG
 5860 5870 5880

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FIG.2G'

LY ILE ASN ASN VAL LEU ASN THR ARG TYR
 GTATCAACAATGTATTAAACACCCGCTACA
 5890 5900 5910
 THR THR TRP GLU ALA ALA ARG GLN LEU PRO S
 CCACATGGGAAGCGGCAACGCCAACCTGCCCA
 5920 5930 5940

ER GLU ALA ALA SER SER THR GLN SER THR
 GGAGCTGCAAGCAGTACCCAAATCAACCC
 5950 5960 5970
 ARG TYR ILE ALA PRO GLY ARG SER TYR PHE A
 GTACATTGCACCAAGGTCCGCACTTCTTG
 5980 5990 6000

LA SER LEU GLU MET LYS PHE *** MET THR
 CCAGTCTTGAAATGAAAGTTTATAATGACC
 6010 6020 6030
 CYS LEU PRO LYS THR ASN PRO ALA LEU . LYS
 TGTTTACCAAGACCAACCCCTGCTTAA
 6040 6050 6060

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FIG.2H'

VAL LYS HIS ARG PHE LEU LYS GLN VAL LEU
 G T C A A G C A C A G A T T T T A A G C A G G T G C T G
 6070 6080 6090
 LEU LEU LEU CYS VAL ASP THR LEU THR ALA
 T T A T T G C T T T G T G T T G A T A C A T T A C A G C A
 6100 6110 6120
 GLN ALA TYR ALA HIS SER HIS HIS THR PRO
 C A G G C G T A C G C C C A C A G C C A T C A T A C G C C C
 6130 6140 6150
 ILE HIS THR PRO THR HIS GLU LEU PRO SER
 A T T C A T A C A C C C A C G C A T G A G C T G C C A T C T
 6160 6170 6180
 ALA ASP ALA LEU SER ASP GLU GLY LEU GLY
 G C T G A T G C C T T T A T C A G A T G A A G G C T T G G G T
 6190 6200 6210
 LYS ASP LEU GLY SER LEU ASP SER LEU ASP
 A A G G A T T T G G G C A G T T T G G A C A G T T T G G A T
 6220 6230 6240

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FIG.2I'

SER PRO ASP GLY LEU GLY ASP GLY LEU GLY
 A G C C A G A T G G T T T G G G T G A T G G T T T A G G C
 6250 6260 6270
 ASP GLY LEU GLY ASP GLY LEU LYS SER ASP
 G A T G G T T T G G G T G A T G G C T T A A A A G T G A T
 6280 6290 6300

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 LYS ALA PRO LEU PRO ILE ASN ALA LEU THR
 A A G C C C C T T A C C C A T C A C G C C T T G A C C
 6310 6320 6330
 ALA HIS GLN THR ASN GLU SER GLN PRO ALA
 G C C C A T C A G A C C A A T G A G A G C C A G C C T G C C
 6340 6350 6360

PRO PRO SER VAL ASP VAL ASN PHE LEU LEU
 C C A C C G A G C G T A G A T G T C A A T T T T A C T T
 6370 6380 6390
 ALA GLN PRO GLU ALA PHE TYR HIS VAL PHE
 G C C C A G C C A G A G G C A T T T T A T C A T G T C T T T
 6400 6410 6420

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FIG.2J'

```

HIS  GLN  ALA  ILE  VAL  GLN  ASP  ASP  VAL  ALA
CATCAAGCGATTGTGCAAGATGATGGCA
6430
        THR  LEU  ARG  LEU  LEU  LEU  PRO  PHE  TYR  ASP
        A C A T T A C G C T T G T T A T T G C C A T T T A T G A C
        6460
ARG  LEU  PRO  ASP  ASP  TYR  GLN  ASP  ASP  VAL
CGCC T G C C T G A T G A T T A T C A A G A T G T T
6490
        LEU  LEU  LEU  PHE  ALA  GLN  SER  LYS  LEU  ALA
        T T G T T G T T A T T T G C C C A A A G T A A A C T T G C C
        6520
LEU  SER  ASP  GLY  ASN  THR  LYS  LEU  ALA  LEU
CTAAGTGATGGCAATAACCAATAATGGCA TTG
6550
        ASN  LEU  LEU  THR  ASP  LEU  SER  ASN  LYS  GLU
        A A T C T G C T G A C C G A T T T G A G T A A C A A G A G
        6580

```


FIG.2K'

PRO THR LEU THR ALA VAL LYS LEU GLN LEU
 C C A A C A C T T A C G G C G G T A A A T T A C A C T T
 6610 6620 6630
 ALA SER LEU LEU LEU THR ASN LYS HIS ASP
 G C T T C C T T G T T G C T G A C C A A C A G C A C G A T
 6640 6650 6660

LYS HIS ALA GLN MET VAL LEU ASP GLU LEU
 A A A C A C G C C C A A A T G G T G C T A G A T G A C T C
 6670 6680 6690
 LYS ASP ASP ALA HIS PHE LEU LYS LEU SER
 A A A G A T G A T G C C C A C T T T T A A A T T A A G C
 6700 6710 6720

LYS LYS GLU GLN ARG TRP VAL LEU SER GLN
 A A A A A G A G C A A A G A T G G G T G C T A T C G C A A
 6730 6740 6750
 SER ARG TYR LEU HIS LYS LYS TYR LYS MET
 A G T C G C T A T T T A C A T A A A A A T A T A A A T G
 6760 6770 6780

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FIG.2L'

GLY LEU ASP LEU GLY ILE ASN TYR LEU HIS
 GGCTTGGATT TGGGCA TCAACTATCTGCAT
 6790 6800 6810
 LEU ASP ASN ILE ASN ALA ALA SER THR ILE
 TTGGATAATA TCAACGCCGCTCCACCATC
 6820 6830 6840

THR GLN PRO ASN ILE LYS LYS ASP ALA PRO
 ACCCAGCCCAATA TAAAGATGCCCCA
 6850 6860 6870
 LYS PRO ALA HIS GLY LEU ALA LEU SER LEU
 AACCTGCTCATGGGCTTGCCCTTATCGCTT
 6880 6890 6900

GLY VAL ASN LYS TYR THR PRO LEU SER HIS
 GGTGTGAATAATA CACGCCGCTTAGTCAT
 6910 6920 6930
 GLY MET SER ILE TYR THR ALA LEU ASP VAL
 GGCA TGAGTATTTATACAGCCCTAGATGTT
 6940 6950 6960

SUBSTITUTE SHEET (RULE 26)

FIG.2M'

ASP GLY LYS PHE TYR ASP ASP LYS SER HIS
 G A T G G T A A A T T T A T G A T G A C A A A G C C A C
 6970 6980 6990
 ASN GLU LEU ALA VAL PHE ALA HIS ALA GLY
 A A T G A A C T G G C G G T T T T G C T C A T G C T G G A
 7000 7010 7020

LEU ARG LYS ASP HIS GLN LYS GLY TYR VAL
 C T A G A A A A G A T C A C C A A A A G G T T A T G T T
 7030 7040 7050
 ASP VAL VAL PRO PHE VAL GLY ARG ILE PHE
 G A T G T C G T A C C T T T T G T T G G G C G T A T T T T T
 7060 7070 7080

ALA THR ASN GLN GLN HIS GLY ARG LEU SER
 G C C A C C A A T C A G C A G C A T G G C A G A T T A T C C
 7090 7100 7110
 PRO ARG LYS ASP SER GLN GLY VAL ALA PHE
 C C C A G A A A G A C A G T C A G G G C G T G G C G T T T
 7120 7130 7140

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FIG.2N'

GLY SER HIS HIS ARG ILE ASN ASP LYS TRP
 GGCAGCCATCATCGGATCAATGATAAATGG
 7150 7160 7170
 GIN ASN ALA PHE PHE ALA ARG MET GLU LYS
 CAAATGCGCTTTTTCACCGCATGGAAAAA
 7180 7190 7200

GLY ASN TYR THR GLU ARG TYR GIN GLY TYR
 GGCAATTATACCGAGCGTTATCAAGGTTAT
 7210 7220 7230
 ASP GLY LYS ARG TYR HIS VAL ASN ASP THR
 GATGGCAAGCGTTATCATGTGAATGACACC
 7240 7250 7260

ILE LEU LEU GIN ASP GLY PRO ASN ARG ARG
 ATTTGTGTGCAAGATGGCCCCAAATCGTCGT
 7270 7280 7290
 TYR SER LEU GLY VAL GLY TYR GIN LEU SER
 TACTCTTTGGGCGGTGGGGTATCAGCTTAGC
 7300 7310 7320

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FIG.20'

HIS LEU GIN ASP ALA THR LYS SER SER HIS
 C A T C T G C C A A G A T G C C A A A A A G C A G T C A T
 7330 7340 7350
 ALA THR LYS ILE HIS PHE GLY VAL LEU GIN
 G C C A C A A A G A T A C A T T T T G G G G T G T T G C A A
 7360 7370 7380

ARG LEU PRO ASN GLY LEU THR VAL GIN GLY
 A G A T T G C C C A A A T G G T C T G A C C G T G C C A A G G T
 7390 7400 7410
 ARG VAL SER ALA GLU ARG GLU ARG TYR HIS
 A G A G T G A G T G C T G A G C G T G A G C G T T A T C A T
 7420 7430 7440

GLY LYS LEU LEU ARG LEU VAL ASN PRO ASP
 G G T A A A T T A T T G C G T C T G G T T A A T C C T G A T
 7450 7460 7470
 ASP VAL TYR ARG THR ASP LYS THR LEU THR
 G A T G T G T A T C G C A C A G A T A A A C C C T A C C
 7480 7490 7500

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FIG.2P'

LEU GLN THR SER ILE TRP HIS LYS ASP ILE
 C T A C A A C C C T C C A T T T G G C A C A A G A C A T T
 7510 7520 7530
 HIS TRP LEU GLY LEU THR PRO LYS LEU THR
 C A C T G G C T T G G A T T A C G C C A A G C T G A C T
 7540 7550 7560
 TYR ARG TYR SER LYS ASN ASN SER ASN LEU
 T A T C G T T A C A G T A A A A T A A C A G T A A C T T A
 7570 7580 7590
 PRO ALA LEU TYR SER HIS ASN LYS GLN ASN
 C C A G C A C T T T A T A G C C A T A C A A C A A A T
 7600 7610 7620
 PHE TYR LEU GLU LEU GLY ARG SER PHE ***
 T T T A T T G G A G C T T G G T C G G T C G T T T A A
 7630 7640 7650

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Restriction map of clone pLD1-8, *M. catarrhalis* strain 4223 *lfr*

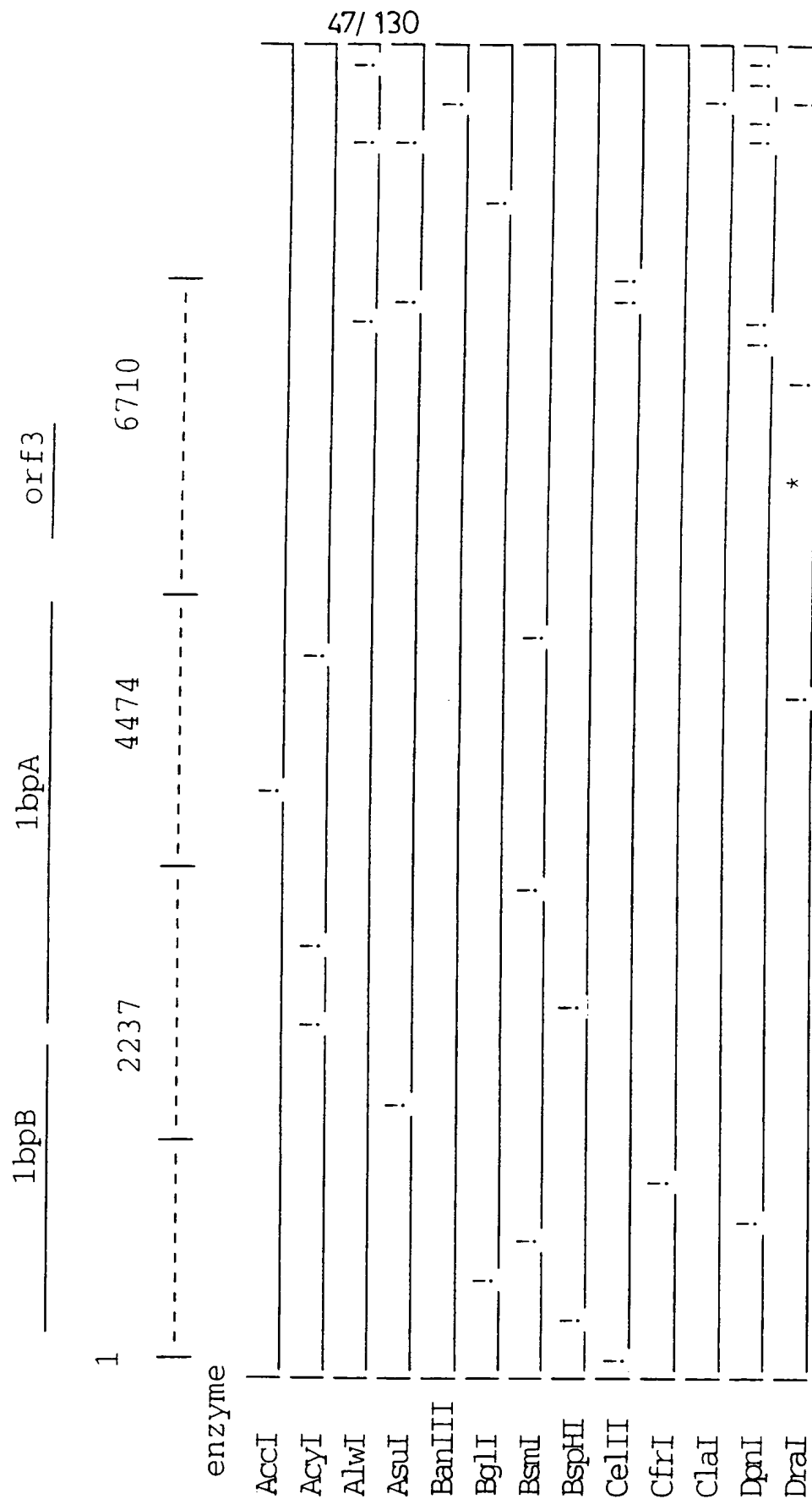


FIG.3A

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EaeI	!	
EarI	!	
EcoRI	!	
EcoRV	!	*
FokI	!	!
HaeII	!	
HaeIII	!	!
HhaI	!	*
HindIII	!	
HpaI	!	
NcoI	!	!
NdeI	!	
PvuII	!	!
ScaI	!	
SphI	!	
SspI	!	!
TagI	!	!

FIG.3B

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FIG.4A *M. catarrhalis* Q8 *lfr* sequence

AAGCTTAGCATGATGGCATCGGCTGATTGT
 10 20 30
 CTTTGGCCCTTGTGTGTGTGTGTGGGAGT
 40 50 60

TGATTGTACTTACCTTAGTGGTGGATGCTT
 70 80 90
GGGCTGATTTAATAATAATCAAGCG
 -35 -10 100 110 120

GTCTTCACAACACACCAAAACGAGATATCAC
 130 140 150
 RBS
 lbp2
 MET SER THR VAL LYS THR PRO HIS ILE PHE
 CATGAGTACTGTCAAAACCCCATATT
 160 170 180

TYR GLN LYS ARG THR LEU SER LEU ALA ILE
 CTACCAAAACGCACCCCTTAGCCCTTGCCAT
 190 200 210
 ALA SER ILE PHE ALA ALA LEU VAL MET THR
 CGCCAGTATTTTTGTGCTGCCCTTGGTGATGAC
 220 230 240

B
4.
G
L

GLY CYS ARG SER ASP ILE SER VAL ASN
 AGGCTGCCCGCTCTGATGACATCAGCGTCAA
 260 270
 ALA PRO ASN VAL THR GLN LEU PRO GLN GLY
 TGCACCCCAATGTTACCCAGCTGCCCAAGG
 280 290 300

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THR VAL SER PRO THR PRO ASN THR GLY HIS	
CACGGTTTCACC AACGCCGAACAAGGTCA	
310	320
	ASP ASN ALA ASN ASN THR ASN ASN GLN GLY
	TGACAA CGCCATAACACCAACAATCAGGG
	340
	350
	360

ASN ASN THR ASP ASN SER THR THR
CAACAACACGGATAACAGCACCAAC
370 380 390
ASP PRO ASN GLY ASP ASN ASN GLN LEU THR
TGACCCAAATGGCGATAACAACCACTGAC
400 410 420

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FIG.4C

GIN ALA GIN LYS THR ALA ALA ALA GLY
 A C A G C G C A A A A A C T G C C C G C C G C A G G 440
 430
 PHE PHE VAL MET GLY LYS ILE ARG ASP THR
 G T T T T T G T G A T G G G T A A A A T T C G T G A T A C 460
 470 480

SER GLU LYS ASN ASP PRO ASP TYR SER ASP
 C A G C G A A A A A A T G A C C C A G A T T A T A G T G A T 500
 490
 ASP LEU LYS GIN GIN TRP LEU GLY LYS LEU
 T G A T T T A A A A C A G C A G T G G C T G G G C A A A T T 520
 530 540

TYR VAL GLY ILE ASP ALA HIS ARG PRO ASP
 A T A T G T T G G T A T T G A T G C C C A T C G C C C A G A 560
 550
 GLY ILE GLY LYS GIN LYS ASN LEU ARG GIN
 T G G C A T C G G A A A A G G T A A A A C T T G C G T C A 580
 590 600

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FIG.4D

PRO ILE THR ALA ASN ASP ILE LYS PRO LEU
 GCCCATCACC GCCAATGACATCAAAACCTT 610
 TYR PHE ASN LYS PHE PRO ALA LEU SER ASP
 GTATTTTAACAAATTCCCTGCATTGCTGA 640
 650
 LEU HIS LEU ASP SER GLU ARG HIS ARG PHE
 TTGCACTTAGACAGTGACGCCATCGTTT 670
 680
 ASP PRO GLN LYS ILE ASN THR ILE LYS VAL
 TGACCCCAAAAGATAAACACCAATTAAAGT 700
 710
 720
 TYR GLY TYR GLY ASN LEU THR THR PRO SER
 GTATGGTTATGGTAACTTAAACAACCAATC 730
 740
 750
 ASN ASN ASN THR HIS ILE ASN HIS GLN GLN
 CAACAACAACACTCACATCAATCATCAGCA 760
 770
 780

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ALA ASP ASN LYS LYS LYS LYS LYS ASN ASN LYS PRO VAL
 AGCTGATAATAAGAAATAACAGCCTGT
 790 800 810
 ASP PRO TYR GLU ASN ILE ARG PHE GLY TYR
 TGACCCCTTATGAATAATATCCGTTTGGGTA
 820 830 840
 LEU GLU LEU GLN GLY SER SER LEU THR GLN
 TCTTGAACTACAAGGAAGCAGCCTGACCA
 850 860 870
 LYS ASN ALA ASP ASN GLN ASN GLU GLN ASP
 AAAATGCCGATAATACAAATGAGCAAGA
 880 890 900
 ARG ILE PRO LYS PRO MET PRO ILE LEU PHE
 CCGCATTCCCAACCCCATGCCCATTTGT
 910 920 930
 TYR HIS GLY GLU ASN ALA SER SER GLN LEU
 TTATCATGGAGAAACGCCAGCAGCAGCT
 940 950 960

FIG.4F

PRO SER ALA GLY LYS PHE ASN TYR THR GLY
 GCCCAGCGCTGGTAATAATTAACTACACAGG 970
 980
 ASN TRP LEU TYR LEU SER ASP VAL LYS LYS
 CAACCTGGCTGTACCTAAGTGATGTCAAAA 1000
 1010
 1020
 ARG PRO ALA LEU SER ALA SER ASP GLU ARG
 ACGCCCTGCCCTTTCAGCATCAGATGAGCG 1030
 1040
 1050
 VAL GLY VAL TYR LEU ASN ALA SER GLY LYS
 AGTGGGGGTCTATCTCAATGCCAGTGGCAA 1060
 1070
 1080
 ALA ASN GLU GLY ASP VAL VAL SER ALA ALA
 AGCCAACGAGGCGGATGTCGTCA GTGCCGC 1090
 1100
 1110
 HIS ILE TYR LEU ASN GLY PHE GLN TYR LYS
 CCACATTTATCTAAACGGCTTTCAATAATA 1120
 1130
 1140

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FIG.4G

HIS THR PRO ALA THR TYR GLN VAL ASP PHE
 GCACGCCCTGCCACTTATCAGGTGGATT
 1150 1160 1170
 ASP THR ASN SER LEU THR GLY LYS LEU SER
 TGACACAAACCTCATTAACAGGCAGCTGTC
 1180 1190 1200

TYR TYR ASP ASN PRO ASN GLN GIN ASN ASN
 CTATTATGACAAATCCCAATCAGCAATAA
 1210 1220 1230
 LYS GLY GLU TYR LEU LYS SER GLN PHE ASP
 TAAAGGCCGAATATCTCAAAAGCCAAATTGA
 1240 1250 1260

THR THR LYS LYS VAL ASN GLU THR ASP VAL
 CACTACCAAAAGTCAATGAACCGATGT
 1270 1280 1290
 TYR GLN ILE ASP ALA LYS ILE ASN GLY ASN
 GTATCAAAATTGATGCCCAAAATCAACGGTAA
 1300 1310 1320

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FIG.4H

ARG PHE VAL GLY THR ALA LYS SER LEU VAL
 CCGCTTTGTCGGGTACGGCCAAATCTTTGGT
 1330 1340 1350
 ASN GLU LYS THR GLN THR ALA PRO PHE ILE
 TAATGAGAAACAACAACCGCACTTTAT
 1360 1370 1380
 LYS GLU LEU PHE SER LYS LYS ALA ASN PRO
 CAAGAGCTGTTCTCCAAATAAGCCACCC
 1390 1400 1410
 ASN ASN PRO ASN PRO ASN SER ASP THR LEU
 CAATAACCCAAACCCCTAATTCAAGACACGCT
 1420 1430 1440
 GLU GLY GLY PHE TYR GLY GLU SER GLY ASP
 AGAAGCGGATTTTATGTTAGTCTGGGCGA
 1450 1460 1470
 GLU LEU ALA GLY LYS PHE LEU SER ASN ASP
 TGAGCTGGCGGGTAATAATTATTCCCAATGA
 1480 1490 1500

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FIG. 4I

ASN ALA SER TYR VAL VAL PHE GLY GLY LYS
 C A A C G C A T C T T A T G T G G T C T T T G G T G G C A A
 1510 1520 1530
 ARG ASP LYS THR THR LYS PRO VAL ALA THR
 A C G A G A C A A A A C G A C T A A A C C T G T C G C C A C
 1540 1550 1560

LYS THR VAL TYR PHE SER ALA GLY PHE GLU
 A A A A C G G T G T A T T T A G T G C A G G C T T T G A
 1570 1580 1590
 LYS PRO SER THR SER PHE VAL ASP ASN GLU
 A A A C C C A G C A C C A G T T T T G T G G A T A A T G A
 1600 1610 1620

THR ILE GLY GLY ILE ILE ASP ARG LYS GLY
 A A C G A T T G G T G G A A T T A T T G A C C G T A A A G G
 1630 1640 1650
 LEU ASN ASN HIS ILE ASN GLU ASP GLU ILE
 G T T A A A T A A T C A C A T T A A T G A A G A T G A A A T
 1660 1670 1680

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FIG. 4J

[illegible][illegible]

Tyr Phe Asn Gly Asp Leu Ser
T T A T T T A A T G G C A A C T A T T A T G A C C T A T C

1810 1820 1830

Ala Ser Arg Val Lys Leu Ala Pro Ala
A G C C A G T C G T G T G A T A A A T T A G C C C C T G C

1840 1850 1860

FIG.4K

ASP ALA VAL LYS ALA ASN GLN SER ILE LYS
 CGATGCTGTC AAGCCAAACCAATCCATTAA
 1870 1880 1890
 GLU LYS TYR PRO ASN ALA THR LEU ASN LYS
 AGAAATAATACCCCTAATGCCACACTAAATAA
 1900 1910 1920
 ASP ASN GLN VAL THR ALA ILE VAL LEU GLN
 GGACACCAAGTTACCGCCATCGTGCTACA
 1930 1940 1950
 GLU ALA LYS ASP ASN LYS PRO TYR THR ALA
 AGAGCCCAAAGATAATAAGCCCTTATACCGC
 1960 1970 1980
 ILE ARG ALA LYS SER TYR GLN HIS ILE SER
 CATTCGTGCCCAAAGCTATCAGCACATCAG
 1990 2000 2010
 PHE GLY GLU THR LEU TYR ASN ASP ALA ASN
 TTTTGGCGAGACGCTGTATACGATGCCAA
 2020 2030 2040

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FIG.4L

GLN THR PRO THR ARG SER TYR PHE VAL GLN
 C C A A C C C A A C A C G C A G T T A T T T G T G C A
 2050 2060 2070
 GLY GLY ARG ALA ASP THR SER THR LEU
 A G G C G G T A G G G C A G A T A C C A G C A C A C T T T
 2080 2090 2100
 PRO GLN ALA GLY LYS PHE THR TYR ASN GLY
 G C C C A G G C A G G T A A T T C A C T T A C A A C G G
 2110 2120 2130
 LEU TRP ALA GLY TYR LEU THR GLN LYS LYS
 T C T T T G G C A G G C T A C C T G A C C C A A A A A A
 2140 2150 2160
 ASP LYS GLY TYR SER ASP ASN ALA GLU THR
 G G A C A A A G G T T A T A G C G A T A A T G C A G A A C
 2170 2180 2190
 ILE LYS GLU LYS GLY HIS PRO GLY TYR LEU
 C A T C A A G G A A A A A G G T C A T C C A G G T T A T C T
 2200 2210 2220

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FIG. 4M

LEU THR GLU ASN PHE THR PRO GLU ASP ASP
 GTT ACC GAA AACTT CACCCAGAGATGA 2230 2240 2250
 ASP ASP LEU THR ALA SER ASP ASP SER
 TGACGATGATTGTGACCGCATCTGATGATTC 2260 2270 2280
 GIN ASP ASP ASN THR HIS GLY ASP ASP
 ACAAGATGATAATACACATGGCGATGATA 2290 2300 2310
 LEU ILE ALA SER ASP ASP SER GLN ASP ASP
 TTTGATTGCAATCTGATGATTCACAGATGA 2320 2330 2340
 ASP ALA ASP GLY ASP ASP SER ASP ASP
 TGACCGAGATGGAGATGACGATTGAGATGA 2350 2360 2370
 LEU GLY ASP GLY ALA ASP ASP ASP ALA ALA
 TTTGGGGTGATGGTGCCAGATGATGACCGCC 2380 2390 2400

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FIG.4N

GLY LYS VAL TYR HIS ALA GLY ASN ILE ARG
 AGG C A A A G T G T A T C A T G C A G G T A A T A T T C G
 2410 2420 2430
 PRO GLU PHE GLU ASN LYS TYR LEU PRO ILE
 C C C T G A A T T T G A A A C A A A T A C T T G C C C A T
 2440 2450 2460

ASN GLU PRO THR HIS GLU LYS THR PHE ALA
 T A A T G A G C C T A C T C A T G A A A A A C C T T T G C
 2470 2480 2490
 LEU ASP GLY LYS ASN LYS ALA LYS PHE GLU
 C C T A G A T G G T A A A A A T A A A G C T A A G T T T G A
 2500 2510 2520

VAL ASP PHE ASN THR ASN SER LEU THR GLY
 A G T G G A T T T A A C A C C A A C A G C C T A A C T G G
 2530 2540 2550
 LYS LEU ASN ASP GLU ARG GLY ASP ILE VAL
 T A A A T T A A C G A T G A G A G A G G T G A T A T C G T
 2560 2570 2580

FIG.4.O

PHE ASP ILE LYS ASN GLY LYS ILE ASP GLY
 C T T G A T A T C A A A A T G G C A A A A T T G A T G G
 2590 2600 2610
 THR GLY PHE THR ALA LYS ALA ASP VAL PRO
 C A C A G G A T T T A C C G C C A A A G C C G A T G T G C C
 2620 2630 2640
 ASN TYR ARG GLU GLU VAL GLY ASN ASN GLN
 A A A C T A T C G T G A A G A G T G G G T A A C A A C C A
 2650 2660 2670
 GLY GLY GLY PHE LEU TYR ASN ILE LYS ASP
 A G G T G G C G G T T T C T T A T A C A A C A T C A A A G A
 2680 2690 2700
 ILE ASP VAL LYS GLY GIN PHE PHE GLY THR
 T A T T G A T G T T A A G G G G C A A T T T T T T G G C A C
 2710 2720 2730
 ASN GLY GLU GLU LEU ALA GLY GIN LEU HIS
 A A A T G G C G A A G A G T T G G C A G G A C A G T T A C A
 2740 2750 2760

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FIG.4P

HIS ASP LYS GLY ASP GLY ILE ASN ASP THR
 T C A T G A C A A A G G C G A T G G C A T C A A T G A C A C 2770
 2780
 ALA GLU LYS ALA GLY ALA VAL PHE GLY ALA
 C G C C G A A A A G C A G G G C T G T C T T G G G C C 2800
 2810 2820
 VAL LYS ASP LYS ***
 T G T T A A G A T A A A T A A G C C C C C C T T C A T C 2830
 2840 2850
 A T C G T T T A G T C G C T T G A C C G A C A G T T G A T G 2860
 2870 2880
 A C G C C C T T G G C A A T G T C T T A A A C A G C A C T 2890
 2900 2910
 T T G A A A C A G T G C C T T G G G C G A A T T C T T G G A 2920
 2930 2940
 T A A A T G C A C C A G A T T T G C C T T G G C T A A T A 2950
 2960 2970
 -10
 T C T T G A T A A A C A T C G C C A T A A A A T A G A A A 2980
 2990 3000
 -35

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FIG.4Q

A T A A G T T A G G A T T T T T A T G T C A A A A
 3010 3020 3030
 RBS MET SER LYS
 Lbp1
 SER ILE THR LYS THR GIN THR PRO SER VAL H
 T C T A T C A C A A A C A C A A C A C C A T C A G T C C
 3040 3050 3060
 65/130
 IS THR MET THR THR HIS ARG LEU ASN LEU
 A T A C C A T G A C C A C G C T T A A C C T T G
 3070 3080 3090
 ALA ILE LYS ALA ALA LEU PHE GLY VAL ALA V
 C C A T C A A A G C G G C G T T A T T G G T G T G G C A G
 3100 3110 3120
 AL LEU PRO LEU SER VAL TRP ALA GIN GLU
 T T T A C C C C T A T C C G T C T G G C G C A A G A G A
 3130 3140 3150
 ASN THR GIN THR ASP ALA ASN SER ASP ALA L
 A C A C T C A G A C A G A T G C C A A C T C T G A T G C C A
 3160 3170 3180

FIG.4R

YS ASP THR LYS THR PRO VAL VAL TYR LEU
 A G A C A C A A A A C C C T G T C G T C T A T T T A G
 3190 3200 3210
 ASP ALA ILE THR VAL THR ALA ALA PRO SER A
 A T G C C A T C A C G G T A A C C G C C C C C A T C T G
 3220 3230 3240

LA PRO VAL SER ARG PHE ASP THR ASP VAL
 C C C C T G T T T C T C G G G T T T G A C A C C G A T G T A A
 3250 3260 3270
 THR GLY LEU GLY LYS THR VAL LYS THR ALA A
 C A G G G C T T G G C A A A C C G T C A A A C C G C T G
 3280 3290 3300

SP THR LEU ALA LYS GLU GLN VAL GLN GLY
 A C A C G C T G G C A A A G A A C A A G T A C A G G G C A
 3310 3320 3330
 ILE ARG ASP LEU VAL ARG TYR GLU THR GLY V
 T T C G T G A T T T G G T G C G T T A T G A A C T G G G
 3340 3350 3360

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FIG.4S

AL SER VAL VAL GLU GLN GLY ARG GLY GLY
 T G A G T G T G G T T G A G C A G G G C G T G G T G G C A
 3370 3380 3390
 SER SER GLY PHE ALA ILE HIS GLY VAL ASP L
 G C A G C G G A T T T G C C A T T C A T G G C G T G G A T A
 3400 3410 3420

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 YS ASN ARG VAL GLY ILE THR VAL ASP GLY
 A A A C C G A G T G G C A T T A C C G T A G A T G G C A
 3430 3440 3450
 ILE ALA GLN ILE GLN SER TYR LYS ASP GLU S
 T T G C C C A A A T T C A A T C C T A C A A G A C G A A T
 3460 3470 3480

ER THR LYS ARG ALA GLY ALA GLY SER GLY
 C C A C T A A G C G A G C T G G G C A G G C T C T G G G G
 3490 3500 3510
 ALA MET ASN GLU ILE GLU ILE GLU ASN ILE A
 C G A T G A A C G A G A T A G A G A T T G A A A C A T T G
 3520 3530 3540

FIG.4T

LA ALA VAL ALA ILE ASN LYS GLY GLY ASN
 CCGCCGTTGCCCATCAATAAAGCGGGTAATG
 3550 3560 3570
 ALA LEU GLU ALA GLY SER GLY ALA LEU GLY G
 CCTTAGAAGCAGGCCTCTGGGTGGGTGGGTG
 3580 3590 3600

LY SER VAL ALA PHE HIS THR LYS ASP VAL
 GTTCGGTGGCGTTTCAATACCAAGATGTGA
 3610 3620 3630
 SER ASP VAL LEU LYS SER GLY ASN ASN LEU G
 GGATGTCCTTAAATACTGGGTAAACAATCTTG
 3640 3650 3660

LY ALA GLN SER LYS THR THR TYR ASN SER
 GTGCTCAAGCAAAACCACTTATAACAGCA
 3670 3680 3690
 LYS ASN ASP HIS PHE SER GLN THR LEU ALA A
 AAAATGACCATTTTAGTCAGACGCTGGCAG
 3700 3710 3720

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FIG.4U

LA ALA GLY LYS THR GLU ARG VAL GLU ALA
 C G G C A G G T A A A C C G A G C G T G T G G A A G C G A 3740
 3730
 MET VAL GLN TYR THR TYR ARG LYS GLY LYS G
 T G G T G C A A T A T A C C T A C C G T A A A G G C A A A G 3760
 3770 3780

LU ASN LYS ALA HIS SER ASP LEU ASN GLY
 A A A C A A A G C A C A C A G C G A C C T A A A T G G C A 3800
 3790
 ILE ASN GLN SER LEU TYR ARG LEU GLY ALA T
 T C A A C C A A A G C C T A T A T C G C T T G G G T G C A T 3820
 3830 3840

RP GLN GLN LYS TYR ASP LEU ARG LYS PRO
 G G C A A C A A A A T A T G A T T T A A G A A A G C C T A 3860
 3850
 ASN GLU LEU PHE ALA GLY THR SER TYR ILE T
 A C G A A C T G T T T G C A G G C A C A G C T A T A T C A 3880
 3890 3900

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FIG.4V

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HR  GLU  SER  CYS  LEU  ALA  SER  ASP  ASP  PRO
CCGAAAGCTGTTTGGCAAGTGATGACCCAA
3910                                3920 3930
      LYS  SER  CYS  VAL  GLN  TYR  PRO  TYR  VAL  TYR  T
      AAGCTGCGTACAATAACCCCTTATGTC TACA
3940                                3950 3960

HR  LYS  ALA  ARG  PRO  ASP  GLY  ILE  GLY  ASN
CCAAAGCCCCGACCAAGATGGTATCGGCAATC
3970                                3980 3990
      ARG  ASN  PHE  SER  GLU  LEU  SER  ASP  ALA  GLU  L
      GCAATTTTCTGAGTTAAGCGATGCTGAAA
4000                                4010 4020

YS  ALA  GLN  TYR  LEU  ALA  SER  THR  HIS  PRO
AAGCAATAATTGGCGTCCACGCCACCCCC
4030                                4040 4050
      HIS  GLU  VAL  SER  ALA  LYS  ASP  TYR  THR  G
      ATGAGGTTGTCCTCTGCCCAAGATTATACAG
4060                                4070 4080

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FIG.4W

LY THR TYR ARG LEU LEU PRO ASP PRO MET
 GCACCTATCGGTTGTGTTACCTGACCCCATGG
 4090 4100 4110
 ASP TYR ARG SER ASP SER TYR LEU ALA ARG L
 ACTATCGTTCAGACTCGTATTGCGACGCC
 4120 4130 4140

EU ASN ILE LYS ILE THR PRO ASN LEU VAL
 TTAACATCAATAATCGCCCAATTTGGTCA
 4150 4160 4170
 SER LYS LEU LEU LEU GLU ASP THR LYS GLN T
 GTAAACTGTTATTAGAGACCAAGCAAA
 4180 4190 4200

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HR TYR ASN ILE ARG ASP MET ARG HIS CYS S
 CATACAATTCGTGATATGCGTCATTGTA
 4210 4220 4230
 ER TYR HIS GLY ALA ARG LEU GLY ASN ASP G
 GTTATCATGGGCGAAGATTGGGCAATGACG
 4240 4250 4260

FIG.4X

LY LYS PRO ALA ASN GLY GLY SER ILE VAL
 GTAGCCCTGCCCAATGGCGGCTCCATTGTCC
 4270 4280 4290
 LEU CYS ASP ASP TYR GLN GLU TYR LEU ASN A
 TTTCGGATGATTATCAAGAGTATCTAAATG
 4300 4310 4320

LA ASN ASP ALA SER GLN ALA SER PHE ARG
 CCAATGACGCATCAAGCATCTATTAGAC
 4330 4340 4350
 PRO GLY ALA ASN ASP ALA PRO ILE PRO LYS L
 CAGGGGCTAATGACGCCCCCATTCCAAAC
 4360 4370 4380

EU ALA TYR ALA ARG SER SER VAL PHE ASN
 TGGCTTATGCCAGAGCAGTGTTTAAAC
 4390 4400 4410
 GLN GLU HIS GLY LYS THR ARG TYR GLY LEU G
 AAGAGCATGGCAAAACTCGCTATGGGTAG
 4420 4430 4440

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FIG.4Y

LY PHE GLU PHE LYS PRO ASP THR PRO TRP
 G T T T G A G T T T A G C C T G A C A C G C C A T G G T
 4450 4460 4470
 PHE LYS GLN ALA LYS LEU ASN LEU HIS GLN G
 T T A A C A A G C A A A T T A A C C T A C A T C A A C
 4480 4490 4500
 LN ASN ILE GLN ILE ILE ASN HIS ASP ILE
 A A A T A T C C A A A T C A T T A A C C A T G A C A T T A
 4510 4520 4530
 LYS LYS SER CYS SER GLN TYR PRO LYS VAL A
 A A A A T C G T G C A G C C A A T A T C C C A A G G T G G
 4540 4550 4560
 SP LEU ASN CYS GLY ILE SER GLU ILE GLY
 A T T A A A T T G T G G C A T C A G T G A A A T T G G G C
 4570 4580 4590
 HIS TYR GLU TYR GLN ASN TYR ARG TYR L
 A T T A T G A A T A T C A A A C A A T T A C C G T T A T A
 4600 4610 4620

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FIG.4Z

YS GLU GLY ARG THR SER LEU THR GLY LYS
 AAGAGGGCGTACCAGTTTGACAGGCAAC
 4630 4640 4650
 LEU ASP PHE ASN PHE ASP LEU LEU GLY GLN H
 TTGATTTTAAATTTTGACCTGCTGGGCCAGC
 4660 4670 4680

IS ASP LEU THR VAL LEU ALA GLY ALA ASP
 ACGATTGACGGTGTTGGCTGGTGCAATA
 4690 4700 4710
 LYS VAL LYS SER GLN PHE ARG ALA ASN ASN P
 AAGTTAAAGCCAAATTCGTGCCAACACC
 4720 4730 4740

RO ARG ARG THR ILE ILE ASP THR THR GLN
 CCAGACGCACAATCATTTGACACCAACCAAG
 4750 4760 4770
 GLY ASP ALA ILE ILE ASP GLU SER THR LEU T
 GCGATGCCATCATTTGATGAAGCAGCTGA
 4780 4790 4800

FIG. 4A'

HR ALA GLN GLU GLN ALA LYS PHE LYS GLN
 CAGCACAGGAGCAAGCCAAATTTAAGCAAT
 4810 4820 4830
 SER GLY ALA ALA TRP ILE VAL LYS ASN ARG L
 CAGGGGCAAGCATGGATTGTCAAAATCGCT
 4840 4850 4860

EU GLY ARG LEU GLU GLU LYS ASP ALA CYS
 TAGGACGCTTAGAAGAAAGACGCGCTGTG
 4870 4880 4890
 GLY ASN ALA ASN GLU CYS GLU ARG ALA PRO I
 GCAATGCCCAATGAATGTGAACGGCGGCCCA
 4900 4910 4920

LE HIS GLY SER ASN GLN TYR VAL GLY ILE
 TTCATGGCAGTAACCAATAATGTGGGCATTA
 4930 4940 4950
 ASN ASN LEU TYR THR PRO ASN ASP TYR VAL A
 AACCTTTATACACCAATAATGATTATGTGG
 4960 4970 4980

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FIG.4B'

SP LEU SER PHE GLY GLY ARG LEU ASP LYS
 A T T A A G T T T G G T G G A C G C T T G G A T A A A C
 4990 5000 5010
 G L N A R G I L E H I S S E R T H R A S P S E R A S N I L E I
 A A C G C A T T C A C A G C A C C G A T T C A A A C A T C A
 5020 5030 5040

LE SER LYS THR TYR THR ASN LYS SER TYR
 T C A G C A A A C T T A C A C C A A A A G C T A T A
 5050 5060 5070
 A S N P H E G L Y A L A A L A V A L H I S L E U T H R P R O A
 A T T T G G A G C G G C G G T T C A T C T G A C A C C T G
 5080 5090 5100

SP PHE SER LEU LEU TYR LYS THR ALA LYS
 A T T T A G C C T G T T G T A T A A A C T G C C A A A G
 5110 5120 5130
 G L Y P H E A R G T H R P R O S E R P H E T Y R G L U L E U T
 G C T T T C G T A C G C C A A G T T T T A T G A A C T G T
 5140 5150 5160

FIG.4C'

YR ASN TYR ASN SER THR ALA ALA GIN HIS
 A C A C T A T A C A G C A C C G C C C A G C A T A
 5170 5180 5190
 LYS ASN ASP PRO ASP VAL SER PHE PRO LYS A
 A A A T G A C C C T G A T G T G T C T T T C C C A A A C
 5200 5210 5220

RG ALA VAL ASP VAL LYS PRO GLU THR SER
 G A G C G G T T G A T G T C A A A C C T G A A A C T T C C A
 5230 5240 5250
 ASN THR ASN GLU TYR GLY PHE ARG TYR GIN H
 A T A C C A A T G A A T A C G G C T T T C G C T A T C A G C
 5260 5270 5280

IS PRO TRP GLY ASP ILE GLU MET SER MET
 A C C C T T G G G G G A T A T T G A G A T G A G C A T G T
 5290 5300 5310
 PHE LYS SER ARG TYR LYS ASP MET LEU ASP L
 T C A A A A G C C G T T A C A A G G A C A T G T T A G A T A
 5320 5330 5340

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FIG.4D'

YS ALA ILE PRO ASN LEU THR LYS ALA GLN
 AAGCCATAACCGAACCTAACCAAGCCAGC 5350
 5360
 GLN GLU TYR CYS LYS ALA HIS LEU ASP SER A
 AAGAGTATTGTAGGCTCATTTGGATTCCA 5380
 5390 5400

SN GLU CYS VAL GLY ASN PRO PRO THR PRO
 ATGAATGTTGGTAATCCACCCAGCCCA 5410
 5420 5430
 LYS THR SER ASP GLU VAL PHE ALA ASN LEU T
 AACCAAGTGATGAGGTATTTGCCCACTTAT 5440
 5450 5460

YR ASN ALA THR ILE LYS GLY VAL SER VAL
 ATAATGCCACCATCAAGGGGTGAGTGTC A 5470
 5480
 LYS GLY LYS LEU ASP LEU HIS ALA MET THR S
 AAGGCAAACTGGATTGCA TGCCATGACAT 5490
 5500 5510 5520

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FIG.4E'

ER LYS LEU PRO ASP GLY LEU GLU MET THR
 C A A A C T G C C A G A T G G T C T T G A A A T G A C C T
 5530 5540 5550
 LEU GLY TYR GLY HIS THR LYS LEU GLY LYS P
 T G G G T T A T G G T C A T A C C A A A T T G G G G A A A T
 5560 5570 5580
 79/130
 HE ASP TYR ILE ALA PRO LYS ASP ALA ASP
 T T G A T T A C A T T G C A C C C A A A G A T G C C G A T G
 5590 5600 5610
 GLY TRP TYR GLN ALA ARG PRO ALA PHE TRP A
 G T T G G T A T C A G G C T C G C C C T G C T T T T G G G
 5620 5630 5640
 SP ALA ILE THR PRO ALA ARG TYR VAL VAL
 A T G C C A T C A C C C C A G C G C C T A T G T G T C G
 5650 5660 5670
 GLY LEU ASN TYR ASP HIS PRO SER GLN VAL T
 G T C T A A A C T A T G A C C A C C C C A G T C A G T A T
 5680 5700

FIG. 4F

RP GLY ILE GLY THR THR LEU THR HIS SER
 GGGCA TTGGCA CAACTTTAACGCACAGCA
 5710 5720 5730
 LYS GLN LYS ASP GLU ASN GLU LEU SER ALA L
 ACAA AAGATGA AATGAGCTAAGTGCCC
 5740 5750 5760

EU ARG ILE ARG ASN GLY LYS ARG GLU ILE
 TAGAATCCGAAATGGCA AAGAGAAATAC
 5770 5780 5790
 GLN THR LEU THR HIS THR ILE PRO LYS ALA T
 AACCTTAACGCACACAAATACCCCAAAGCCT
 5800 5810 5820

YR THR LEU LEU ASP MET THR GLY TYR TYR
 ATACCTTACTGGACATGACAGGCTATTATA
 5830 5840 5850
 SER PRO THR GLU SER ILE THR ALA ARG LEU G
 GCCCAACTGAGAGCATCACCGCTCGCTTG
 5860 5870 5880

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FIG.4G'

LY ILE ASN ASN VAL LEU ASN THR ARG TYR
 G T A T C A A C A A T G T A T T A A A C A C C C G C T A
 5890 5900
 THR THR TRP GLU ALA ALA ARG GIN LEU PRO S
 C A C C A C A T G G G A A G C G G C A C G C C A A C T G C C C A
 5910 5920 5930 5940
 ER GLU ALA ALA SER SER THR GIN SER THR
 G C G A A G C T G C A A G C A G T A C C C A A T C A A C C C
 5950 5960 5970
 ARG TYR ILE ALA PRO GLY ARG SER TYR PHE A
 G T T A C A T T G C A C C A G G T C G C A G T T A C T T T G
 5980 5990 6000
 ORF 3
 LA SER LEU GLU MET LYS PHE *** MET THR
 C C A G T C T T G A A A T G A A G T T T A A T A T G A C C
 6010 6020 6030
 CYS LEU PRO LYS THR ASN PRO ALA LEU LYS
 T G T T A C C A A G A G A C C A C C C T G C T T T A A A
 6040 6050 6060

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FIG.4H'

VAL LYS HIS ARG PHE LEU LYS GLN VAL LEU
 GTCAAGCACAGATTTTTAAAGCAGGTGCTG
 6070 6080 6090
 LEU LEU LEU CYS VAL ASP THR LEU THR ALA
 TTTATTGCTTTGTTGTTGATTATTAACAGCA
 6100 6110 6120

GLN ALA TYR ALA HIS SER HIS HIS THR PRO
 CAGGCGTACGCCCAACAGCCATCATACGCCC
 6130 6140 6150
 ILE HIS THR PRO THR HIS GLU LEU SER SER
 ATTCAATACCCACGCCATGAGCTGTCTCT
 6160 6170 6180

ALA ASP ALA LEU SER ASP GLU GLY LEU GLY
 GCTGATGCTTTATCAGATGAGGCTTGGGT
 6190 6200 6210
 LYS ASP LEU GLY SER LEU ASP SER PRO ASP
 AAGGATTGGGCGAGTTTGGACAGCCCAT
 6220 6230 6240

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FIG. 4I'

GLY LEU GLY ASP GLY LEU GLY ASP GLY LEU
 GGT TGG GTG ATGGT TTAGGCG ATGGT TTG
 6250 6260 6270
 GLY ASP GLY LEU LYS SER ASP LYS THR PRO
 GGTGATGGCTTAAAGTGA TAAACCCCT
 6280 6290 6300

LEU PRO ILE ASN ALA LEU THR VAL ASN GLN
 TACCCATCAACGCCCTTGACCGTTAATCAG
 6310 6320 6330
 SER ASN GLU SER GLN PRO ALA PRO PRO SER
 AGCAATGAGAGCCAGCC TGCCCA CCGAGC
 6340 6350 6360

VAL ASP VAL ASN PHE LEU LEU ALA GLN PRO
 GTAGATGTCAATTTT TTA CTTGCCCA GCCA
 6370 6380 6390
 GLU ALA PHE TYR HIS VAL PHE HIS GLN ALA
 GAGGCATTTTATCATGTCTTCTTCA TCAAGCG
 6400 6410 6420

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FIG. 4J'

ILE VAL GLN ASP ASP VAL ALA THR LEU ARG
 A T G T G C A A G A T G A T G T G G C A A C A T T A C G C
 6430 6440 6450
 LEU LEU LEU PRO PHE TYR ASP ARG LEU PRO
 T T G T T A T T G C C A T T T T A T G A C C G C C T G C C T
 6460 6470 6480

ASP ASP TYR GLN ASP ASP VAL LEU LEU LEU
 G A T G A T T A T C A A G A T G A T G T T T G T T G T T A
 6490 6500 6510
 PHE ALA GLN SER LYS LEU ALA LEU SER ASP
 T T T G C C C A A A G T A A A C T T G C C C T A A G T G A T
 6520 6530 6540

GLY ASN THR LYS LEU ALA LEU ASN LEU LEU
 G G C A A T A C C A A A T T G G C A T T G A A T C T G C T G
 6550 6560 6570
 THR ASP LEU SER ASN LYS GLU PRO THR LEU
 A C C G A T T T G A G T A A C A A A G A G C C A C A C T T
 6580 6590 6600

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85/130

FIG.4K'

THR ALA VAL LYS LEU GLN LEU ALA SER LEU
 ACGCGGTAAATAATTACAACTTGCTTCCCTTG
 6610 6620 6630
 LEU LEU THR ASN LYS HIS ASP LYS HIS ALA
 TTGCTGACCAACAAGCACGATAAACACGCC
 6640 6650 6660

GLN MET VAL LEU ASP GLU LEU LYS ASP ASP
 CAATGGTGCTAGATGAACTCAAGATGAT
 6670 6680 6690
 ALA HIS PHE LEU LYS LEU SER LYS LYS GLU
 GCCACCTTTTATAAATTAAAGCAAAAGAG
 6700 6710 6720

GLN ARG TRP VAL LEU SER GLN SER ARG TYR
 CAAAGATGGGTGCTATCGCAAGTCGCTAT
 6730 6740 6750
 LEU HIS LYS LYS TYR LYS MET GLY LEU ASP
 TTACATAAATAATATAAATGGGCTTGGA
 6760 6770 6780

FIG. 4L'

LEU GLY ILE ASN TYR LEU HIS LEU ASP ASN
 TTGGGCATCAACTATCTGCAATTGGATAAT
 6790 6800
 ILE ASN ALA ALA SER THR ILE THR GLN PRO
 ATCAACGCCCGCCTCCACCATCACCCAGCCC
 6820 6830 6840

ASN ILE LYS LYS ASP ALA PRO LYS PRO ALA
 AACATTAAAGATGCCCCAAACCTGCT
 6850 6860 6870
 HIS GLY LEU ALA LEU SER LEU GLY VAL ASN
 CATTGGGCTTGCCCTTATCGCCTTGGTGTGTAAT
 6880 6890 6900

LYS TYR THR PRO LEU SER HIS GLY MET SER
 AATACACGCCGCTTAGTCA TGGCATGAGT
 6910 6920 6930
 ILE TYR THR ALA LEU ASP VAL ASP GLY LYS
 ATTTATACAGCCCTAGATGTGTGTTGTTAA
 6940 6950 6960

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FIG. 4M'

PHE TYR ASP ASP LYS SER HIS ASN GLU LEU
 TTTTATGATGACAAAGCCACAAATGAACTG
 6970 6980 6990
 ALA VAL PHE ALA HIS ALA GLY LEU ARG LYS
 GCGGTTTTTTCCTCATGCTGGACTAAGAAA
 7000 7010 7020

ASP HIS GLN LYS GLY TYR VAL ASP VAL VAL
 GATCACCAAAAGGTTATGTTGATGTCGTA
 7030 7040 7050
 PRO PHE VAL GLY ARG ILE PHE ALA THR ASN
 CCTTTTGTGCGCGTATTTTGGCCACCAAT
 7060 7070 7080

GLN GLN HIS GLY ARG LEU SER PRO ARG LYS
 CAGCAGCATGGCAGATTATCCCCAGAAA
 7090 7100 7110
 ASP SER GLN GLY VAL ALA PHE GLY SER HIS
 GACAGTCAGGCGGTGGCGTTTGGCAGCAT
 7120 7130 7140

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FIG.4N'

HIS ARG ILE ASN ASP LYS TRP GLN ASN ALA
 C A T C G G A T C A A T G A T A A A T G G C A A A A T G C G
 7150 7160 7170
 PHE PHE ALA ARG MET GLU LYS GLY ASN TYR
 T T T T T T G C A C G C A T G G A A A A G G C A A T T A T
 7180 7190 7200
 THR GLU HIS TYR GLN GLY TYR ASP GLY LYS
 A C C G A G C A T T A T C A A G G T T A T G A T G G C A A G
 7210 7220 7230
 ARG TYR HIS VAL ASN ASP THR ILE LEU LEU
 C G T T A T C A T G T G A A T G A C A C C A T T T G T T G
 7240 7250 7260
 GLN ASP GLY PRO ASN ARG ARG TYR SER LEU
 C A A G A T G G C C C A A A T C G T C G T T A C T C T T T G
 7270 7280 7290
 GLY VAL GLY TYR GLN LEU SER HIS LEU GLN
 G G C G T G G G G T A T C A G C T T A G C C A T C T G C A A
 7300 7310 7320

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FIG. 4O'

ASP ALA THR LYS SER SER HIS ALA THR LYS
 G A T G C A A C A A A G C A G T C A T G C C A C A A A G
 7330 7340 7350
 ILE HIS PHE GLY VAL LEU GLN ARG LEU PRO
 A T A C A T T T T G G G G T G T T G C A A A G A T T G C C A
 7360 7370 7380

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ASN GLY LEU THR VAL GLN GLY ARG VAL SER
 A A T G G T C T G A C C G T G C A A G G T A G A G T G A G T
 7390 7400 7410
 ALA GLU ARG ARG TYR HIS GLY LYS LEU
 G C T G A G C G T G A G C G T T A T C A T G G T A A A T T A
 7420 7430 7440

LEU ARG LEU VAL ASN PRO ASP ASP VAL TYR
 T T G C G T C T G G T T A A T C C C T G A T G T G T A T
 7450 7460 7470
 ARG THR ASP LYS THR LEU THR LEU GLN THR
 C G C A C A G A T A A A C C C T A A C C C T A C A A C C
 7480 7490 7500

FIG.4P'

SER ILE TRP HIS LYS ASP ILE HIS TRP LEU
 TCCATTGGCCACAAAGACATTCACTGGCTT 7510
 7520
 GLY LEU THR PRO LYS LEU THR TYR ARG TYR
 GGATTACGCCCAAAGCTGACTTATCGTTAC 7530
 7540 7550 7560

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 SER LYS ASN ASN SER ASN LEU PRO ALA LEU
 AGTAAATAACAGTAACCTTACCGACTT 7570
 7580 7590
 TYR SER HIS ASN LYS LYS GIN ASN PHE TYR LEU
 TATAGCCATAACAACAATAATTTATTG 7600
 7610 7620

GLU LEU GLY ARG SER PHE ***
 GAGCTTGGTCGGTCGTTTTTAA 7630
 7640

1bpB 1bpA orf3



FIG. 5A

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EcoRI	_____!
EcoRII	_____!_____!
EcoRV	_____!_____*
FokI	_____!_____!_____!
HaeII	_____!_____
HaeIII	_____!_____!_____!
HhaI	_____!_____!_____!_____*
HindIII	_____!_____!_____!
HpaI	_____!_____
NcoI	_____!_____!_____!
NdeI	_____!_____!_____!
PvuII	_____!_____!_____!_____!
ScaI	?!_____?
ScrFI	_____!_____!_____!
SphI	_____!_____!
SspI	_____!_____?_____!_____!
XmnI	_____!_____*

FIG. 5B

FIG.6A

Alignment of Lbp2 proteins

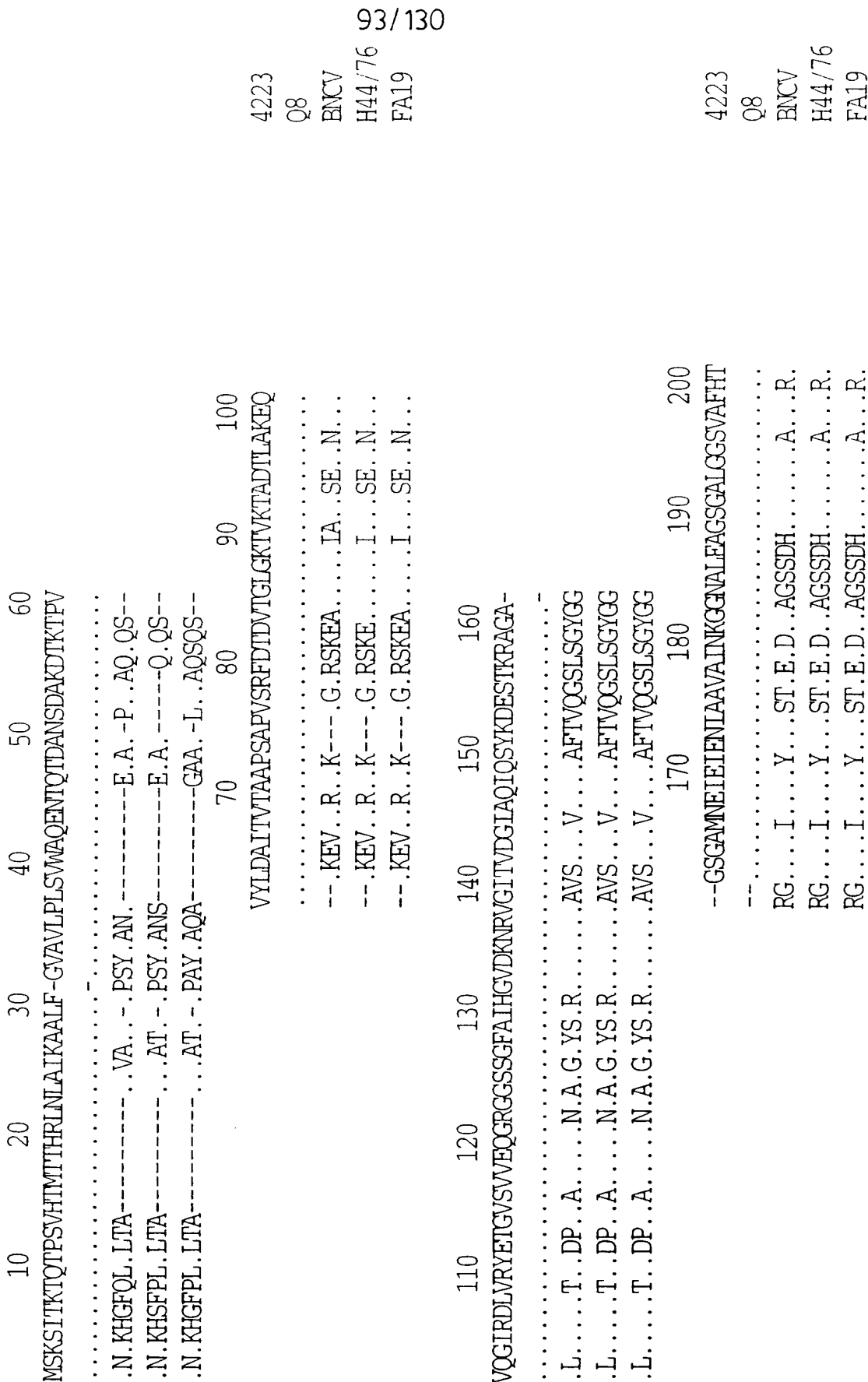


FIG. 6B

210	220	230	240	250	260		
KDVSDVLKSGKNLGAQSKITTYNSKNDHFSQTLAAAGKTERVEAMVOYTYRKKGKKNKAHSDL							
.....N.....							
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.THP.G.I							
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.TRP.G.I							
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.TRP.G.I							
			270	280	290	300	
			-NGINQSLYRLGAWQKYDL-RKPNELFAGTSYTTESCLAS				4223
			-.....T.....				Q8
			AD.VAYGIN..D.FR.T.GI-K..S.GGEYFLAEG..E.KP				BNCV
			AD.VAYGID..D.FR.T..I-Q.QNKKAIFYLAEG..E.KP				H44/76
			AD.VAYGID..D.FR.T..IK..TT.P.--FLAEG.NT.KP				FA19
							94 / 130
310	320	330	340	350	360		
DDPKSCVQYPYVTKARPDGIGNRNFSELSDAEKAQYLAETHPEWWSAKDYTGIRLLPD							
.....T.....							
VAKVAGNGNYLANQLN.WKERIEQNQP..AE.E.MVREAQAR..NL..QA...GG.I...							
AAKLACNGNYLANQLN.WFEERKNNQS..AE.E.MVREAQAR..NL..QA...GG.I...							
VAKLAGYGIYLANQLN.WKERIEQNQP..AE.E..VREAQAR..NL..QA...GG.I...							
			370	380	390	400	
			PMDYRSDSYLARLNKITYPNLVSKLLLEDIKQTYNIRDM				4223
						Q8
		G.W..K.GYRFGGRHYVGGVF.....R.D....				BNCV
		G.W..K.GYRFGGRHYVGGVF.....R.D....				H44/76
		G.W..K.GYRFGGRHYVGGVF.....R.D....				FA19



FIG.6C

410	420	430	440	450	460		
RHCSYHGARLGNCGKIPANGSIVLCDDYQEIYINANDASQALFRPGANDAPIPKLAYARSSV							
.....S.....							
TEKQ.Y.TDEAKKFRDKS.—VYDG...FRDG.YFVPNIEE—WKGDQKLRGIG.K.S.TK—							
TEKQ.Y.TDEATKFSDKS.—VYDG...FRDG.YFVPNIEE—WKGDK.LVKGIG.K.S.TK—							
TEKQ.Y.TDEAEKFRDKS.—VYDG...FRDG.YFVPNIEE—WKGDK.LVKGIG.K.S.TK—							
			470	480	490	500	
			FNQEHCKTRY—GLSFEF---KPDTPWFKQAKINLHQQNIQIIN				4223
		G.....				Q8
			.ID..HRR.RM..LYRYENE.YSDN.ADK.V.SFDK.GVATD.				ENCV
			.ID..HRR.RM..LYRYENEAYSIN.ADK.V.SFDK.GVATD.				H44/76
			.ID..HRR.RM..LYRYENE.YSDN.ADK.V.SFDK.GVATD.				FA19
							95/130
510	520	530	540	550	560		
HDIKKSCSQYPKVDLNCGISEIGHYEQ—NNRYKEGRASLTGKLDNFNFDL—LGQHDLITVLG							
.....T.....							
NTL.IN.AV..A..KS.RA.ADKP.S.DSSDRFH.R.QHNV.NASFEKSLKNKWIK.H..LGF.							
NTL.IN.AV..A..KA.RA.ADKP.S.DSSDRFH.R.QHNV.NALFEKSLKNKWIK.H..LGF.							
NTL.IN.AV..A..KA.RA.ADKP.S.DSSDRFH.R.QHNV.NASFEKSLKNKWIK.H..LGF.							
			570	580	590	600	
			ADKVKSQFRANNPRRTIIDTTQGDALIDESTLTAQEQAK				4223
						Q8
			Y.ASNAIS.PEQLSHNAARISEYSDYT.KGD-----				ENCV
			Y.AS.AIS.PEQLSHNAARISEFSDYA.DGKY-----				H44/76
			Y.AS.AIS.PEQLSHNAARISE—STGF..KNQD-----				FA19

710	720	730	740	750	760
TAKGFRTPSFYELVYNSTAQHKNDPVSF	PKRAVDVKPETSNTINEYGF	FRYQHPWG	DVEM		
.....
ASS.....Q..	FGIDIYH-----	YPKGQRPAL.S.KAANR.I.	LQWKGDF.FL.I		
ASS.....Q..	FGIDIYH-----	YPKGQRPAL.S.KAANR.I.	LQWKGDF.FL.I		
ASS.....Q..	FGIDIYH-----	YPKGQRPAL.S.KAANR.I.	LQWKGDF.FL.I		
		770	780		
		SMFKSRYKMLDKAIPNLTKAQ-QE			
	
		.S.RN..T..IAV.DHK-	.LPN.A		
		.S.RN..T..IAV.DHK-	.LPN.A		
		.S.RN..T..IAV.DQK-	.LPDSM		

NSDOCID: <WO__9855606A2_1_>

FIG. 6E

[illegible]

Alignment of M. catarrhalis Lbp2 proteins

110	120	130	140	150
FFVMGKIRDTSEKNDPDYSDDLKQW----	LGKLYVGIDAHRPDGIGCKNLRQPITAND			
.....P.....N.V.....Q.....T.....				
.....TK..QGSVHTAGQ.LQ.L.TKEP.....T.T.....D.				
	160	170	180	190
	200			
	IKPLYFNKFPALSDLHLDSEHRHFDQKINTIKVGYGNLTTPS			
K.L.....			
	.T....D...KI.....ENSE.V.AK.A.N.I....A.SS.A			

210	220	230	240	250	260
NNNTHNHQQA	DNKKNNKPV	DPYENIRFG	YLELQGS	SLTQKNAD	QNEQDR-IPKPMPILF
K...Y.....	P.DK.-.....
K.P.YM.Y.EQ	I.K.G.D.Q.....	M.RELD.NK.G.....	SDKN.A.IFTT.T..
		270	280	290	300
		YHGENASSQLPSAGKFN Y TG N MLYLS D VKKR P ALSASDER			
	D.			
	TH.K...D.E.....T.....F.DKT.DK			

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FIG.7B

310	320	330	340	350	360	
VGVLNASC	NEGDVSA	HIYNGFQYK	HTPATYQ	VDFTINS	LTKLSYD	NPQNN
.....S	TA
T.F.S	R.S.LS.KS.Q.T.KK.TA		
		370	380	390	400	
		KGEYLK	SQFD	TIKKVNE	TDVYQIDAKING	RFVGTAKSLV
		Q.K.I
		D.R.IRDAET
					I
410	420	430	440	450	460	
NEKTQ	TAP	TIKEL	FSKKAN	PNPNP	NSD	ILEGGFYGESG
..N.E
DIN.NVDTF		
		470	480	490	500	
		RDKTTK	PVATK	TVVFS	SAGFEK	PSTSFVDNETIGGI
	DRNS.K
	ETGES
					G
510	520	530	540	550		
LN----	NHNEDE	IIP-SDD	SYGYTWG	KEKQFTKKV	SSSQVWPAYFGQ	HDKFYFN
..DAVNEK	DNCD	PT-..ER	DEFPEKKAEA
..DEWN.Q	..TV.V	NKE..E	NY.R.NINA.V	KN
		560	570	580	590	600
		YYDLSAS	RVDKL	APADAV	KANQSIKE	KYPNATINKUNQVTA
	S
	KEAN	..GVSQ	DTST.K	..LA..D	..KVST..K..K....

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FIG.7C

610 620 630 640 650 660
EAKDKPYTAIRAKSYQHISFGETLYNDANQTPTRSYFVQGRADTSTILPQAGKFTVNG

.....H.....D.....V.....NKGK.....Q.V.Q.S.....K.....

LK.I.TAFAD-I..T..AR.T.

670 680 690 700

LWAGYLITQKKDKGYSDNAETIKKEKHPGYLLTENFTPEDD

.....I.....N.E.....K...QD.....D.....

.....KDED...Q..LKD.I..KD.I.Q..

T.EARISKPIQWNNHADKKA-----

Q8
4223
VH19
H44/76

710 720 730 740 750

DDD---LTASDDSDNNIHGDDDLIASDDSQDDADGDDSDDLGCGADDDAAGKVYHA

....DA.....A.....

...DDS.....T.....

760 770 780 790 800

GNIRPEFENKYLPINEPHEKTFALDGKNKAKFEVDENINSLTG

.....D...D.....

.....D.N.D.....

-----E.D...GEK.IS.

Q8
4223
VH19
H44/76

FIG.7D

810 820 830 840 850
 KINDERGIV-FDIKNGKIDGIGTAKADVNYREEV-GNQGGFLYNIKIDIVKG

T.TEKN.VQPA.H.E..V.E.N..H.T.RTRDNGINLS..DSTNPPSFKANLL.T.
 RDNGINLS..GSTNPPSFKANLL.T.

860 870 880

QFFGTNGEELAGQLHHDKGD--GINDTAEKA-----
QY.....
 R.....T.....
 G.Y.PKA...G.IIFND.KSL..TEGT.NKVE-ADVDDVDVDVD
 G.Y.PQA...G.IIFND.KSL..TEDT.NEAE-AEVEAEAGVG--
 G.Y.PQAA..G.IIFND.KSL..TEDI.NEVEAEADV-----

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 Q8
 4223
 VH19
 BNCV
 H44/76
 FA19

890
 -----GAVEGAVKD-----K*
 -----*
 -----*
 ADADVEQLKPEVKPQF.V...K..NKEVE.*
 -----EQLKPEAKPQF.V...K..NKEVE.*
 -----EQLEPEVKPQF.V...K..NKEVE.*

Q8
 4223
 VH19
 BNCV
 H44/76
 FA19

FIG.8A *M. catarrhalis* strain 4223 Tbp2/Lbp2 comparison

<p>MKHIPLT--TLCVAISA---VLLTACGGSGS-NPPAPT-----PIPNASGSGNTG MSTVKTP..FYQKR..SL..ASIFAALVM.G.RSDDI.V.A.NV.QLPQGTVS...TGH-..N ** *</p>	Tbp2 Lbp2
<p>NTGNAGG-TDNTANAGTGGTNSGTGSANTPEPKYQDVPTKNEKDKVSSIQEPAMG-----YGM ..N.Q.NN...STSTDPN.D.NQLTQ.QKTAAGFFVMG.I-R.TSPKNDPDYSNDLVQQWQG ** *</p>	Tbp2 Lbp2
<p>ALSKINLHNRQDTPLDEKNIITLDGKKQVAEGKKSPLPFSLDVENKLLDGYIAKMNVDKNAIGD K.YVGIDAH.P.GIGTG..LRQITANDIKPLYFNKF.ALS.L-----HL.SERHRF *</p>	Tbp2 Lbp2
<p>RIKKGKEISDEELAKQIKEAVRKSHEFQQVLSLENKIFHSNDGTTKATTRDLKYVDYGYLAN DP..L.TIKVYGYGNLTTPSKNNTYINH..ADNKKN..PVDPYENIRFGYLELQGSSTQKNADT ** *</p>	102/130 Tbp2 Lbp2
<p>DGNYLTVKTDKLNLPVGVGFYNGTTAKELPTQDAVKYKGHWDFMTDVANRRNRFSEVKENSQ PNDKDRIPK-----MPIL..H.ENASSQ..SAGKFN.T.N.LYLS..KK.PALSASDDRV-- *</p>	Tbp2 Lbp2
<p>AGWYYGASSKDEYNRLLTKEDSAPDGHSGEYGHSSSEFTVNFKEKLTGKL-----F -.V.LN..G.SNEGDVVSAAHIYLN.FQYKHTPAT-YQ.D.DTNS...SYDNPQNQTAQGY *</p>	Tbp2 Lbp2
<p>SNLQDRHKGNVTKTERYDIDANIHGNNRFRGSATA-----SNKNDTSKHPFTSDAN-----NR IKS.FDTTKK.NE.DV.Q...K.N...V.T.KSLVNENTETAPFI.EL.SKK..PNNPNPNSDT *</p>	Tbp2 Lbp2

FIG. 8B

LEGGFYGPKEELAGKFLTNDNKLFGVFGAKRESKAEKTEAILDAYALGTFNTSNATTTTFTE Tbp2
ES.D.....S...ASYV...G..DKTDKPVATKTVYFS.AFEKP----- Lbp2

Protein	Sequence
Tbp2	KQLDNFGNAKKLVLGSTVIDLVPTDKNEFTKDKPESATNEAGETLMVNDEVSVKTYGKNFEYL
Lbp2	-----

KKFGELSIGGSHSVFLQGERTATTGEKAVPTTGTAAYLGNWVGYYITGKDTGTGTGKSFTDAQDVAD	Tbp2
-----	Lbp2
	103/ 130
FFDIDFGNKSVSGKLI TKGRQDPVFSITGQIAGNGWGTGTASTTKADAGGYKIDSSSTGKSI AIKDA	Tbp2
----- . IKNG . . . GTGFTAKADVPNY	Lbp2

-----NVTGFGYPNAN-EMGGSFTHN-----ADDS-----KASVVFGTKRQQEVK * Tbp2
PREEVGN.QG...LYNIKDIDVK.Q.FGTNGEEL.GQLQYDKGDINDTAE..GA...AVKD---. * Lbp2

* * * * *

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FIG.9A Alignment of *M. catarrhalis* Lbp3 sequences

10	20	30	40	50	4223
MTCLPKTNPALKVKHRFLKQVLLLCVDTLTAQAYAHSHHTPIHTPTHEL					Q8
.....	
60	70	80	90	100	4223
PSADALSDEGLGKDLGSLDSDSPDGLDGLDGLDGLKSDKAPLPINA					Q8
S.....	
.....	
110	120	130	140	150	4223
LTAHQTNESQPAPPSVDVNFLLAQPEAFYHVFHQAIQVQDDVATLRLLLPF					Q8
..VN.S.....	
.....	
160	170	180	190	200	4223
YDRLLPDDYQDDVLLLFAQSKLALSDGNTKLALNLLTDLNKEPTLTAVKL					Q8
.....	
210	220	230	240	250	4223
QLASLLLTNKHDKHAQMVLDELKDDAHFLKLSKKEQRWVLSQSRYLHKKY					Q8
.....	
260	270	280	290	300	4223
KMGLDLGINYLHLDNINAASTITQPNIKKDAPKPAHGLALSLGVNKYTPL					Q8
.....	
310	320	330	340	350	4223
SHGMSIYTALDVGKIFYDDKSHNELAVFAHAGLRKDHQKGYVDVVPFVGR					Q8
.....	



FIG.9B

360	370	380	390	400	
IFATNQHGRLSPRKDSQGVAFGSHHRINDKWQNAFFARMEKGN YTERYQ					4223
.....	H..	Q8
410	420	430	440	450	
GYDGKRYHVNDTILLQDGNRRYSLGVGYQLSHLQDATKSSHATKIHFV					4223
.....	Q8
460	470	480	490	500	
LQRLPNGLTVQGRVSAERERYHGKLLRLVNPDDVVRTDKTLTLQTSIWHK					4223
.....	Q8
510	520	530	540		
DIHWLGLTPKLT YRYSKNNSNLPALYSHNKQNFYLELGRSF*					4223
.....	*	Q8

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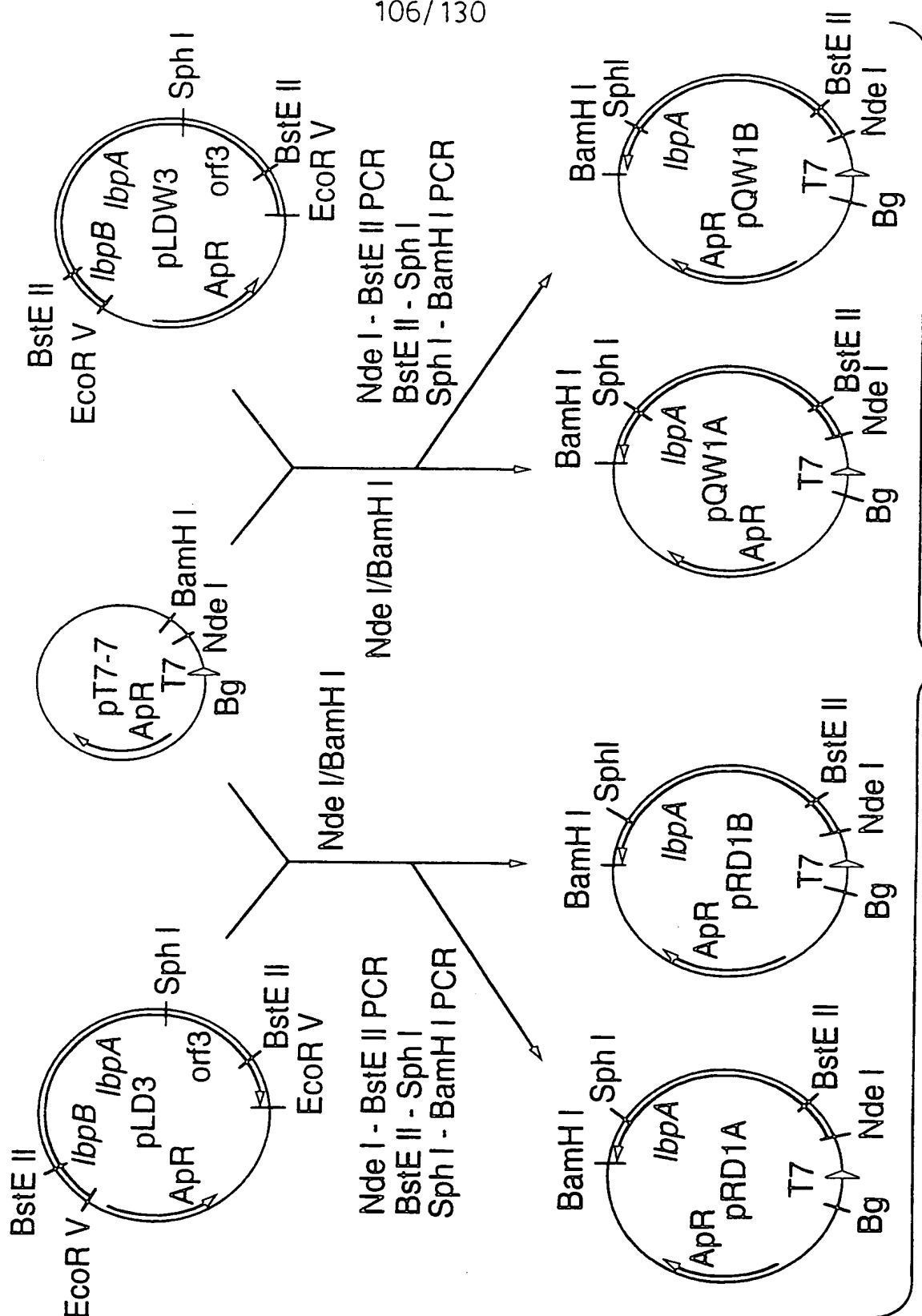


FIG.10

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EXPRESSION OF Q8 AND 4223 rLbp 1 PROTEINS.

FIG. 11A.

Q8 rLbp1

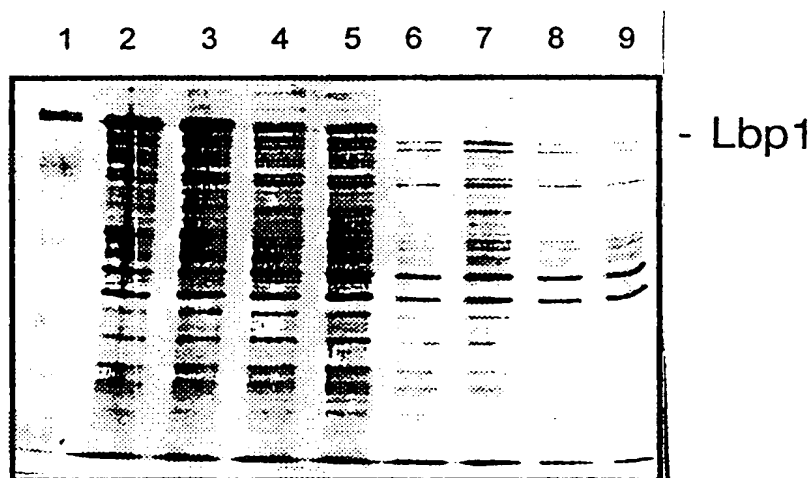
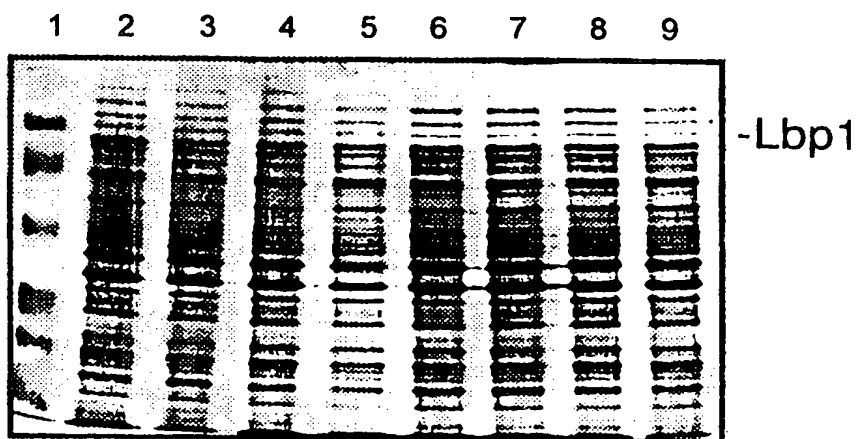


FIG. 11B.

rLbp1



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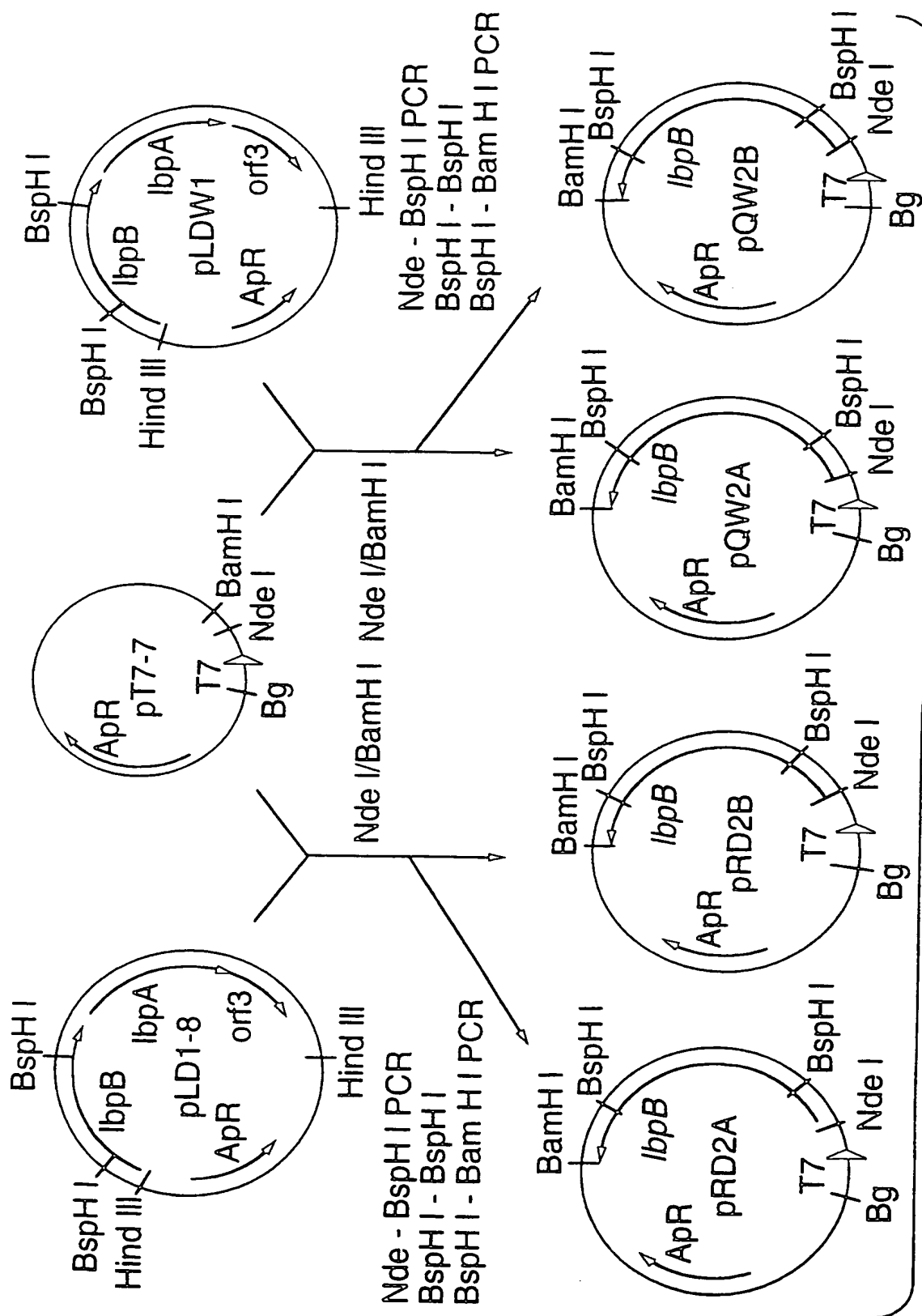


FIG.12

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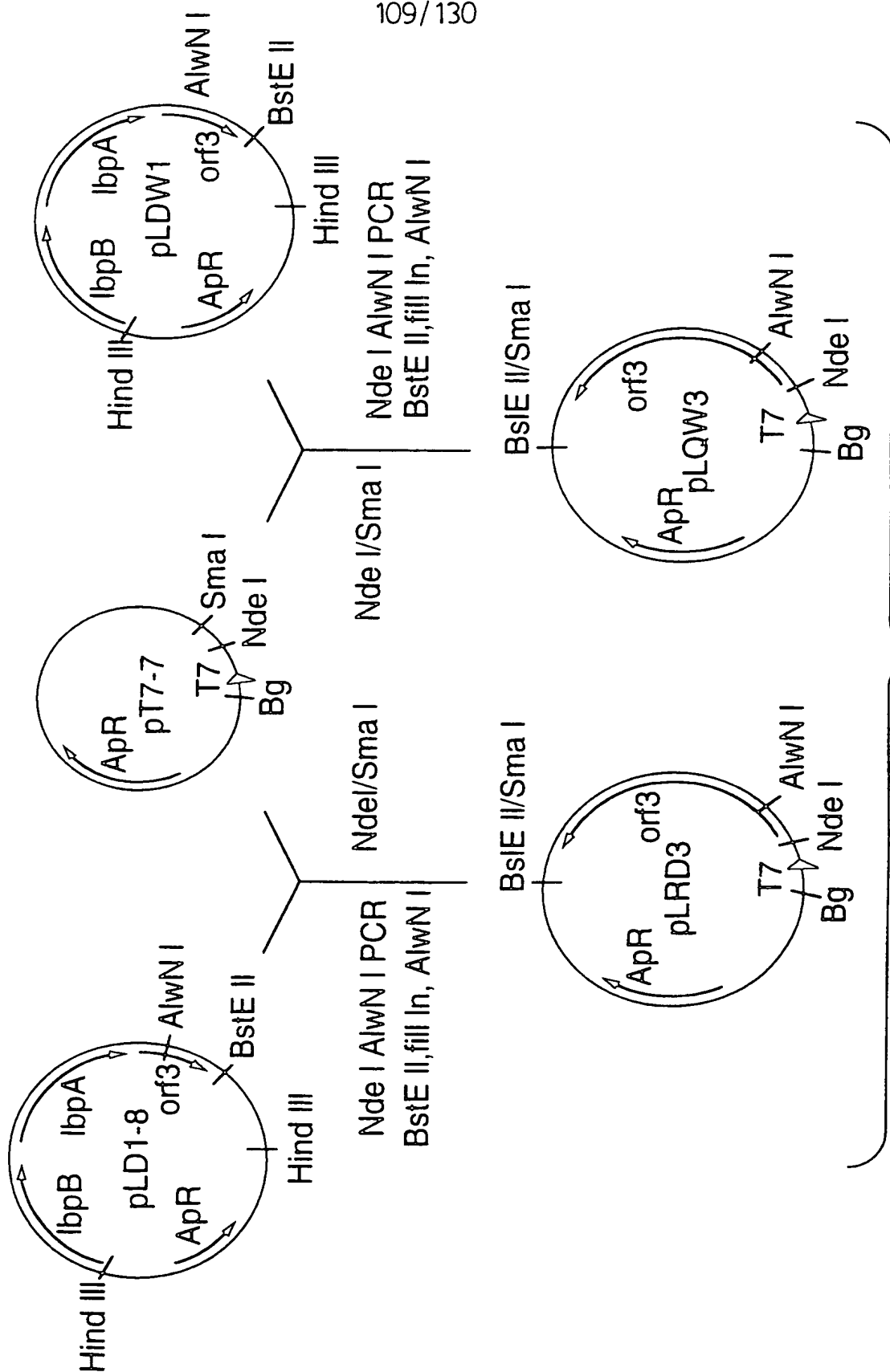
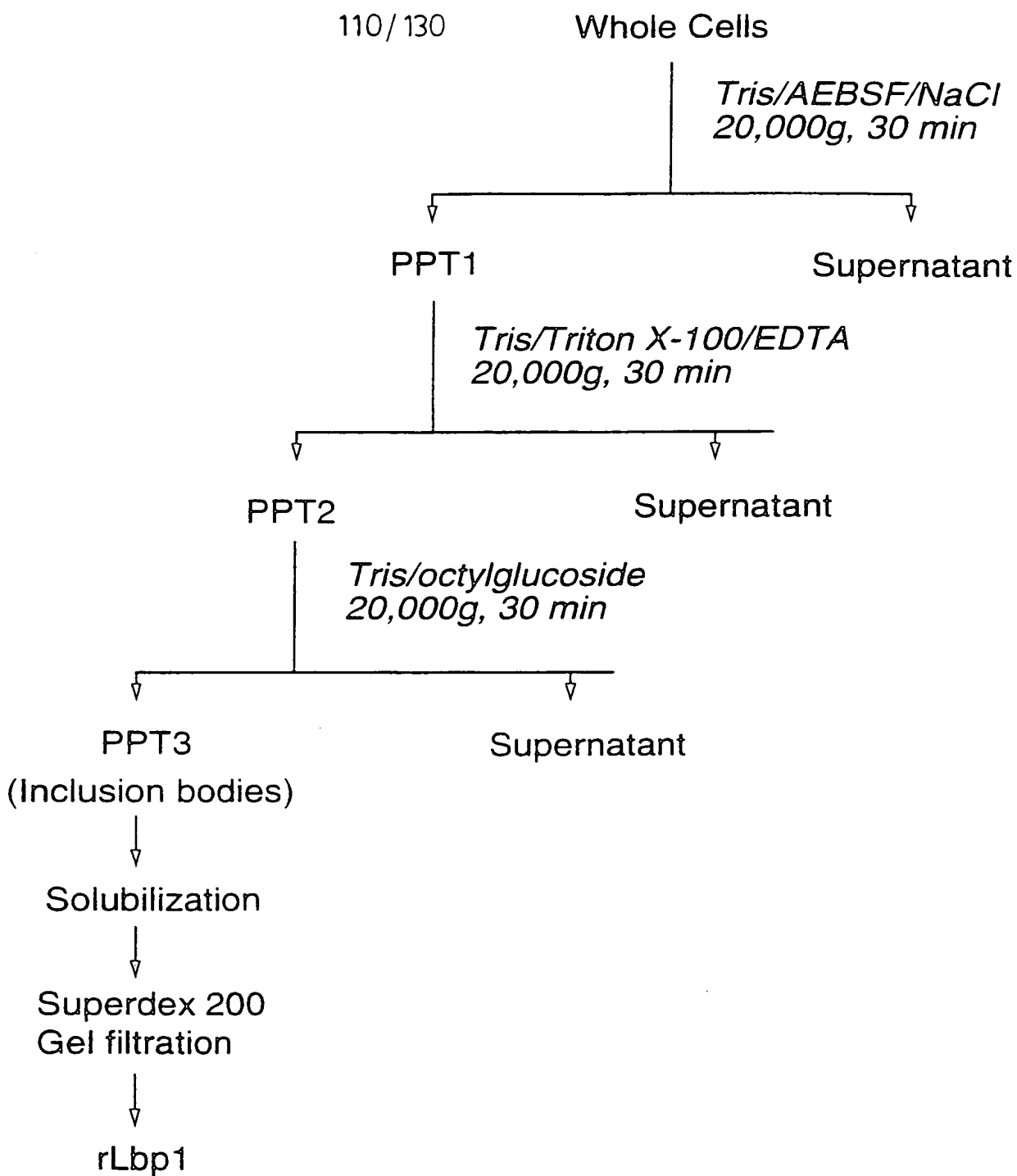


FIG. 13

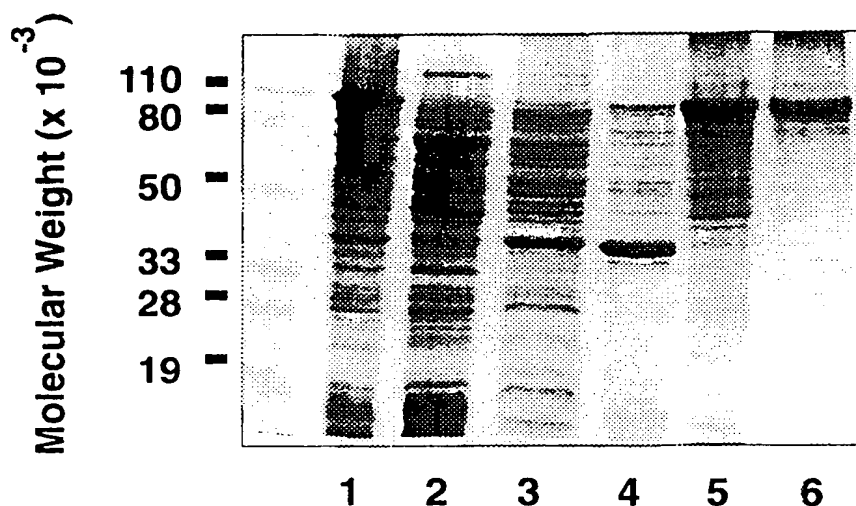


Purification scheme for rLbp1 expressed from E.coli

FIG.14

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Purification of Q8 rLbp1 from *E. coli*



1. *E. coli* whole cells
2. Soluble proteins in 50 mM Tris/ NaCl extraction
3. Soluble proteins in Tris/ Triton X-100 extraction
4. Soluble proteins in Tris/ octylglucoside extraction
5. rLbp1 inclusion bodies
6. rLbp1

FIG.15.

FIG. 16A

Nucleotide and deduced amino acid sequence of the VH19 lbpB gene.

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```

MET SER THR VAL LYS VAL PRL HIS ILE PHE
A T G A G T A C T G T C A A A G T C C C C C A C A T T T T C
10 20 30
TYR GLN LYS ARG THR LEU SER LEU ALA ILE
T A C C A A A A C G C A C C C T T A G C C T T G C C A T C
40 50 60
ALA SER ILE PHE ALA VAL VAL MET THR
G C C A G T A T T T T G C T G C C G T G G T G A T G A C A
70 80 90
GLY CYS ARG SER ASP ASP ILE SER VAL ASN
G G C T G C C G C T C T G A T G A C A T C A G C G T C A A T
100 110 120
ALA PRO ASN VAL THR GLN LEU PRO GLN GLY
G C A C C C A A T G T T A C C C A A C T G C C C C A A G G C
130 140 150
THR VAL SER PRO ILE PRO ASN THR GLY HIS
A C G G T T T C A C C A A T A C C G A A C A C A G G T C A T
160 170 180
```


FIG.16B

ASP ASN THR ASN ASN THR ASN ASN GLN GLY
 G A C A A C C C A A T A C A C C A A C A A T C A G G G C
 190 200 210
 ASN ASN THR ASP ASN SER THR SER THR THR
 A A C A A C A C G G A T A A C A G C C A C C A G C A C A A C T
 220 230 240

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ASP PRO ASN GLY ASP ASN ASN GLN LEU THR
 G A C C C A A A T G G C G A T A A C A A C C A A C T G A C A
 250 260 270
 GLN ALA GLN LYS THR ALA ALA ALA ALA GLY
 C A A G C A C A A A A A A C T G C C C G C C G C C A G G G
 280 290 300

PHE PHE VAL MET GLY LYS ILE ARG ASP THR
 T T T T T G T G A T G G G T A A A A T T C G T G A T A C C
 310 320 330
 SER GLU LYS ASN ASP PRO ASP TYR THR LYS
 A G C G A A A A A A A T G A C C C A G A T T A T A C C A A A
 340 350 360

FIG.16C

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```

ASP LEU GLN GLY SER VAL HIS THR ALA GLY
G A T T A C A A G G C A G C G T A C A T A C A G C A G G G
370                               380
GLN GLY LEU GLN TYR LEU GLY THR LYS GLU
C A A G G C T T A C A G T A C T T A G G C A C C A A A G A G
400                               410
420

PRO ARG PRO ASP GLY THR GLY THR GLY LYS
C C T C G G C C A G A T G G C A C A G G T A C A G G T A A A
430                               440
ASN LEU ARG GLN PRO ILE THR ALA ASP ASP
A A C T T A C G C C A G C C C A T C A C A G C T G A T G A C
460                               470
480

ILE THR PRO PRO LEU TYR PHE ASP LYS PHE PRO
A T T A C A C C A C T T T A T T T G A T A A A T T C C C C
490                               500
LYS ILE SER ASP LEU HIS LEU GLU ALS SER
A A A A T A T C C G A T C T G C A C C T A G A A A C A G C
520                               530
540

```

FIG.16D

GLU HIS VAL PHE ASP ALA LYS LYS ALA ASN
 G A G C A T G T G T T T G A T G C C T A A A A A A G C A A A T
 550 560 570
 ASN ILE LYS ILE TYR GLY TYR SLY ALA LEU
 A A C A T C A A A A T A T A T G G T T A T G G T G C A T T G
 580 590 600

SER SER PRO ALA LYS ASN PRO THR TYR MET
 T C A T C A C C T G C C A A A A C C C A C C T A C A T G
 610 620 630
 ASN TYR GLN GLN GLU GLN ASN ILE LYS ASN
 A A T T A T C A A C A A G A A C A A A C A T C A A A A C
 640 650 660

LYS LYS PRO GLY ASP ASP TYR GLN ASN ILE
 A A A A C C A G G C G A T G A T T A T C A A A C A T T
 670 680 690
 ARG PHE GLY TYR MET GLU LEU ARG GLU LEU
 C G T T T G G C T A T A T G G A G C T A A G A G A G C T G
 700 710 720

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FIG.16E

ASP LEU ASN LYS LYS GLY ALA ASP THR GLN
 G A C C T A A A T A A A A G G T G C A G A C A C C C A G
 730 740 750
 SER ASP LYS ASN ARG ALA ILE PHE THR
 A G C G A C A A G A A C C G T G C C A T C A T T T C A C C
 760 770 780

THR PRO THR LEU PHE TYR HIS GLY GLN ASN
 A C A C C T A C T T A T T T A T C A T G G T G A G A A T
 790 800 810
 ALA SER THR HIS LEU PRO LYS ALA GLY LYS
 G C C A G C A C C C A T C T G C C C A A A G G C G G T A A A
 820 830 840

PHE THR ASP ASP LYS VAL GLY THR TYR PHE
 T T T G A C T A T G A G G G C A A T T G G T T G T A T C T G
 850 860 870
 ASN SER THR ARG LYS SER ASN GLU GLY ASP
 A C C G A T G T C A A A A A C G C C C A T T T T A G A T
 880 890 900

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FIG.16F

LYS THR ASP ASP LYS VAL GLY THR TYR PHE
 A A A C A G A C G A T A A G T A G G C A C T T A T T T
 910 920
 ASN SER THR ARG LYS SER ASN GLU GLY ASP
 A A C T C A A C C A G A A A A T C A A A T G A A G G C G A T
 940 950 960

LEU VAL SER ALA ALA HIS ILE TYR LEU ASN
 T T G G T G A G T G C A G C A C A T T T A T C T A A A C
 970 980 990
 SER PHE LYS TYR LYS HIS THR PRO ALA THR
 A G C T T T A A A T A T A A C A C C C C G G C C A C T
 1000 1010 1020

TYR SER VAL ASP PHE ASP GLN ASN THR LEU
 T A T A G C G T G G A C T T T G A T C A A A A T A C C C T A
 1030 1040 1050
 LYS GLY LYS LEU SER TYR TYR ASP ASN PRO
 A A A G G C A A A T T G T C T T A T T A T G A C A C C C A
 1060 1070 1080

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FIG. 16G

ASN LYS GLN THR ALA ASP GLY ARG TYR ILE
 A A C A G C A A A C A G C C G A T G G G C G T T A T A T C
 1090 1100
 ARG SER GLN PHE ASP THR ASP LYS VAL
 A G A A G T C A G T T T G A T A C C G A C A A A A G G T C
 1120 1130 1140

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 ASN GLU ALA ASP VAL TYR GLU ILE ASP ALA
 A A T G A A G C C G A T G T C T A T G A G A T T G A C G C C
 1150 1160 1170
 LYS ILE ASN GLY ASN ARG PHE THR GLY THR
 A A G A T T A A T G G C A A C C G C T T T A C T G G C A C A
 1180 1190 1200

ALA LYS SER LEU ILE ASP ASP ASN THR ASN
 G C C A A A T C T T T G A T T G A T G A T A C A C C A A T
 1210 1220 1230
 THR ALA PRO PHE VAL LYS GLU LEU PHE SER
 A C C G C A C C T T T G T T A A A G A G C T G T T C T C C
 1240 1250 1260

FIG. 16H

```

LYS  LYS  ALA  ASN  PRO  ASN  PRO  ASP  PRO
A A A A G C C A A T C C C A A C C C A G A C C C
1270                                     1290
      ASN  SER  ASP  THR  LEU  GLU  GLY  GLY  PHE  TYR
      A A C T C A G A T A C G C T A G A A G G C G G G T T T A T
      1300                                     1310 1320
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GLY  GLU  SER  GLY  ASP  GLU  LEU  ALA  GLY  LYS
G G T G A G T C G G G C G A T G A G C T G G C G G G T A A A
1330                                     1340 1350
      PHE  LEU  SER  ASN  ASP  ASN  ALA  THR  PHE  VAL
      T T T T T A T C C A A T G A C A A C G C A A C T T T G T G
      1360                                     1370 1380
VAL  PHE  GLY  GLY  LYS  ARG  ASP  LYS  THR  THR
G T C T T T G G T G G C A A A C G A G A C A A A C G A C C
1390                                     1400 1410
      GLU  PRO  VAL  ALA  THR  LYS  THR  VAL  THR  PHE
      G A C C T G T C G C C A C A A A A C G G T G T A T T T
      1420                                     1430 1440

```

FIG.16I

SER THR GLY PHE GLU LYS POE SER THR SER
 AGTACAGGATTTGAA A A A C C C A G C A C C A G C
 1450 1460 1470
 PHE VAL GLY ASN GLU GLU ILE GLY SER ILE
 TTTGTTGGCAATG A A G A G A T T G G T A G C A T T
 1480 1490 1500

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 ILE ASP GLY LYS LYS LEU ASN ASP GLU VAL
 ATGACGGTTAA A A G T T A A T G A T G A A G T C
 1510 1520 1530
 ASN ASN GLN ILE GLU ASP GLU THR VAL PRO
 AATAATCA A A T T G A A G A T G A A C T G T C C C T
 1540 1550 1560

VAL SER ASN LYS LYS GLU TYR TYR GLU TYR ASN
 GTCAGTAATAA A A G A A T A T T A T G A A T A T A A T
 1570 1580 1590
 TYR GLY ARG PRO ASN LYS GLN PHE THR LYS
 TATGGACGACCC A A C A A A C A A T T C A C C A A A
 1600 1610 1620

FIG. 16J

LYS ILE ASN ALA SER VAL GLN LYS ASN PRO
 A A A T A A A C G C C A G C G T C C A A A A A A C C C T
 1630 1640 1650
 ALA TYR PHE GLY GLN HIS ASP LYS PHE TYR
 G C T T A T T T G G T C A G C A T G A T A A G T T T T A T
 1660 1670 1680

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PHE ASN GLY ASN TYR TYR ASP LEU SER ALA
 T T T A A T G G T A A C T A T T A T G A C T T A T C A G C C
 1690 1700 1710
 LYS GLU ALA ASN LYS LEU GLY VAL THR ASP
 A A G A A G C A A A C A A G C T T G G T G T C T C C C A A
 1720 1730 1740

ASP THR SER THR ASN LYS SER ILE LEU ALA
 G A T A C C A G C A C C A A T A A G A G T A T T T T G G C T
 1750 1760 1770
 LYS TYR PRO ASP ALA LYS VAL SER THR ASP
 A A A T A C C C A G A T G C C A A A G T A A G C A C A G A C
 1780 1790 1800

FIG.16K

ASN LYS VAL THR LYS ILE VAL LEU GIN
 AATAAGTTACCAAAATCGTTCTACAACA
 1810 1820 1830
 ALA LYS ASP LYS PRO TYR THR ALA ILE HIS
 GCCAAGATAAGCCGTATACCGCCATTCA
 1840 1850 1860

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 ALA LYS SER TYR ASP HIS ILE SER PHE GLY
 GCCAAAGCTATGACCAACATCAGTTTGGT
 1870 1880 1890
 GLU VAL LEU TYR ASN ASP ASN LYS GLY ASN
 GAAGTATTGTATAATGATAACAAGGCAAC
 1900 1910 1920

PRO THR ARG SER TYR PHE VAL GIN GLY GLY
 CCAACAGCAGTTATTTGTGCAAGGCGGT
 1930 1940 1950
 GIN ALA ASP VAL SER THR GIN LEU PRO SER
 CAGCGGATGTCAGTACTCAGCTGCCAGT
 1960 1970 1980

FIG. 16L

ALA GLY LYS PHE THR TYR ASN GLY LEU TRP
 GCAGGTAAATTCACCTATAATGGTCTTTGG
 1990 2000 2010
 ALA GLY TYR LEU THR GLN LYS LYS ASP LYS
 GCAGGCTACCTGACCCAGAAAGACAA
 2020 2030 2040

GLY TYR SER LYS ASP GLU ASP THR ILE LYS
 GGTATAGCAAGATGAGGATACCATCAAG
 2050 2060 2070
 GLN LYS GLY LEU LYS ASP TYR ILE LEU THR
 CAAAGGCTCTTAAAGATTATATTGACC
 2080 2090 2100

LYS ASP PHE ILE PRO GLN ASP ASP ASP
 AAGACTTTATCCCAAGATGACGATGAC
 2110 2120 2130
 ASP ASP ASP SER LEU THR ALA SER ASP ASP
 GATGACGATAGTTTGACCGCATCTGATGAT
 2140 2150 2160

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FIG. 16M

SER GLN ASP ASP ASN THR HIS GLY ASP ASP
 TCACAGATGATAATACACATGGCGATGAT
 2170 2180 2190
 ASP LEU ILE ALA SER ASP ASP SER GLN ASP
 GATTGATTGCACTCTGATGATTCAACAAGAT
 2200 2210 2220

ASP ASP THR ASP GLY ASP ASP ASP SER ASP
 GATGACACAGATGGCGATGACGATTCAAGAT
 2230 2240 2250
 ASP LEU GLY ASP GLY ALA ASP ASP ASP ALS
 GATTGGGTGATGGTGCAAGATGATGACGCC
 2260 2270 2280

ALA GLY LYS VAL TYR HIS ALA GLY ASN ILE
 GCAGGC AAAGTGTAATCATGCAAGGTAATAT
 2290 2300 2310
 ARG PRO GLU PHE GLU ASN LYS TYR LEU PRO
 CGCCCTGAATTGAAACAACAATACTTGCCC
 2320 2330 2340

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FIG. 16N

ILE ASN GLU PRO THR HIS GLU LYS THR PHE
 A T T A A T G A G C C T A C T C A T G A A A A A C C T T T
 2350 2360 2370
 ALA LEU ASP GLY LYS ASN LYS ALA LYS PHE
 G C C C T A G A T G G T A A A A T A A G G C T A A G T T T
 2380 2390 2400

ASP VAL ASN PHE ASP THR ASN SER LEU THR
 G A T G T A A A C T T T G A C A C C A C A G C C T A A C T
 2410 2420 2430
 GLY LYS LEU ASN ASP GLU ARG GLY ASP ILE
 G G T A A A T T A A C G A T G A G A G A G G T G A T A T C
 2440 2450 2460

VAL PHE ASP ILE LYS ASN GLY LYS ILE ASP
 G T C T T T G A T A T C A A A A T G G C A A A A T T G A T
 2470 2480 2490
 GLY THR GLY PHE THR ALA LYS ALA ASP VAL
 G G C A C A G G A T T T A C C G C C A A A G C C G A T G T G
 2500 2510 2520

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FIG. 16O

PRO ASN TYR ARG GLU GLU VAL GLY ASN ASN
 CCA AACTATCGTGAGAGAGTGGGTAAACAAC
 2530 2540 2550
 GLN GLY GLY PHE LEU TYR ASN ILE LYS
 CAGGTGGCGGTTTCTTATACAAATCAAA
 2560 2570 2580

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ASP ILE ASP VAL LYS GLY GLN PHE PHE GLY
 GATATTGATGTTAAGGGGCAATTTTTGGC
 2590 2600 2610
 THR ASN GLY GLU LEU ALA GLY ARG LEU
 ACAATGGCGAAGAGTTGGCAGGACGGTTA
 2620 2630 2640

HIS HIS ASP LYS GLY ASP GLY ILE THR ASP
 CATCATGACAAAGGCGATGGCATCACTGAC
 2650 2660 2670
 THR ALA GLU LYS ALA GLY ALA VAL PHE GLY
 ACCGCCGAAAGCAGGGGCTGCTTTGGG
 2680 2700 2710

ALA VAL LYS ASP LYS ***
 GCTGTTAAAGATAATAA
 2710

Partial restriction map of *M. catarrhalis* strain VH19 *lbpB*

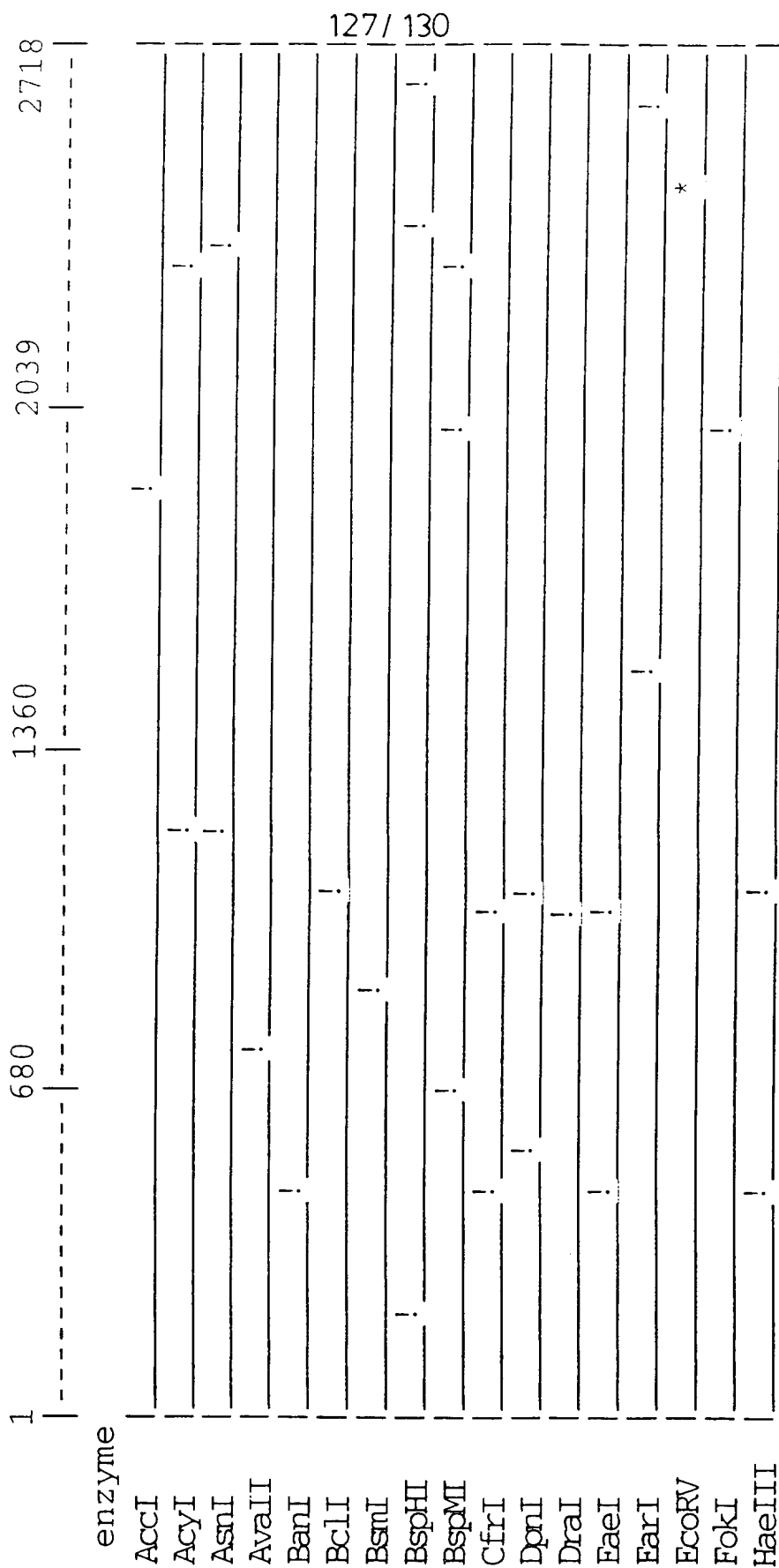


FIG.17A

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HindIII	!
HpaII	!
MaeI	!
MboI	!
MnlI	!
NciI	!
NspBII	!
NspI	!
PaiI	!
PvuII	!
ScaI	!
SspI	!
StyI	!
XbaI	!

FIG.17B

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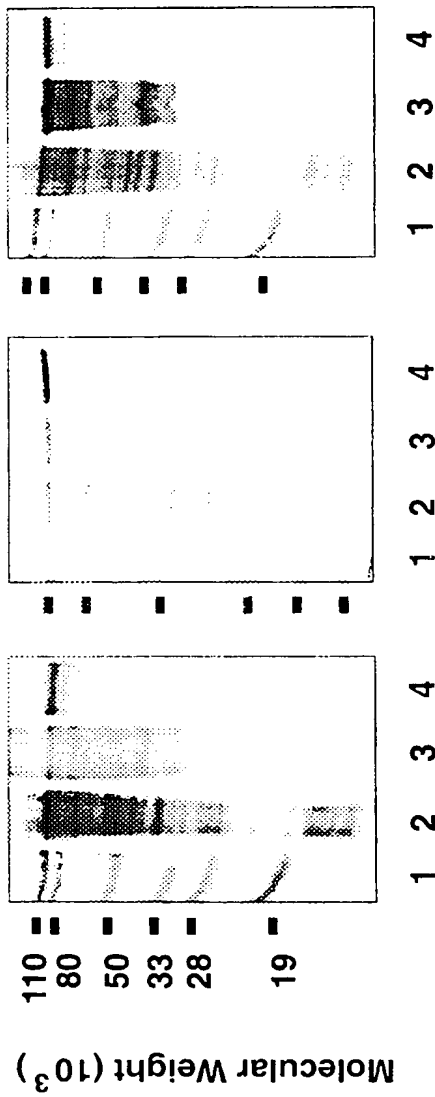


FIG.18A.

FIG.18B.

FIG.18C.

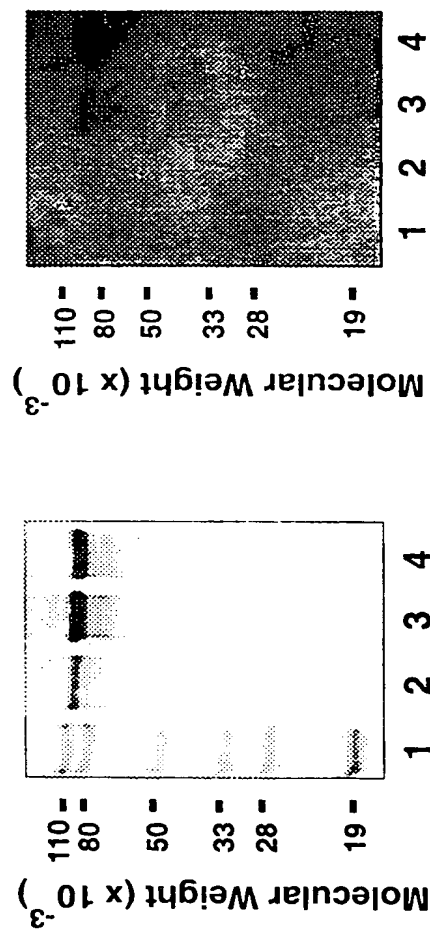


FIG.19A.

FIG.19B.

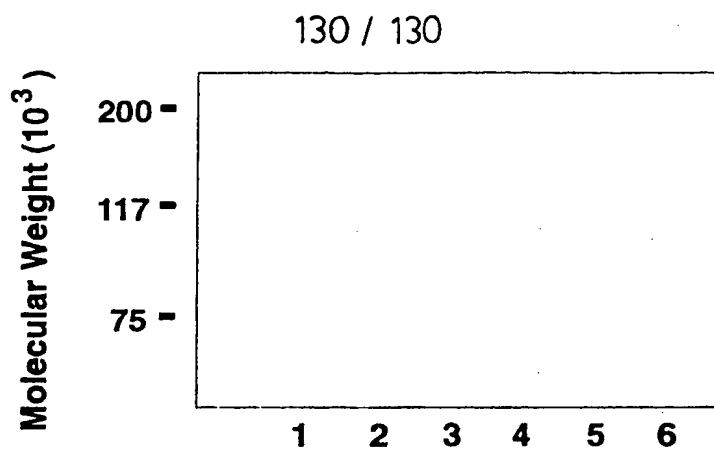


FIG.20A.

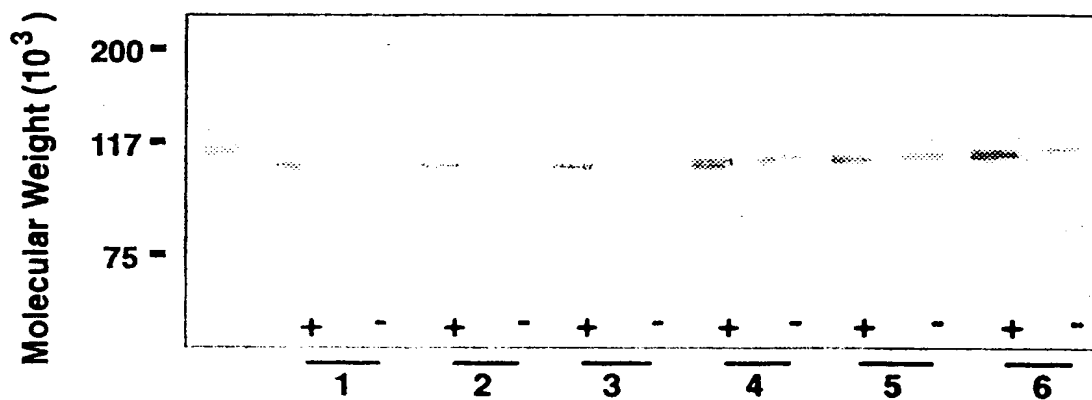


FIG.20B.

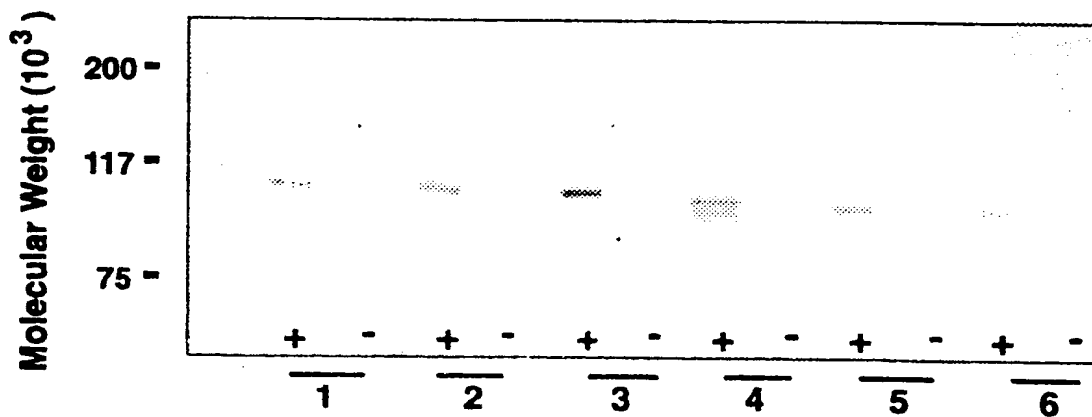


FIG.20C.



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/12, C07K 14/79, 14/22, A61K 38/17, G01N 33/68, C12Q 1/68	A3	(11) International Publication Number: WO 98/55606 (43) International Publication Date: 10 December 1998 (10.12.98)
(21) International Application Number: PCT/CA98/00544 (22) International Filing Date: 2 June 1998 (02.06.98) (30) Priority Data: 08/867,941 3 June 1997 (03.06.97) US 09/074,658 8 May 1998 (08.05.98) US (71) Applicant (for all designated States except US): CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Av- enue West, North York, Ontario M2R 3T4 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): LOOSMORE, Sheena, M. [CA/CA]; 70 Crawford Rose Drive, Aurora, Ontario L4G 4R4 (CA). DU, Run-Pan [CA/CA]; (CA). WANG, Quijun [CA/CA]; 299 Chelwood Drive, Thornhill, Ontario L4J 7Y8 (CA). YANG, Yan-Ping [CA/CA]; Apartment 709, 120 Torresdale Avenue, Willowdale, Ontario M2R 3N7 (CA). KLEIN, Michel, H. [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA). (74) Agent: STEWART, Michael, I.; Sim & McBurney, 6th floor, 330 University Avenue, Toronto, Ontario M5G 1R7 (CA).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> (88) Date of publication of the international search report: 4 March 1999 (04.03.99)
(54) Title: LACTOFERRIN RECEPTOR GENES OF MORAXELLA (57) Abstract Purified and isolated nucleic acid molecules are provided which encode lactoferrin receptor proteins of <i>Moraxella</i> , such as <i>M. catarrhalis</i> , or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid sequence may be used to produce recombinant lactoferrin receptor proteins Lbp1, Lbp2 or ORF3 of the strain of <i>Moraxella</i> free of other proteins of the <i>Moraxella</i> strain for purposes of diagnostics and medical treatment. Furthermore, the nucleic acid molecule may be used in the diagnosis of infection.		

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INTERNATIONAL SEARCH REPORT

Inter. Application No
PCT/CA 98/00544

A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/12 C07K14/79 C07K14/22 A61K38/17 G01N33/68 C12Q1/68		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 6 C07K C12N A61K G01N C12Q		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CA 2 162 193 A (CONNAUGHT LAB) 3 May 1997	20-22, 24,29,30
Y	see the claims see abstract; figures 1-3; examples 1,4 see page 4, line 21 - page 7, line 15	3,4, 6-19,23, 25-28
X	OGUNNARIWO J A AND SCHRYVERS A B: "Rapid identification of bacterial transferrin and lactoferrin receptor protein genes" JOURNAL OF BACTERIOLOGY, vol. 178, no. 24, December 1996, pages 7326-7328, XP002083300	1,2,5, 31,32
Y	see abstract; figure 1; table 1 see page 7327 - page 7328 <div style="text-align: center;">---</div> <div style="text-align: center;">-/--</div>	3,4, 6-19,23, 25-28
<div style="display: flex; justify-content: space-between;"> <input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex. </div>		
* Special categories of cited documents : <div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed </div> <div style="width: 45%;"> "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "&" document member of the same patent family </div> </div>		
Date of the actual completion of the international search <div style="text-align: center; font-weight: bold;">5 November 1998</div>		Date of mailing of the international search report <div style="text-align: center; font-weight: bold;">23/12/1998</div>
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016		Authorized officer <div style="text-align: center; font-weight: bold;">Oderwald, H</div>

INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 98/00544

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 90 12591 A (UNIV TECHNOLOGIES INT ;SCHRYVERS ANTHONY BERNARD (CA)) 1 November 1990 see abstract; claims 5-8,16-26; figure 1; examples 1,2 see page 5, line 4 - page 6, line 26 see page 7, line 30 - page 8, line 1 see page 10, line 8 - page 11, line 8 -----	20,24,29
X	SCHRYVERS A B ET AL: "COMPARATIVE ANALYSIS OF THE TRANSFERRIN AND LACTOFERRIN BINDING PROTEINS IN THE FAMILY NEISSERIACEAE" CANADIAN JOURNAL OF MICROBIOLOGY, vol. 35, no. 5, May 1989, pages 409-415, XP002020995 cited in the application see abstract; figures 1,4-6 see page 410, paragraph 9 see page 414, paragraph 2 -----	20-22,24
T	DU R-P ET AL: "Cloning and expression of the Moraxella catarrhalis lactoferrin receptor genes" INFECTION AND IMMUNITY, vol. 66, no. 8, August 1998, pages 3656-3665, XP002083301 see the whole document -----	1-32

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA 98/ 00544

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claim(s) 30
is(are) directed to a method of treatment of the human/animal
body, the search has been carried out and based on the alleged
effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such
an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please see additional sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all
searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment
of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report
covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is
restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1, 5-20, 24, 29-32 all partially and 2, 21.

Lbp1 polypeptide, corresponding nucleic acids, vectors containing said nucleic acid, host cells transformed with said vector, immunogenic composition containing said nucleic acid or protein, methods for detection and diagnostic kit involving said nucleic acid.

2. Claims: 1, 5-20, 24, 29-32 all partially and 3, 22.

Same as in invention 1 but comprising Lbp2.

3. Claims: 1, 5-20, 24, 29-32 all partially and 4, 23, 25-28.

Same as in invention 1 but comprising ORF3.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/CA 98/00544

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
CA 2162193 A	03-05-1997	NONE	
WO 9012591 A	01-11-1990	US 5292869 A	08-03-1994
		AU 649950 B	09-06-1994
		AU 5526190 A	16-11-1990
		EP 0528787 A	03-03-1993
		JP 4506794 T	26-11-1992
		NZ 247967 A	24-02-1995
		US 5141743 A	25-08-1992

